

UNIVERSIDADE ESTADUAL DE MARINGÁ  
CENTRO DE CIÊNCIAS BIOLÓGICAS  
DEPARTAMENTO DE BIOLOGIA  
PROGRAMA DE PÓS-GRADUAÇÃO EM ECOLOGIA DE  
AMBIENTES AQUÁTICOS CONTINENTAIS

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**Efeitos da altitude, longitude e latitude na composição histórica e ecológica dos anfíbios da Floresta Atlântica brasileira**

Maringá  
2016

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Tese apresentada ao Programa de Pós-Graduação em Ecologia de Ambientes Aquáticos Continentais do Departamento de Biologia, Centro de Ciências Biológicas da Universidade Estadual de Maringá, como requisito parcial para obtenção do título de Doutor em Ciências Ambientais.

Área de concentração: Ciências Ambientais

Orientador: Prof. Dr. Rogério Pereira Bastos

Maringá  
2016

"Dados Internacionais de Catalogação-na-Publicação (CIP)"  
(Biblioteca Setorial - UEM. Nupélia, Maringá, PR, Brasil)

M827e Moraes, Ricardo Lourenço de, 1976-  
Efeitos da altitude, longitude e latitude na composição histórica e ecológica dos anfíbios da Floresta Atlântica brasileira / Ricardo Lourenço de Moraes.-- Maringá, 2016.  
330 f. : il. (algumas color.).

Tese (doutorado em Ecologia de Ambientes Aquáticos Continentais)--Universidade Estadual de Maringá, Dep. de Biologia, 2016.  
Orientador: Prof. Dr. Rogério Pereira Bastos.

1. Anfíbios - Ecologia - Distribuição - Floresta Atlântica - Brasil. 2. Anfíbios - Ecologia - Modelagem - Floresta Atlântica - Brasil. 3. Anfíbios - Evolução - Floresta Atlântica - Brasil. I. Universidade Estadual de Maringá. Departamento de Biologia. Programa de Pós-Graduação em Ecologia de Ambientes Aquáticos Continentais.

CDD 23. ed. -597.8170981  
NBR/CIP - 12899 AACR/2

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Aprovado em: 08 de Agosto de 2016.

Local de defesa: Anfiteatro Prof. “Keshiyu Nakatani”, Nupélia, Bloco G-90, *campus* da Universidade Estadual de Maringá.

*“Dedico este trabalho a  
minha mãe que sempre  
me apoiou, e agora lá  
de cima, me abençoa.”*

## AGRADECIMENTOS

A minha esposa Meline por todo apoio, ajuda e parceria em todas as etapas de minha carreira científica;

Aos filhos Lia, Ísis e Uriel por entenderem e permitirem que eu escrevesse este trabalho;

Ao professor Dr. Rogério Pereira Bastos pela orientação, amizade, auxílio em todas as etapas deste trabalho e, principalmente, por ter acreditado no meu potencial;

As queridas e competentes Aldenir e Jucemara, Salete e João, e a todos os professores do PEA;

Aos grandes parceiros e amigos Felipe Campos, Rodrigo Ferreira, Rafael Rosa, Vinicius Batista, Leo Malagoli, Fernando Lansak-Toha, Leilaine Talita, Helen Cassia, Mileny Otani, Amanda Fernandes, Claudio Zawadzki, Gabriel Deprá, Marion Haruki, Dayani Bailly, Iuri Dias, Mirco Solé, Tiago Rangel, Amanda Covre, Renan Oliveira, Guilherme Figueiredo, Caio Carniatto, Evanilde Benedito, Weferson Graça pelas importantes contribuições nesse estudo;

Ao Nupélia e ao PEA pelo fornecimento da infra-estrutura e desenvolvimento da pesquisa;

Ao CNPq pelo apoio financeiro;

Aos meus pais, por todo amor e dedicação;

Muito obrigado!

# **Efeitos da altitude, longitude e latitude na composição histórica e ecológica dos anfíbios da Floresta Atlântica brasileira**

## **RESUMO**

Identificar quais os fatores bióticos ou abióticos que influenciam na riqueza e dispersão das espécies, são importantes ferramentas para se entender os processos ecológicos para a manutenção das espécies. A Floresta Atlântica foi altamente devastada restando poucos fragmentos da suas florestas originais. Anfíbios são considerados o grupo de vertebrados mais ameaçados e cerca de 90% das espécies que ocorrem na Floresta Atlântica brasileira, são endêmicos deste bioma. Nos avaliamos as espécies de anfíbios que ocorrem na Floresta Atlântica quanto a sua capacidade de dispersão e adaptação. Buscou-se entender os processos abióticos que influenciam a riqueza e endemismo de espécies bem como as características funcionais e filogenéticas que influenciam na distribuição das espécies. Avaliamos refúgios ecossistêmicos e evolutivos durante o presente e futuro, auxiliando estratégias de conservação no futuro. Traçamos a história evolutiva dos mecanismos antipredação bem como a sua importância na distribuição das espécies. Os resultados mostraram que espécies mais adaptadas as alterações antrópicas possuem maior distribuição geográfica. A temperatura, topografia e áreas alteradas pelo homem são uma importante barreira geográfica. Áreas de altitudes serão os novos refúgios durante a era Antropocênica e repetem as mesmas áreas que foram usadas como refugio durante a era Pleistocênica. Mecanismos antipredação podem ter origem puramente filogenética, sendo que quando isto acontece, podem determinar novos ou ajustes de mecanismos existentes, e espécies com maior diversidade de mecanismos também possuem maior capacidade de dispersão.

**Palavras-chave:** Ectotermicos. Características funcionais. Filogenia. Evolução. Dispersão.

# **Effects of altitude, longitude and latitude in the historical and ecological composition of amphibians in the Brazilian Atlantic forest**

## ***ABSTRACT***

Identify the biotic or abiotic factors that influence species richness and dispersion, are important tools for understanding the ecological processes to maintain of the species. The Atlantic Forest was highly devastated leaving few fragments of its original forests. Amphibians are considered the most threatened group of vertebrates and about 90% of the species that occur in the Brazilian Atlantic Forest, are endemic to this biome. We evaluate amphibian species that occur in the Atlantic Forest as its ability to dispersion and adaptation. We attempted to understand the abiotic processes that influence the richness and endemism of species as well as the functional traits and phylogenetics features that influence the distribution of species. We evaluate ecosystem and evolutionary refuges for the present and future, helping conservation strategies in the future. Traced the evolutionary history of antipredator mechanisms and their importance in the distribution of species. The results showed that more adapted species to anthropogenic changes have greater geographic distribution. The temperature, topography and areas altered by humans are an important geographical barrier. Altitude areas will be the new refuges during the Antropocene age and repeat the same areas that were used as refuge during the Pleistocene age. Antipredator mechanisms may have origin purely phylogenetic may determine new or adjustments of existing mechanisms, species with the greatest diversity of mechanisms also have greater ability to dispersion.

***Keywords:*** Ectothermic. Functional traits. Phylogeny. Evolution. Dispersion.



Tese elaborada e formatada conforme as normas das publicações científicas  
*Diversity and Distributions*. Disponível em:  
<[http://onlinelibrary.wiley.com/journal/10.1111/\(ISSN\)1472-4642/homepage/ForAuthors.html](http://onlinelibrary.wiley.com/journal/10.1111/(ISSN)1472-4642/homepage/ForAuthors.html)>,  
*Global Change Biology*. Disponível em:  
<[http://onlinelibrary.wiley.com/journal/10.1111/\(ISSN\)1365-2486/homepage/ForAuthors.html](http://onlinelibrary.wiley.com/journal/10.1111/(ISSN)1365-2486/homepage/ForAuthors.html)>  
e *Evolution*. Disponível em:  
<[http://onlinelibrary.wiley.com/journal/10.1111/\(ISSN\)1558-5646/homepage/ForAuthors.html](http://onlinelibrary.wiley.com/journal/10.1111/(ISSN)1558-5646/homepage/ForAuthors.html)>

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## 1 INTRODUÇÃO GERAL

A busca pela compreensão dos fatores ambientais associados à riqueza de espécies em uma determinada área, tal qual às variações ocorrentes no espaço e tempo, tem sido um dos principais temas de pesquisa dos ecólogos (BEGON et al. 2007). Para tentar entender os fatores que influenciam a diversidade de espécies em larga escala, foi proposto por Brown & Maurer (1989) o estudo da *Macroecologia*, sendo os padrões geográficos (latitude e altitude), a característica funcional “tamanho do corpo” um dos focos principais. A variação de fatores abióticos pode influenciar a composição das comunidades ao longo de gradientes espaciais, como em relação à latitude (i.e., os fatores climáticos relacionados a ela), onde a riqueza de espécies é inversamente proporcional à latitude (PIANKA 1966; GASTON 2000). Alternativamente, a altitude (i.e., os fatores abióticos relacionados a ela) também influencia na riqueza de espécies, sendo menor nas altitudes maiores (MACARTHUR 1972; STEVENS 1992; RAHBK 1995). Contudo, em áreas de maior altitude pode ocorrer maior número de espécies sensíveis e endêmicas, como ocorre com os anfíbios devido as temperaturas mais amenas e alta umidade (CRUZ & FEIO 2007).

Os anfíbios estão sofrendo um acentuado declínio (STEININGER et al. 2001; LIPS & DONNELLY 2002; HAYES et al. 2010; BECKER & ZAMUDIO 2011). Os principais fatores deste declínio seriam a fragmentação e a perda do hábitat, levando as pequenas populações à redução da diversidade genética, aumentando o risco de deriva genética e endogamia (YOUNG et al. 1996; BECKER et al. 2007; DIXO et al. 2009; LOURENÇO-DE-MORAES et al. 2012). Estes fatores tornam as populações mais suscetíveis aos efeitos estocásticos devido ao

menor potencial evolutivo (HOLT et al. 2004). Além disso, outros fatores estão associados ao declínio de anfíbios, como o aumento da radiação ultravioleta-B (POUNDS et al. 2006; BLAUSTEIN & JOHNSON 2003), doenças letais causadas por fungos (LIPS et al. 2003), tráfico ilegal de animais (PISTONI & TOLEDO 2010), alterações da legislação ambiental (TOLEDO et al. 2010) e introdução de espécies exóticas (KATS & FERRER 2003).

A Floresta Atlântica originalmente se estendia desde a costa do Rio Grande do Norte até o Rio Grande do Sul, e devido à excessiva pressão antrópica, hoje restam alguns últimos remanescentes com um total aproximado de 98.000 Km<sup>2</sup> (MORELLATO & HADDAD 2000), representando cerca de 16% da cobertura original (RIBEIRO et al. 2009). Com uma grande concentração de espécies endêmicas de plantas e animais, é considerada um dos trinta e cinco *hot spots* mundiais mais importantes para biodiversidade (MYERS et al. 2000). A Floresta Atlântica possui um total estimado de mais de 1.800 espécies de vertebrados terrestres, sendo que aproximadamente 36% destas espécies são endêmicas deste bioma (CI 2005). Cerca de 90% das espécies de anfíbios descritas para o Brasil são endêmicos da Floresta Atlântica (HADDAD et al. 2013). Um fator que pode ser relacionado à alta riqueza deste bioma é o grande número de *habitat* e de micro-habitat, favorecendo o número de espécies hábitat-especialistas (HADDAD 1998). Além disso, é provável que exista um alto grau de isolamento entre as populações, dado pelo relevo altamente acidentado com rios e montanhas que podem representar barreiras biogeográficas, associado as suas características fisiológicas e pressões bióticas como predação (HADDAD 1998; MARQUES et al. 1998; WELLS 2007).

Muitos anfíbios que vivem restritos nas proximidades ou nos topos de morros estão sofrendo um forte impacto sob um cenário de aquecimento climático ou micro-climático, porque as condições ambientais adequadas para sua sobrevivência, podem não existir mais localmente e algumas espécies desapareceram (WAKE 2012). Estudos com anuros apontam menor riqueza em maiores altitudes (SCOTH Jr. 1976; FAUTH et al. 1989; NAVAS 2002; 2006). No Brasil os estudos testando os efeitos da altitude em comunidades são escassos, apontando redução da riqueza com o aumento da altitude (GIARETTA et al. 1997; 1999). Contudo, ainda existe uma carência de conhecimento taxonômico sobre as espécies de altitude, as quais estão sendo descritas recentemente (e.g. CRUZ et al. 2007; ALVES et al. 2009; RECODER et al. 2010; NAPOLI et al. 2011; GAREY et al. 2012; LOURENÇO-DE-MORAES et al. 2014).

Investigamos em macroescala a atual situação dos anfíbios da Floresta Atlântica brasileira. Para isso investigou-se o padrão de distribuição das espécies e relacionou-os com a topografia, referências espaciais e variáveis ambientais e analisou quais características funcionais auxiliam na dispersão das espécies promovendo o atual padrão de diversidade beta em toda a Floresta Atlântica brasileira; os atuais *hotspots* de diversidade funcional e filogenética, identificando os refúgios climáticos das espécies baseado nos padrões ecossistêmicos e evolutivos para o futuro; e a origem dos mecanismos antipredação e verificou as características funcionais que influenciam na diversidade de mecanismos antipredação bem como a diversidade destes mecanismos e sua relação com a distribuição das espécies.

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## 2 FUNCTIONAL TRAITS EXPLAIN THE CURRENT DISTRIBUTION OF THE AMPHIBIAN SPECIES IN BRAZILIAN ATLANTIC FOREST

### *Abstract*

Dispersion processes of species of the most important ways to understand how communities interact in macroecological studies. The functional abilities of the species, such as its plasticity throughout of the various environments in a macroscale, can be decisive for the dispersion process and may reflect the beta diversity of a locality or region. In this study we assessed the potential dispersion of amphibian species based on their functional traits, we analyze the influence of abiotic factors on species richness, endemism, and beta diversity pattern. Our results show that the current change of forest affect direct the beta diversity pattern because the most species adapted to this new environment altered by anthropic actions to increase their range. Species with functional characteristics such as body size (medium and large) and the habitat (open area and open/forest areas) are the main traits that explain the current pattern of beta diversity. The richness pattern is determined to historical processes in synergy with abiotic variables and species with more suitable phenotypic specializations to anthropogenic changes are the least threatened in the current geological Age.

**Key words:** Anthropocene; beta diversity; ectothermics; Anura; Gymnophiona

## CARACTERÍSTICAS FUNCIONAIS EXPLICAM A ATUAL DISTRIBUIÇÃO DAS ESPÉCIES DE ANFÍBIOS NA FLORESTA ATLÂNTICA BRASILEIRA

### **Resumo**

Os processos de dispersão das espécies são uma das formas mais importantes para compreender como as comunidades interagem em estudos macroecológicos. As habilidades funcionais das espécies, bem como sua plasticidade ao longo dos vários ambientes em uma macroescala, pode ser decisivo para o processo de dispersão e pode refletir a diversidade beta de uma localidade ou região. Neste estudo avaliou-se a dispersão potencial das espécies de anfíbios com base em suas características funcionais, analisamos a influência de fatores abióticos sobre a riqueza de espécies, endemismo e o padrão de diversidade beta. Os resultados mostram que a alteração original das florestas afeta diretamente o padrão de diversidade beta, isso devido a maioria das espécies adaptadas a este novo ambiente alterado pelas ações antropicas, aumentam a sua área de distribuição. Espécies com características funcionais, tais como o tamanho do corpo (médio e grande) e *habitat* (área aberta e áreas abertas/florestal) são as principais características que explicam o atual padrão de diversidade beta. O padrão de riqueza está determinado a processos históricos em sinergia com variáveis abióticas, e espécies com especializações fenotípicas mais adequadas as alterações antropicas são as menos ameaçadas na Era geológica atual.

**Key words:** Antropoceno; diversidade beta; ectotérmicos; Anura; Gymnofiona

## 2.1 INTRODUCTION

Understanding the processes of dispersion, distribution and permanence of species are likely the most studied topics of macroecologists and biogeographers. Among the processes that can address these patterns, two theories are much discussed: the niche theory (Hutchinson, 1957), where biotic and abiotic factors determine the composition patterns and distribution of species, and the neutral theory (Hugbell, 2001), where simple random stochastic processes are responsible for the distribution, extinction and dispersal of species. It is recognized that the current pattern of dispersion and distribution of species is related to a combination of historical and contemporary events (Ricklefs, 1987; Oberdorff, 1997; Svenning & Skov, 2007; Carnaval & Moritz, 2008; Carnaval *et al.*, 2009; Baselga *et al.*, 2012; Silva *et al.*, 2014). The dispersion of a species is often related by the interactions of characteristics such as body size, local abundance, and probability of extinction with geographical range size (Brown & Maurer, 1989; Gaston, 1990; Lawton, 1993). Therefore, one of the issues to understand is how communities are organized and distributed and their ecological and evolutionary questions (Gaston, 1991).

Endothermic species with restricted ranges are more limited by local topographical features such as mountain ranges, rivers, valleys and coastal areas; whereas animals with larger geographical scale should be limited more by large climatic zones (Brown, 1995; Brown & Maurer, 1989). However, ectothermic species do not follow a pattern, is have the climatic zones as the main limiting factor, responding differently to different groups (Pfrender *et al.*, 1998). Therefore, climate and dispersal limitation (e.g. body size) are both critical determinants of species ranges for a variety of taxonomic groups in different regions (Baselga *et al.*, 2012). Small species for

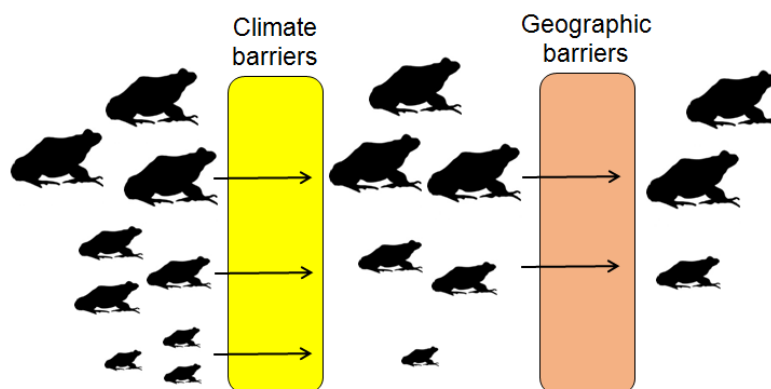
example, can lost water for the environmental and suffer desiccation (MacLean, 1985), furthermore the interaction between species (predation and competition) and adaptive processes to the environment (generalist, specialist or opportunistic) are hypotheses that can determine the difference in the composition of species in a particular locality (Bell, 2001; Legendre *et al.*, 2005). In this sense, potential dispersal of the ectothermic species for many times, is related to their morphological and physiological characteristics as a limiting factor (Jimenez-Valverde *et al.*, 2015) and this may be a key point to understand these processes. Functional traits may determine species composition to the characteristics of a particular ecosystem (Díaz *et al.*, 2007) and determine the species dispersion processes (Gómez-Rodrigues *et al.*, 2015). As well as body size tends to be higher in cold climates (Bergmann's rule, Bergmann, 1847) but this pattern can vary depending on the taxon (Ashton & Feldman, 2003; Angilletta *et al.*, 2004; Cruz *et al.*, 2005; Pincheira-Donoso *et al.*, 2007; Shelomi, 2012). Another pattern related to body size and Rappaport's rule which the species are generally smaller at lower latitudes than at higher latitudes (Rappaport, 1982). However, this rule has generated many doubts because to be little tested, especially being a pattern observed in the northern Hemisphere (Gaston *et al.*, 1998; Whitton *et al.*, 2012).

Amphibians are sensitive to environmental changes (Blaustein *et al.*, 1994) and because of its physiological characteristics sensitive to the environment are limited to dispersal (Duellman & Trueb, 1994). Furthermore, many are prey to vertebrates and invertebrates (Wells, 2007), which makes amphibians dependent on their adaptive morphological and physiological specialization to succeed in dispersing. The Atlantic Forest is considered one of the world hotspots due to its high species diversity, endemism and degree of threat to extinction. (Myers *et al.*, 2000). Anthropogenic



impacts, mostly over the last 500 years, has dramatically altered the landscape (Ribeiro *et al.*, 2009). These impacts have been responsible for the well documented decline of amphibian populations (Lambert, 1997; Lips & Donnelly, 2002; Becker *et al.*, 2007, 2010; Ferreira *et al.*, 2016). Understanding beta diversity patterns and evaluating the different compositions (turnover or nested) along of latitudinal and longitudinal gradient be an important tool for understanding the processes of dispersion of the species (Baselga, 2008, 2010).

Knowing that amphibians are limited to dispersion due to their physiological characteristics (e.g. body size), we evaluated the beta diversity of amphibians in the Atlantic Forest and evaluated the ability of species dispersion based on their functional traits. For this, we tested the hypothesis that (i) functional traits promoting greater adaptation to new environmental conditions (e.g. body size and specializations of habitat) may explain the current pattern of amphibians composition in the Atlantic Forest (Fig. 1); (ii) species that occur of Atlantic Forest are smaller in high altitudes and smaller in lower latitudes. We evaluated species composition of territories inside in this biome, and correlate the richness and endemism of species to environmental variables and check the average body size of species in different topographies and spatial references.

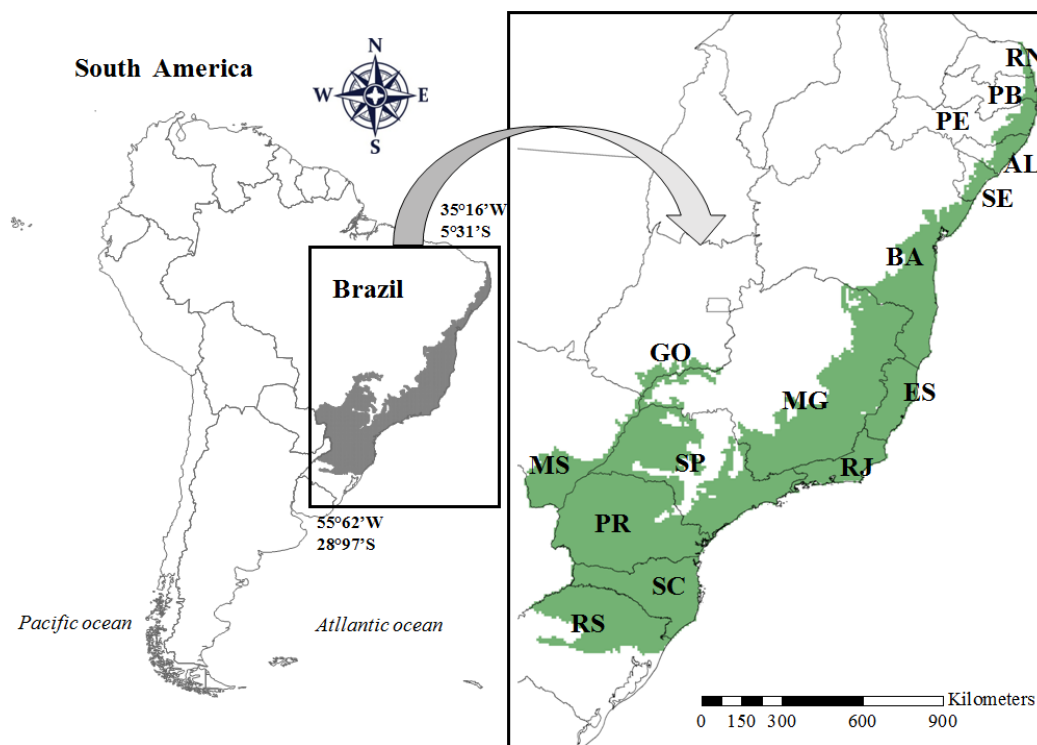


**Figure 1.** Hypothetical diagram showing that larger and more adapted species to environmental variables can be dispersed more easily in the current Atlantic Forest. Larger species lose less water to the environment than small or medium body sized species. Larger and medium species have more energy and ability to disperse by geographical barriers.

## 2.2 METHODS

### 2.2.1 Study region

The study area was the Atlantic Forest hotspot, is a biome that has latitudinal range extending into the tropical and subtropical regions (Myers *et al.*, 2000; Ribeiro *et al.*, 2009). The longitudinal range favor differences in forest composition due to a diminishing gradient in rainfall from coast to interior (Ribeiro *et al.*, 2009) and an altitudinal range encompassing the elevational limits of the mountain from 0-2000 m a.s.l. (Cavarzere & Silveira, 2012). Originally this biome covered around 150 million ha with heterogeneous environmental conditions provided by a wide range of climatic belts and vegetation formations (Tabarelli *et al.*, 2005; Ribeiro *et al.*, 2009) (Fig. 2).



**Figure 2.** Map showed the study area of Brazilian Atlantic Forest hotspot (in gray) in South American territory. The acronyms of territories studied: RN = Rio Grande do Norte, PB = Paraíba, PE = Pernambuco, AL = Alagoas, SE = Sergipe, BA = Bahia, ES = Espírito Santo, MG = Minas Gerais, RJ = Rio de Janeiro, SP = São Paulo, GO = Góias, PR = Paraná, MS = Mato Grosso do Sul, SC = Santa Catarina, RS = Rio Grande do Sul.

The Brazilian Atlantic Forest occurs in 15 states (Fig 2). For better understand the pattern of beta diversity, we analyzed these 15 states for aid visualize the differences of species compositions (richness and endemism) and a possible dispersion route. We divided the territories based in i) size of territory; ii) forest composition of each territory; and iii) abiotic environmental features of each territory (Olson *et al.*, 2001; Ribeiro *et al.*, 2009, Worldclim database see below).

Two states have all their territory included as composition of the species, RJ (Rio de Janeiro state) and ES (Espírito Santo state) because to be smaller and their

forest composition similar. Four states have their territories separated in eastern and western, because of be large states and with deferent's forest composition (eastern rain forest, western seasonal forest): EPR (Eastern of Paraná state), WPR (Western of Paraná state), ESC (Eastern of Santa Catarina state), WSC (Western of Santa Catarina state), ERS (Eastern of Rio Grande do Sul state), WRS (Western of Rio Grande do Sul state); and the "SMGM" refers to four connected states in seasonal forests (include Western of São Paulo -S, North of Mato Grosso do Sul -M, South of Góias -G and extreme South of Minas Gerais - M). The region of Mato Grosso do Sul state localized in the western of Paraná state was analyzed only, MS (Mato Grosso do Sul state). The Pernambuco (PE), Sergipe (SE), Alagoas (AL), Paraíba (PB) and Rio Grande do Norte (RN) states were included in an only region N (Northeast), because the smaller territories (inside of biome), similar forest composition and abiotics environmental features.

Finally two states were separated and region North and South, because the large territory and different forest composition and abiotics environmental features, SBA (Southern of Bahia state), NBA (Northern of Bahia state), SMG (Southern of Minas Gerais state) and NMG (Northern of Minas Gerais state). In total sixteen territories of the study were assessed (see Fig. 3d).

### 2.2.2 Sampling

Based on the list of species that occur in the Brazilian Atlantic Forest according to Haddad *et al.* (2013), we used species range maps obtained from the International Union for Conservation of Nature (IUCN) database (<http://www.iucnredlist.org/technical-documents/spatial-data>) to determine the full

recorded range of species distribution. In addition, we conducted amphibian sampling in 11 protected areas (PA) of Atlantic Forest (see Appendix 1, Fig. S1), as a complementary fieldwork and to supplement the dataset with observed functional traits (body size, reproductive mode, habitat, activity, poison patterns and habit).

In total, we assessed the geographical ranges of 529 amphibian species (519 Anurans and 10 Gymnophionas) covered by our grid system. We used ArcGIS 10 software (ESRI, 2010) to build presence/absence matrices from the species distribution data by superimposing a grid system with cells of 0.1 latitude/longitude degrees, creating a network with 10,359 grid cells. We followed Frost (2016) for the amphibian nomenclature.

### 2.2.3 Environmental variables

We averaged six environmental variables for each grid cell, which included one topographic (altitude), one biotic (tree cover) and four climatics (annual precipitation, mean annual temperature, annual evapotranspiration, and net primary productivity). We obtained altitude, annual precipitation and mean annual temperature from the World Clim database with 0.05° resolution (<http://www.worldclim.org/>). We also obtained annual evapotranspiration (AET), from the Geonetwork database (<http://www.fao.org/geonetwork/srv/>); net primary productivity (NPP) from the Numeral Terra dynamic Simulation Group (<http://www.ntsg.umd.edu/data>), and land cover from Global Forest Change 2000-2014 database ([http://earthenginepartners.appspot.com/science-2013-global-forest/download\\_v1.2.html](http://earthenginepartners.appspot.com/science-2013-global-forest/download_v1.2.html)). All of these variables are known to represent either

potential physiological limits for amphibians or barriers to dispersal (Vasconcelos *et al.*, 2010; Silva *et al.*, 2012). We draw the maps using the software and ArcGis10 (ESRI, 2010).

#### 2.2.4 Functional traits

We characterized ecological traits of 529 amphibian species according to Haddad *et al.*, (2013), ecological information available in IUCN data base, literatures on the original descriptions of the species and to our field observations. Ecological traits were: body size, development mode (direct or indirect), habitat (forested areas, open areas, and both open and forested areas), activity time (nocturnal, diurnal, and both), poisonous (toxic, unpalatable or bad odour, and non-toxic), habit (arboreal, phytotelmate, terrestrial, cryptozoic, fossorial, rheophilic, semi-aquatic, and aquatic) (See Appendix 1, Table S1 for details and references).

#### 2.2.5 Data Analyses

##### 2.2.5.1 *Environmental variables vs species richness and species endemics*

We evaluated the response of species richness and areas with richness of species endemic (separated) to the predicted variables: altitude, annual precipitation, mean annual temperature, annual evapotranspiration, net primary productivity, and tree cover. We defined the endemic species than occur in only one of 16 territories defined above (see Fig. 3d) and calculated the richness species and richness of endemic species for each grid cell of 0.05° (10,359 cells) separately. We used permutation multivariate

analysis of variance (PERMANOVA), with 1,000 permutations based on a Euclidean distance matrix through the “adonis” function in the package 'vegan' (Oksanen *et al.*, 2013) in R (R Development Core Team, 2012).

#### 2.2.5.2 *Distribution vs functional traits*

In order to evaluate the response of the geographical range of each species to the predicted body size (categorized - small <3cm, medium  $\geq 3$  and <10cm, large >10cm – according to Haddad *et al.*, 2013), poisonous, development mode, habit, habitat and activity for Anura and body size, development mode, habitat and habit for Gymnophiona. We also used permutation multivariate analysis of variance (PERMANOVA), with 1,000 permutations. In addition, we used Boxplots to visualize the traits that better explained species distribution. These analyzes were performed separately for Anuran and Gymnophiona due their different characteristics of ecological traits and body size (Haddad *et al.*, 2013). We calculated for each grid cell of 0.05° (10,359 cells) and performed these analyses using the package 'vegan' in R.

#### 2.2.5.3 *Effects of body size vs richness*

We used simple linear models to test the association of body size (original body size of each species) of amphibians on species richness. For this, we calculated species mean of body size for each grid cell of 0.05° (10,359 cells). After, we evaluated the relationship between species mean of body size to variations of latitude, longitude and

altitude. We performed these analyses using the package 'vegan' (Oksanen *et al.*, 2013) in R (R Development Core Team, 2012).

#### 2.2.5.4 *Similar species groups*

First, we examined whether there was independence of spatial correlation of species composition among the 16 territories (matrix of spatial data vs matrix of species composition). For this, we used Pearson correlation tests using the Mantel permutation (Legendre & Legendre, 1998).

Second, we used a similarity measure (i.e. Euclidean distance matrix) of the 16 territories to rank the groups of similar species composition. Ordination of the 16 territories was based on this faunal dissimilarity matrix, which was submitted to a nonparametric multidimensional scaling analysis (NMDS, Legendre & Legendre 1998) and the most likely solution was evaluated by Pearson correlation. The calculation of the variance is captured by a regression matrix from the original distances (Bray-Curtis) and the final array distances (Euclidean).

Third, we draw a dendrogram by taking Euclidean distance as the measure of resemblance and average linkage procedure as the linkage rule (Fig. 3c, b). These tests were computed in R using the “vegan” package.



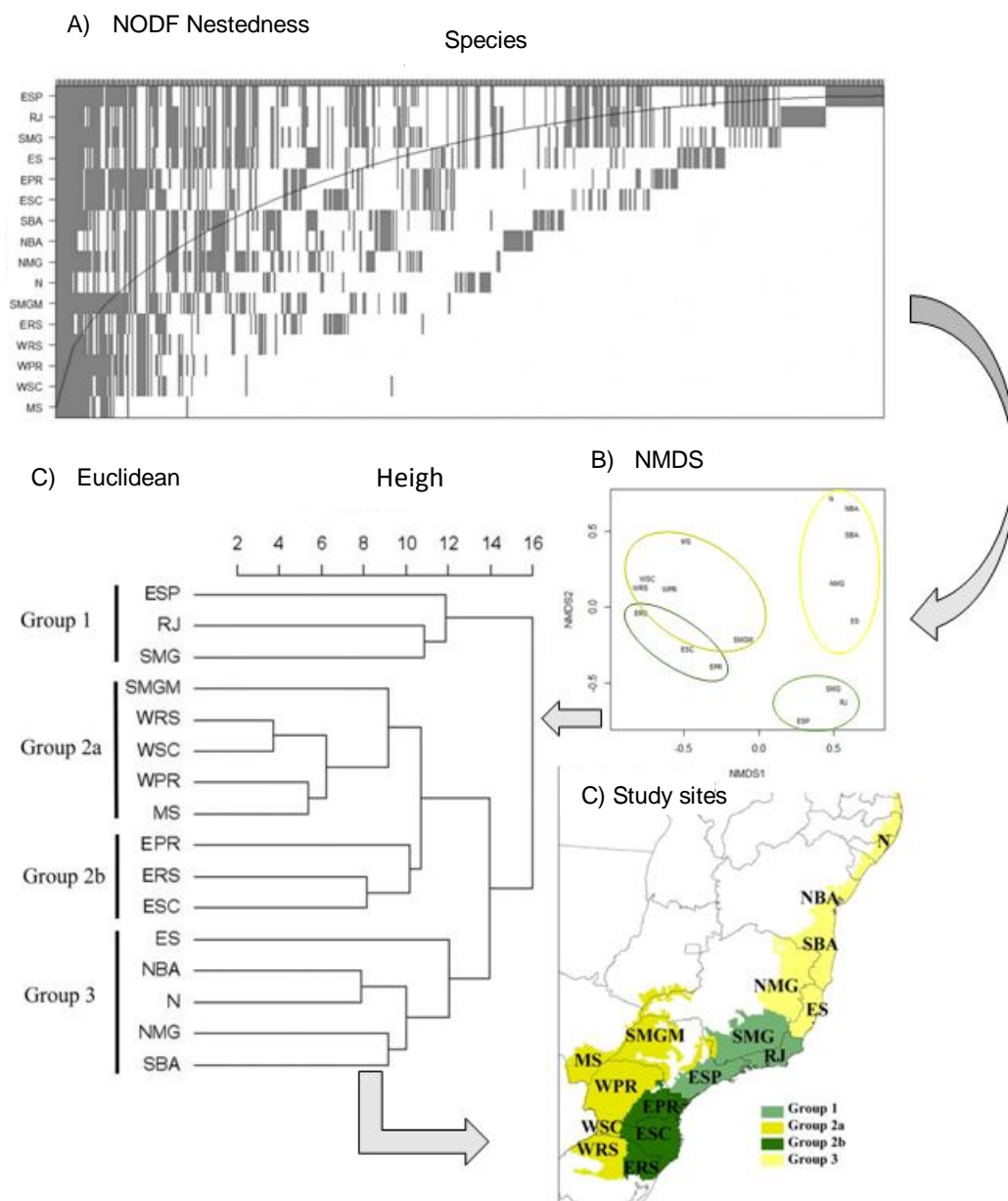
#### 2.2.5.5 *Beta diversity measures*

First, we used two complementary metrics for beta diversity analysis by considering species presence and absence. In order to determine if the pattern among anuran communities is nested, we calculated a nestedness metric based on overlap and decreasing fill (index NODF) of Almeida-Neto *et al.*, (2008) and Ulrich *et al.*, (2009) across the 16 territories. The matrix in decreasing both columns and rows, columns ranking the areas according to their species richness and ordering the species in lines, the most frequent to the rarest. Creating and ordering the territories as larger biota and its subsets biota. To perform this analysis, original matrices were submitted to 1,000 simulations. We performed this analysis for all localities separately using the package “vegan” in R.

Second, we conducted beta diversity partitioning and computed distance matrices using pairwise dissimilarities  $\beta_{sor}$  (i.e. measure total beta diversity),  $\beta_{sim}$  (i.e. measure spatial turnover), and  $\beta_{nes}$  (i.e. measure nesting) (Baselga, 2010). We computed these analyses among the 16 territories to show the directions of the species distributions. We considered larger nesting values means more species similar to the area of major species richness, and larger numbers of turnover means less similar species composition. We follow the sequence provided by NODF for analysis of beta diversity partitioning (Fig. 3a).

To available the effects of variation of species composition, we use the classification of Veloso *et al.*, (1991) based on the type of forest composition and topography. We computed the analyses in four ways: 1) all altitudes, 2) from 0-300m of

altitude, 3) from 300-700m of altitude, 4) from 700-2000m of altitude. All these tests used the “betapart” package (Baselga & Orme, 2012) in R.

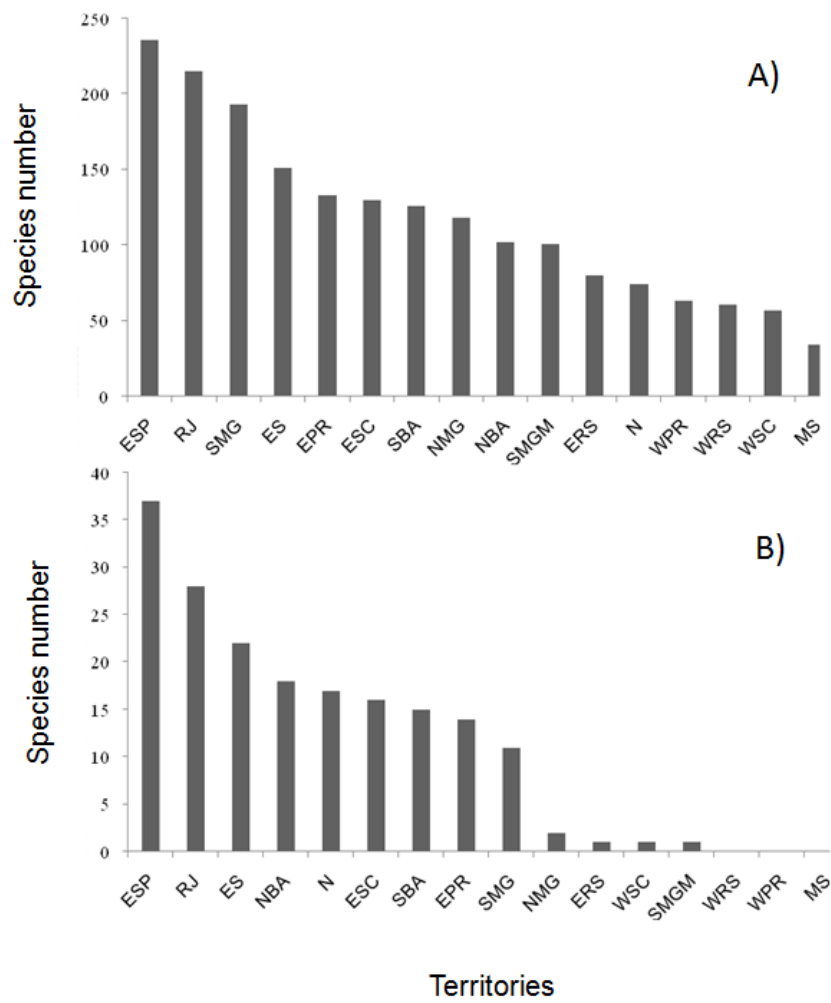


**Figure 3.** Steps designed to determine the beta diversity in Atlantic forest. A) NODF created to determine the territories with greater species richness and subset, B) NMDS used to spatially group the territories based on distance and species composition, C) visualization cluster of results obtained by NMDS, and D) created map showing groups similarities.

## 2.3 RESULTS

We found that the abiotics enviromental features explained 59.5% of the species richness distribution according to PERMANOVA values. The temperature was the main variable, (39.3%) followed by precipitation (11.4%), and NPP (8.6%) (Table 1). For territories of species endemic the abiotics enviromental features explained 26% of the endemism. Temperature was the main variable (22%), followed by altitude (2%) (Table 2 and see Fig. 4 for territories and their respective scores of species richness and endemism).

The functional traits explained 15.8% of distribution the anuran species in the Atlantic Forest according to PERMANOVA values. The habitat was responsible for about 9.0%, body size 3.0%, habit 1.0% and palatability 0.8% of the variation in the species distribution. The development mode and activity traits did not showed any significant relationship to species distribution (Table 3). For Gymnophiona, two main traits explained 88.8% of the species distribution. Habitat was responsible for about 67.4% and development mode 15.7% of the variation in the species richness distribution. The body size and habit traits did not showed any relationship to species distribution (Table 4).



**Figure 4.** Scores of species richness (A) and endemism (B) in the territories considered as sampling units.

**Table 1.** Association of amphibian species richness to environmental variables in the Brazilian Atlantic Forest by the PERMANOVA.

Variables	df	<i>F</i> model	<i>R</i> <sup>2</sup>	<i>P</i> value
Altitude	1	4.4	0.00017	0.036
AET - Evapotranspiration	1	38.3	0.0015	0.001
NPP - Net primary production	1	2198	0.08597	0.001
Precipitation	1	2917.4	0.11411	0.001
Temperature	1	10046.3	0.39294	0.001
Tree cover	1	10.9	0.00043	0.001
Residuals	10352	–	0.40489	–
Total	10358	–	1.00	–

**Table 2.** Association of amphibian species endemics to environmental variables in the Brazilian Atlantic Forest by the PERMANOVA.

Variables	df	<i>F</i> model	<i>R</i> <sup>2</sup>	<i>P</i> value
Altitude	1	413.2	0.02948	0.001
AET - Evapotranspiration	1	19.3	0.00137	0.001
NPP - Net primary production	1	4.1	0.00029	0.049
Precipitation	1	0.3	0.00002	0.578
Temperature	1	3202.0	0.22845	0.001
Tree cover	1	25.5	0.00182	0.001
Residuals	10352	–	0.73856	–
Total	10358	–	1.00	–

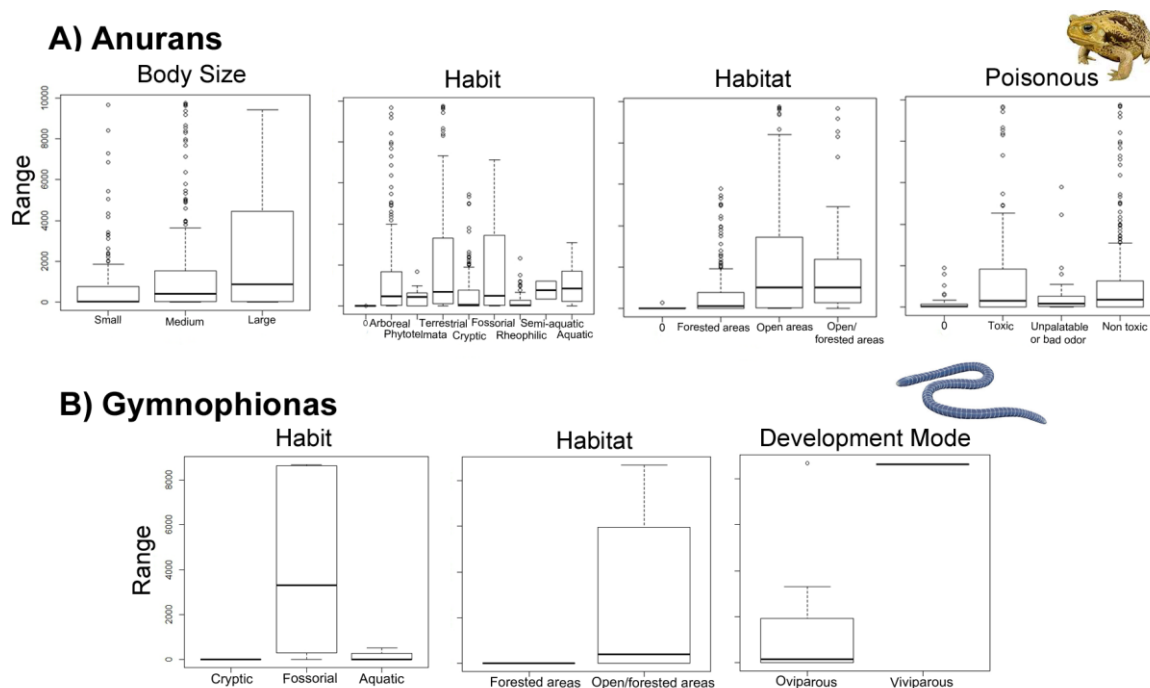
**Table 3.** Association of species range to the functional traits to Anuran in the Brazilian Atlantic Forest by the PERMANOVA.

Variables	df	<i>F</i> model	<i>R</i> <sup>2</sup>	<i>P</i> value
Activity	1	0.086	0.00014	0.772
Body size	1	22.14	0.03613	0.001
Poisonous	1	5.011	0.00818	0.021
Habit	1	12.056	0.01967	0.002
Habitat	1	57.575	0.09394	0.001
Development mode	1	1.013	0.00165	0.312
Residuals	515	–	0.84029	–
Total	521	–	1.00000	–

**Table 4.** Association of species range to the functional traits to Gymnophionas in the Brazilian Atlantic Forest by the PERMANOVA.

Variables	Df	<i>F</i> model	<i>R</i> <sup>2</sup>	<i>P</i> value
Body size	1	1.346	0.02823	0.282
Habit	1	2.674	0.05609	0.175
Habitat	1	32.126	0.67401	0.001
Development mode	1	7.519	0.15775	0.027
Residuals	4	–	0.08392	–
Total	8	–	1.00000	–

The Boxplot reveal the main responsible of largest distribution in Anura is found between species of open areas and species of open and forested areas, species of large body size, terrestrial and fossorial species, and toxic and non-toxic species (Fig.5a). The largest distribution in Gymnophiona is found among species of open and forested areas and species of indirect development mode (Fig. 5b).

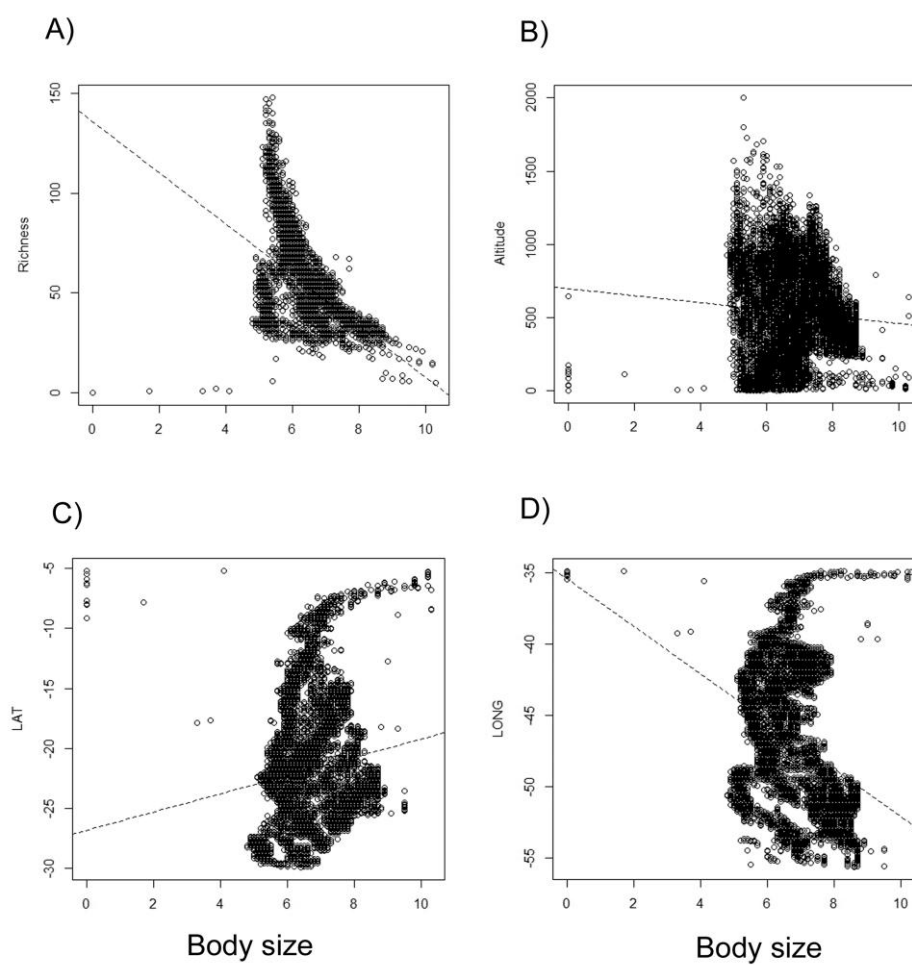


**Figure 5.** Box plot showing the traits selected by PERMANOVA and its variations that explained the distribution of species.

The body size showed high relation with distribution and species richness. Our results of linear models of mean body size was significant in all analyzes ( $p < 0.001$ ). Species richness had high relationship to smaller body size ( $r^2 = 0.398$ ). The average body size increases by increasing longitude ( $r^2 = 0.103$ ). The average body size decreases by increasing latitude, ( $r^2 = 0.025$ ) and altitude ( $r^2 = 0.006$ ) (Table 5, Fig. 6).

**Table 5.** Correlation used a linear regression of body size vs. species richness, longitude, latitude and altitude.

Variables	Body size	
	$r^2$	$P$
Richness	0.398	$p < 0.001$
Longitude	0.103	$p < 0.001$
Latitude	0.025	$p < 0.001$
Altitude	0.006	$p < 0.001$



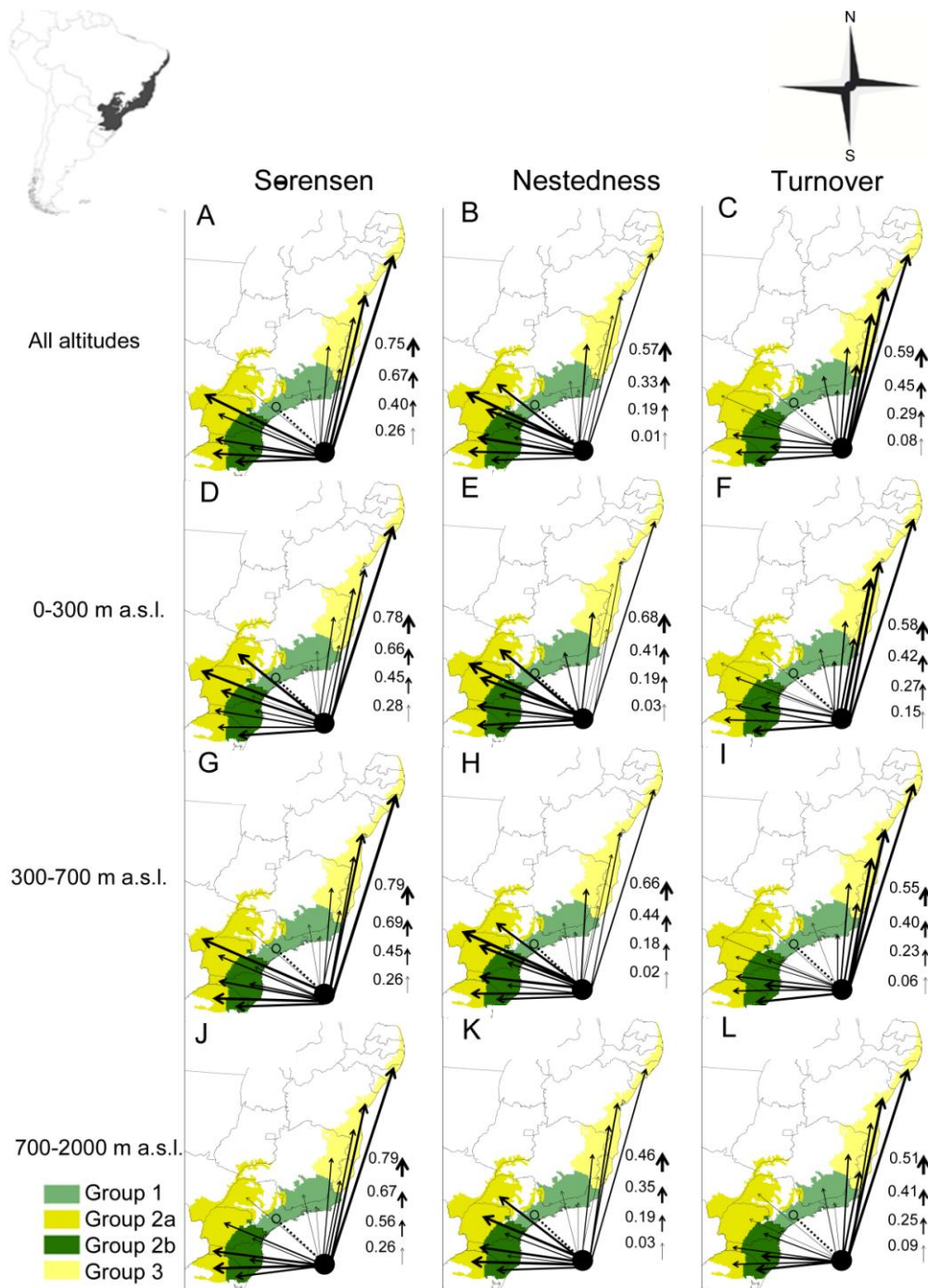
**Figure 6.** A) Relation between body size and species richness, B) altitude, C) latitude (LAT), and D) longitude (LONG).



The Mantel tests indicated spatial correlation of species composition and distance among the 16 territories ( $r^2 = 0.30$ ,  $p = 0.019$ ). The dendrogram used the scores of the NMDS axes (7 dimensions,  $r^2 = 0.99$ ,  $p < 0.001$ ), showed the presence of three groups of similarity faunal regions in Atlantic Forest (Fig. 3b-d). The major biota was ESP, RJ and SMG (group 1), characterized by humid forests major species richness and endemism. The group 2 was divided into two subgroups; one formed by MS, WPR, WSC WRS, and SMGM (subgroup 2a), with drier forests and seasonality with less species richness and endemism, and another one formed by EPR, ESC and ERS (subgroup 2b), characterized by humid forests with presence of Araucarias forest. The group 3 was formed by ES, NMG, SBA, NBA and N, characterized by hot forests and little or no seasonality, its second major group with species richness and endemism (see Fig. 4).

Results of the NODF values indicated significant nesting (NODF = 38.7,  $p < 0.001$ ; N rows = 57.7, N columns = 38.6) showing the areas ESP, RJ and SMG as the major biota (Fig. 3a). The beta diversity partitioning revealed that the highest values were between groups 1 and 3 ( $\beta_{sor}$  mean  $0.684 \pm 0.093$ ) followed by the group 2a ( $\beta_{sor}$  mean  $0.675 \pm 0.126$ ). Among the group 3, the values increase by increasing of the difference in species composition according to altitude, decreasing  $\beta_{sim}$  and increasing  $\beta_{nes}$ ; wherein the territory N has highest  $\beta_{nes}$  (0.281) among the species that occur 700-2000m a.s.l., and the territory SBA has highest values of  $\beta_{sim}$  (0.582) among the species composition that occur in 300m a.s.l. There is similar pattern among the group 2a, however the  $\beta_{nes}$  decreases with increasing altitude and  $\beta_{sim}$  decreases then increases slightly at higher altitudes, wherein the territory MS has highest values of  $\beta_{nes}$  (0.669) whereas that territory WRS has highest values of  $\beta_{sim}$  (0.397) both among

species which occur between 300-700m a.s.l.. Among the groups 1 and 2b the values of  $\beta_{sor}$  increases abruptly with increasing altitude ( $\beta_{sor}$  mean 0.215-0.609). The main difference in this value of  $\beta_{sim}$  (0.353-0.395) increases according to increasing the differences in the species composition between altitudes. The territory ERS had the highest value of  $\beta_{nes}$  (0.281) among the species that occur from 700-2000 m a.s.l. and the highest value of  $\beta_{sim}$  (0.480) among the species that occur 300-700m a.s.l. (see Table 6 and Fig. 7).



**Figure 7.** Beta diversity of amphibians in the Brazilian Atlantic Forest. Thinner arrows indicate smaller values, thicker arrows larger values. Beta diversity values are from the large biota ESP (circle) for the subsets.

**Table 6.** Partitioning of beta diversity. Analyses were made from the territory with greater species richness (ESP) from the other subsets. A mean of the results of each territories and each group and presented for evaluation difference between the species similarity groups. In bold are shown the greatest beta diversity values.

Groups	Sorensen ( $\beta_{sor}$ )				Nestedness ( $\beta_{nes}$ )				Turnover ( $\beta_{sim}$ )			
	All altitudes	0-300m	300-700m	700-2000m	All altitudes	0-300m	300-700m	700-2000m	All altitudes	0-300m	300-700m	700-2000m
ESP	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
RJ	0.282	0.263	0.263	0.269	0.035	0.016	0.029	0.039	0.247	0.247	0.234	0.229
SMG	0.329	<b>0.403</b>	0.268	0.314	0.075	0.195	0.069	0.060	0.254	<b>0.416</b>	0.199	0.254
Mean	0.203	0.222	0.177	0.194	0.037	0.070	0.033	0.033	0.167	0.332	0.217	0.242
DP $\pm$	0.178	0.205	0.153	0.170	0.037	0.108	0.035	0.031	0.144	0.119	0.025	0.017
<b>Group2a</b>												
SMGM	<b>0.460</b>	<b>0.751</b>	<b>0.446</b>	<b>0.448</b>	0.361	<b>0.551</b>	0.381	0.357	0.099	0.200	0.065	0.091
WPR	<b>0.666</b>	<b>0.719</b>	<b>0.736</b>	<b>0.654</b>	<b>0.459</b>	<b>0.452</b>	<b>0.444</b>	<b>0.460</b>	0.206	0.267	0.210	0.194
WSC	<b>0.734</b>	<b>0.713</b>	<b>0.727</b>	<b>0.741</b>	<b>0.418</b>	<b>0.436</b>	0.393	<b>0.447</b>	0.316	0.277	0.333	0.294
MS	<b>0.770</b>	<b>0.750</b>	<b>0.757</b>	<b>0.000</b>	<b>0.682</b>	<b>0.574</b>	<b>0.669</b>	0.000	0.088	0.176	0.088	0.000
WRS	<b>0.744</b>	<b>0.710</b>	<b>0.749</b>	<b>0.785</b>	0.367	0.330	0.353	<b>0.460</b>	0.377	0.379	0.397	0.326

Mean	<b>0.675</b>	<b>0.729</b>	<b>0.683</b>	<b>0.657</b>	<b>0.457</b>	<b>0.469</b>	<b>0.448</b>	<b>0.431</b>	0.217	0.260	0.218	0.226
DP ±	0.126	0.020	0.133	0.150	0.132	0.098	0.128	0.050	0.128	0.079	0.146	0.106
<b>Group2b</b>												
EPR	<b>0.491</b>	0.248	<b>0.457</b>	<b>0.499</b>	0.197	0.248	0.237	0.194	0.293	0.154	0.220	0.305
ESC	<b>0.612</b>	0.148	<b>0.592</b>	<b>0.595</b>	0.158	0.148	0.181	0.168	<b>0.454</b>	<b>0.429</b>	<b>0.410</b>	<b>0.427</b>
ERS	<b>0.747</b>	0.250	<b>0.736</b>	<b>0.733</b>	0.247	0.250	0.256	0.281	<b>0.500</b>	<b>0.478</b>	<b>0.480</b>	<b>0.452</b>
Mean	<b>0.616</b>	0.215	<b>0.595</b>	<b>0.609</b>	0.201	0.215	0.225	0.214	<b>0.416</b>	0.353	0.370	0.395
DP ±	0.128	0.058	0.140	0.118	0.044	0.058	0.039	0.059	0.109	0.175	0.134	0.079
<b>Group3</b>												
ES	<b>0.576</b>	<b>0.511</b>	<b>0.546</b>	<b>0.567</b>	0.119	0.095	0.142	0.153	<b>0.457</b>	<b>0.416</b>	<b>0.404</b>	<b>0.414</b>
SBA	<b>0.702</b>	<b>0.673</b>	<b>0.693</b>	<b>0.679</b>	0.130	0.091	0.152	0.268	<b>0.571</b>	<b>0.582</b>	<b>0.541</b>	<b>0.412</b>
NBA	<b>0.757</b>	<b>0.718</b>	<b>0.743</b>	<b>0.765</b>	0.159	0.144	0.187	0.258	<b>0.598</b>	<b>0.574</b>	<b>0.556</b>	<b>0.507</b>
NMG	<b>0.599</b>	<b>0.576</b>	<b>0.589</b>	<b>0.589</b>	0.201	0.199	0.207	0.243	0.398	0.378	0.382	0.346
N	<b>0.787</b>	<b>0.757</b>	<b>0.791</b>	<b>0.798</b>	0.233	0.195	0.245	0.281	<b>0.554</b>	<b>0.562</b>	<b>0.545</b>	<b>0.517</b>
Mean	<b>0.684</b>	<b>0.647</b>	<b>0.672</b>	<b>0.680</b>	0.168	0.145	0.187	0.241	<b>0.516</b>	<b>0.502</b>	<b>0.486</b>	<b>0.439</b>
DP ±	0.094	0.102	0.103	0.103	0.048	0.052	0.042	0.051	0.085	0.098	0.085	0.072

## 2.4 DISCUSSION

Our results showed that mean annual temperature has high influence in the amphibian richness and endemic species of the Atlantic Forest. The temperature between 19° C and 21° C is the optimal range for amphibian, promoted high species richness. Because of their physiological characteristics, amphibians are dependent of humidity and mild temperatures (Wells, 2007; Crump, 2010), thus is expected the correlation of species richness to precipitation and temperature (Casemiro *et al.*, 2007; Ortiz-Yusty *et al.*, 2013; Vasconcelo *et al.*, 2014). The correlation of species richness with NPP and vegetation also is strongly related to rainfall; plants are strongly correlated to water-balance (Bradford *et al.*, 2003; Rueda *et al.*, 2010). Moreover, altitude areas have a higher number of endemic species that due to the milder temperature is higher humidity. However, lowland areas also have endemism, and this may be related to historical events (Carnaval *et al.*, 2012).

Because of high humidity and diversity available microhabitats, many Neotropical amphibians evolved to decrease its size and many species are miniaturized (Rittmeyer *et al.*, 2012). The miniaturized species are more susceptible to humidity loss (MacLean, 1985) and many are restricted to forest formations preserved to maintain the necessary physiological features (Lourenço-de-Moraes *et al.*, 2012, 2014, Ferreira *et al.*, 2016). Because of this, miniaturized species are concentrated in areas of milder temperatures, higher rainfall, and higher vegetation cover. For amphibians in Atlantic Forest, high latitudes, longitudes and altitudes tend to have smaller species. Therefore, the relationship between species richness to the mean body size of species is directly related to environmental conditions. This may have led to a restricted gene flow

between populations and thus accelerate genetic differentiation (Pabijan *et al.*, 2012). Consequently, it may have limited these species in rainforests where such genetic differentiation is most commonly observed (Rodríguez *et al.*, 2015), as these factors may have contributed to the large number of small species in small intervals.

The association for the current dispersion or expansion of its occurrence areas, are to adaptation for live in open areas, have medium to large size and be toxic. In our study, it is the species with the largest body size have better dispersion capabilities (specialists in open areas or edge forest) as species *Rhinella shineideri*, *Odontophrynus americanus* and *Siphonops annulatus* (Haddad *et al.*, 2013). Species with the same features than this species but not toxic or unpalatable, also had higher ranges possibly because of other factors that may assist in their range expansion. Species with a variety of antipredator mechanisms may be more likely to avoid a wider range of predators (Lourenço-de-Moraes *et al.*, 2016), that allow achieve success in the dispersion. In addition, species of open areas with higher ranges also occur in drier biomes such as the Cerrado and the Caatinga.

According to our results, the current dispersion of species is directly related to their functional traits, being the habitat in which the species and adapted their biggest asset - species than tolerate high temperatures and low humidity rates. Most of the species that occur in the Atlantic Forest are under 30mm. Small species lose water more quickly to the environment (MacLean, 1985). Deforestation has limited most species of the Atlantic Forest to small scattered fragments. Even small species as *Dendropsophus nanus* and *D. minutus* have great ability to dispersion due to their ability of occurring in open areas (Haddad *et al.*, 2013). Many species of open areas are expanding or expanded their ranges due to forest destruction. Species of open areas are not found in

forests, or they are found in low abundance, we suggest that these species are not generalists, but specialists of open areas and opportunistic.

Because of their functional traits our results showed that the species of open areas to disperse from West to East and North of Atlantic Forest, and this influence of the results of beta diversity (nestedness). The mountains of Serra do Mar and Mantiqueira can be limiting geographical barriers for small amphibians and strictly forest habits (Haddad, 1998; Morelato & Haddad, 2000). Moreover, the geographical barrier Rio Doce divides the territory ES and part of NMG (Bates *et al.*, 1998; Costa *et al.*, 2000) and influenced in dispersion and composition on species of group 3. ES has southern (group 1) and northern (group 3) species of the Atlantic Forest, while NMG is more related to northern (group 3) species of the Atlantic Forest. Few strictly forest species have large ranges in the Atlantic Forest. *Haddadus binotatus* is a species with great range possibly it was dispersed by the drier forests of the West, or during the Glacial period due to ocean regression (Atlantic Forest hypothesis, see Leite *et al.*, 2016). Moreover, it is possible that this is a species complex (Dias *et al.*, 2011). The same applies to small species with wide distribution as *Dendropsophus minutus* in which Gehara *et al.*, (2014) indicated through molecular techniques that is a species complex.

Due to the consequent expansion of open areas species (trait habitat), strongly reflected in the beta diversity values. Our results pointed out nesting differences between the groups 1 and 2a, showing a decrease of nesting by increasing altitude and increasing turnover for the species occurring between 700-2000m and 0-300m altitude. Between the groups 1 and 2b, there is a reduction of nesting and increased of turnover by increasing elevation which has slight increase in beta diversity between species that



occur 300-700m altitude. Between the groups 1 and 3 the nesting increases with increasing altitude, reducing the turnover. These results indicate differences in the compositions according to its topographic location. The main differences between the compositions are among the species that occur in the lowland (0-300m) and hilltops (700-2000m). The group 2b indicates higher endemism rate on the mountainous region while the group 3 in the lowlands. Therefore, mountain areas are important geographical barriers to amphibian species in the Atlantic Forest.

According to our data, it is possible to separate the Atlantic Forest in three major regions of endemism (groups 1, 2b and 3). The beta diversity values corroborate the hypothesis of endemism during the Pleistocene glacial (Carnaval *et al.*, 2009; Carnaval *et al.*, 2014). The two extremes of this biome (i.e. Southern most and Northern most), have higher turnover rates compared to group 1. The group 3 has shared genera with the Amazon forest as *Pristimantis*, *Adelophryne* and *Alophryne* - genera that do not occur in other groups of the Atlantic Forest. These data support the hypothesis connection between the North of the Atlantic Forest and the Eastern Amazon rainforest (Batalha-Filho *et al.*, 2013; Sobral-Souza *et al.*, 2015). On the other hand, the group 3 also received species from the Atlantic Forest as *Hypsiboas faber*. The 2a and 2b group also have higher turnover rates at some territories. In the group 2a, WRS and in group 2b the territories ESC and ERS. The South of the Atlantic Forest region had a strong influence of the Western Amazon composition and the Andean forests (Batalha Filho *et al.*, 2013; Sobral-Souza *et al.*, 2015). Most *Melanophryniscus*, *Scythrophrys* and *Lymnomedusa* species of the genus occur in group 2. These genera tolerate colder areas of the Atlantic Forest. Moreover the nesting territories of greatest values were in group 1 and group 2a. This group has areas of warmer forests and more pronounced seasonality due to these

species that occur at these points, especially in MS and WPR are mostly species of open areas. In the SMGM area, species that occur at this point are directly related to the Cerrado biome; many species of open areas and forest edges occur at this point. Finally, our data point groups 1, 2b and 3 as the areas with the highest rates of endemic and rare species.

Historical events (e.g. Pleistocene glacial) can be the main direction of the species richness of amphibians in the Atlantic Forest. This richness provided by these events direct speciation and different specializations with creating various functional traits, which depending on the specialization, will direct the dispersion of species and a current historical panorama. This process is a cycle that has been repeated for thousands of years, however the current change on the planet by human actions, can direct a mass extinction process in the Anthropocene Age (Barnoski *et al.*, 2011; Dirzo *et al.*, 2014). The current spatial composition and dispersion of species is related to anthropogenic actions and the species most suitable for the new environment prosper, while forest and exclusive species of the Atlantic Forest, are even more endemic and endangered. Our results can help define new strategies aimed at conservation of the original forests of the Atlantic Forest biome, promoting connections between the reserves and the creation of new reserves.

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### **3 PHYLOGENETIC AND FUNCTIONAL DIVERSITY REVEAL NEW CLIMATIC REFUGES FOR AMPHIBIANS IN THE BRAZILIAN ATLANTIC FOREST**

#### **Abstract**

Climate changes have occurred on Earth over millions of years. To survive these changes the species took refuge in places where the changes were less drastic. Climate change is a real panorama, and conservation strategies should be developed. Ectothermic species, are especially sensitive of the climate change, which makes them good references for studies involving climate change. Analyze the losses against the ecosystem and evolutionary values, can help to understand the processes and minimize their impact. In the present study we analyzed the functional and phylogenetic diversity of amphibians in the Atlantic Forest in the present day and for 2080. Our results showed drastic reduction in species richness, ecosystem and evolutionary function at lower latitudes and low altitudes. The species tend to disperse to areas of high latitudes and high altitudes where temperatures are milder, tending to the same points that have been suggested as refuges during the Pleistocene. Our results present new refuges for the Atlantic forest that provide aid to making decisions regarding conservation strategies.

**Key words:** Altitude; Anthropocene; climatic change; ectothermics; Anura; Gymnophiona

### **A DIVERSIDADE FILOGENÉTICA E FUNCIONAL REVELAM NOVOS REFÚGIOS CLIMÁTICOS PARA ANFÍBIOS NA FLORESTA ATLÂNTICA BRASILEIRA**

#### **Resumo**

As mudanças climáticas têm ocorrido na Terra ao longo de milhões de anos. Para sobreviver a essas mudanças as espécies se refugiaram em locais onde as mudanças foram menos drásticas. A mudança climática é um panorama real e estratégias de conservação devem ser desenvolvidas. Espécies ectotérmicas, são especialmente sensíveis às mudanças do clima, o que as torna boas referências para estudos envolvendo alterações climáticas. Analisar as perdas ecossistêmicas e os valores evolutivos das espécies, pode ajudar a compreender os processos de extinção e minimizar o seu impacto. Analisou-se a diversidade funcional e filogenética dos anfíbios na Floresta Atlântica para o presente e para 2080. Os resultados mostraram redução drástica na riqueza de espécies, perda ecossistêmica e função evolutiva em latitudes e altitudes mais baixas. As espécies tendem a se dispersar para as áreas de altas latitudes e altitudes, onde as temperaturas são mais amenas, tendendo aos mesmos pontos que foram sugeridas como refúgios durante o Pleistoceno. Os resultados apresentam novos refúgios para a Floresta Atlântica e fornecem auxílio à tomada de decisões sobre estratégias de conservação no futuro.

**Palavras chaves:** Altitude; Antropoceno; mudanças climáticas; ectotermicos; Anura; Gymnophiona

### 3.1 INTRODUCTION

Over millions of years the Earth has undergone several transformations that seem to appear cyclically (Raup & Sepkoski, 1982). During these changes, the animals need to take refuge in areas with milder environments and resource availability (Haffer, 1969; Mayr & O'Hara, 1986; Bush, 1994; Bush & Oliveira, 2006; Carnaval *et al.*, 2009; Bush *et al.*, 2011). These events lead to massive extinctions (Raup & Sepkoski 1982; Bambach, 2006; Jablouski, 1994; Barnorsky *et al.*, 2011). The actual Age (Anthropocene) can to be the sixth massive species extinctions (Barnosky *et al.*, 2011; Dirzo *et al.*, 2014).

Anticipated the climate changes in biogeographic patterns its important because may also result in changes to functional and phylogenetic diversity and alterations to organism-mediated ecosystem goods and services (Montoya & Raffaelli, 2010; Cardinale *et al.*, 2011; Prather *et al.*, 2012), as well as evolutionary historical processes (Thuiller *et al.*, 2011; Pio *et al.*, 2014). Because of this, studies available about climatic change should be evaluated phylogenetic and functional diversity integrated (Sobral & Ciaccianruso, 2012). Thus, it is possible to have a broader view of historical and ecosystem reasoning processes. Despite the assumption that similar phylogenetically species has the same ecosystem role, it sometimes may not be true (Webb *et al.*, 2002). The phylogenetic structure of meetings depends on as the ecological characteristics evolved (Sobral & Ciaccianruso, 2012). The ecosystem functioning and stability are often correlated with evolutionary process changes, producing several implications for ecological and human wellbeing on short time scales (Alberti, 2015).

Climate change is a widespread threat to global biodiversity (Thomas *et al.*, 2004; Pereira *et al.*, 2010) by continuously promoting changes in physiological and ecological processes that directly affect the distribution and persistence of species in an environment (Stenseth *et al.*, 2002; MacDonald *et al.*, 2004; Huey *et al.*, 2009). Studies evaluating how climate changes affect the individual performance (Huang *et al.*, 2013; Holt & Jorgensen, 2015), demographic dynamics (Lukoscheck *et al.*, 2013; Pomara *et al.*, 2014), composition and species richness (Lemes & Loyola, 2013; Ferro *et al.*, 2014) are examples of the complex relationship of as has been addressed from diversity of perspectives of this process with the living organisms. Predictive outcomes have included adaptation to novel conditions (Quintero & Wiens, 2013), shift, expansion or retraction of ranges (Ferro *et al.*, 2014; Lemes *et al.*, 2014), isolation to unaffected areas or climatic refuges (Puschendorf *et al.*, 2009), and species extinctions (Thomas *et al.*, 2004). Such climate change effects are the reasons for which there is a growing consensus that management decisions for biodiversity conservation must be taken in light of this phenomenon (Araújo & Rahbek, 2007).

Ecological niche models (ENMs), also referred to as species distribution models (SDMs) (Peterson *et al.*, 2011; Rangel & Loyola, 2012), have been increasingly used to estimate species ranges for future scenarios of climate change (Peterson *et al.*, 2011). These models can be used to evaluate the current and future hotspots of phylogenetic (Thuiller *et al.*, 2011; Loyola *et al.*, 2013; Pio *et al.*, 2014) and functional diversity, acting as an efficient conservation tool (Del Toro *et al.*, 2015). Ectothermic animals are highly susceptible to climate change (Pounds *et al.*, 2006; Sinervo *et al.*, 2010) due to the interdependence of their behavioral-physiological functions in relation to external environment (Ribeiro *et al.*, 2012). Amphibians are sensible the environmental changes

(Lourenço-de-Moraes *et al.*, 2014) due their metabolic features (Duellman & Trueb, 1994) at high temperatures amphibians lose water to the atmosphere (Wells, 2007) and can suffer local extinction (Becker *et al.*, 2007). Because this, study using ENMs may be effective tools in predicting the possible dispersions than the climate change can promote for the amphibians species (Pie *et al.*, 2013; Ribeiro *et al.*, 2015) and enable advance shares to conservation.

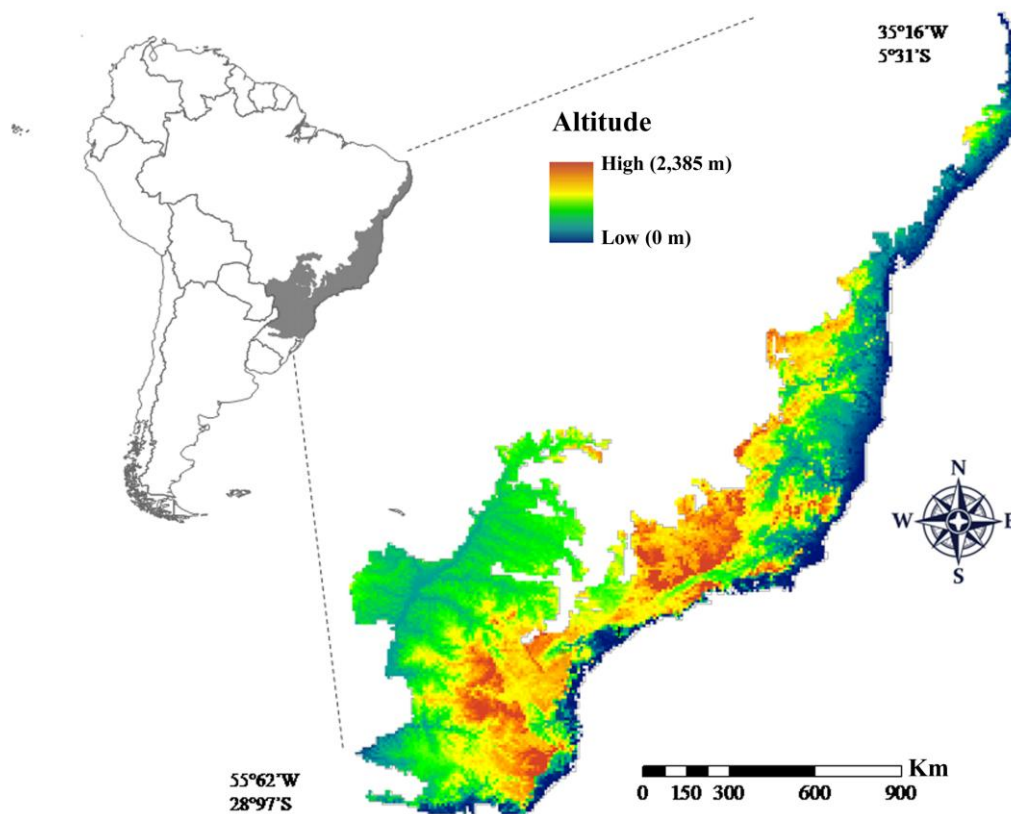
The consequences of human activities go beyond the loss of species and various studies reported losses of both evolutionary history and functional trait diversity at different landscapes (Purvis *et al.*, 2000; Flynn *et al.*, 2009; Mayfield *et al.*, 2010). The claimed Anthropocene age, follows drastic climate change (especially warmer temperature) causing that jointly a massive defaunation (Dirzo *et al.*, 2014). Recent studies have suggested that areas of altitude may be important refuges for vertebrates during the Anthropocene Age (Loyola *et al.*, 2013; Lemes *et al.*, 2014). In this context, we tested the hypothesis that the elevation areas are refuges for amphibians, and may maintain their ecosystem and evolutionary services. For this we analyzed the potential distribution, phylogenetic and functional diversity on present and future of Atlantic Forest species. We correlated this diversity with altitude in both periods and analyzed the retraction of the species in the direction to higher areas. In addition, we evaluated the species conservation status for 2080.



## 3.2 MATERIALS AND METHODS

### 3.2.1 *Study region*

Our analyses focused on the Atlantic Forest Biodiversity Hotspot (Myers *et al.*, 2000), which originally covered around 150 million ha with heterogeneous environmental conditions provided by a wide range of climatic belts and vegetation formations (Tabarelli *et al.*, 2005; Ribeiro *et al.*, 2009). This biome has an altitudinal range encompassing the elevational limits of the mountain chains of Serra do Mar and Serra da Mantiqueira (Cavarzere & Silveira, 2012) a longitudinal range harboring differences in forest composition due to a diminishing gradient in rainfall from coast to interior and a latitudinal range extending into the tropical and subtropical regions, (Ribeiro *et al.*, 2009) (Fig. 1).



**Figure 1.** Map showed the study area of Brazilian Atlantic Forest hotspot (in gray) in South American territory.

### 3.2.2 *Spatial species data*

We obtained spatial data of amphibian species from a joint research through four procedures types: (i) we built a dataset with all the species distributed in the Atlantic Forest according to Haddad *et al.* (2013); (ii) we included maps of geographical ranges for each species from the IUCN Red List of Threatened Species database (IUCN, 2015); (iii) we conducted a complementary fieldwork comprising the major Atlantic Forest remnants of Brazil, to supplement the dataset with observed functional traits, as body

size, reproductive mode, habitat, activity, poison patterns, habit and calling site (see Appendix 2 Figure S1); (iv) we modelled the potential distribution for the present and future using ecological niche modeling, and calculated the diversity functional and phylogenetic diversity based on species richness generated by the model for the current time and for 2080.

### 3.2.3 *Ecological niche modeling, species richness and turnover*

We used ArcGIS 10 software (ESRI, 2010) to build presence/absence matrices from the species distribution data by superimposing a grid system with cells of 0.1 latitude/longitude degrees, creating a network with 10,359 grid cells. In total, we assessed the geographical ranges 451 amphibian species (5 Gymnophionas and 446 Anurans) covered by our grid system.

Considering that species occurrence patterns are determined at large-scales by responses of organisms to different environmental conditions (reflecting the Grinnellian component of the ecological niche, *sensu* Soberón, 2007), we used ecological niche models (ENMs) to predict the distribution area of amphibians species in the Atlantic Forest. For this, we used the species occurrence matrix and the layers of climatic-environmental variables, resulting on a suitability matrix, which we used to modeling and mapping the potential distribution of each species evaluated.

We used the following bioclimatic variables in the modeling process: (i) annual mean temperature, (ii) annual temperature range, (iii) precipitation of wettest month, (iv) precipitation of driest month and (v) precipitation of warmest quarter. We obtained these variables for the present and future (mean of simulations for 2080-2100) from.

CMIP5 – Coupled Models Intercomparison Project Phase 5 (<http://cmip-pcmdi.llnl.gov/cmip5/>; and also at <http://ecoclimate.org>, see Lima-Ribeiro, 2015), and downscaled to the resolution of  $0.05^\circ$ . We also used altitude as predictor of richness and dispersion, through the data available from the WorldClim Global Climate Data ([www.worldclim.org](http://www.worldclim.org)). We assumed the temporal stationary of this variable to perform future predictions. For future, we used the greenhouse gas concentration trajectory corresponded to the Representative Concentration Pathway (RCP) 4.5, that represents a moderated emission scenario within a optimistic context. We used simulations provided by four Atmosphere-Ocean General Circulation Models (AOGCMs): CCSM, CNRM, MIROC and MRI, which were obtained from CMIP5 (Coupled Model Intercomparison Project – Phase 5). Original data resolution varied from  $1^\circ$  to  $2.8^\circ$  (in longitude and latitude) and both current and future climate variables were rescaled to fit our grid resolution.

We performed four conceptually and statistically different ENMs based on presence data (i.e. only occurrences are known, absences are unknown – pseudo-absence), using the algorithms (i) Bioclim (BIO, Busby, 1991) based on bioclimatic envelope logic; (ii) Gower Distance and Euclidean Distance (GD, EUD, Carpenter *et al.*, 1993) based on environmental distance approach; (iii) Maximum Entropy (ME, Phillips *et al.*, 2006) and random forest (RF, Breiman 2001) based on machine learning technique; and (iv) Ecological Niche Factor Analysis (ENFA, Hirzel *et al.*, 2002) based on multivariate analysis, and Genetic Algorithm for Rule set Production (GARP, Stockwell & Noble, 1992). Given the particularities of each model, different predictions have been provided, generating uncertainties about which model is more appropriate to represent the geographical distribution of species (Diniz-Filho *et al.*, 2009). To

overcome this uncertainty and minimize errors, we employed the ensemble forecasting approach, which offers a consensus of multiple models (Araújo & New, 2006). The main idea of ensemble forecasting is that different sources of errors will affect each niche model in different ways and, by obtaining a consensus result of these models, errors will tend to cancel each other out and produce a trustworthy and more conservative solution (Diniz-Filho *et al.*, 2010a). Assuming that the consensual model (CONS) reduces uncertainty and error associated with alternative ENMs, we interpreted only the range sizes from the CONS model.

We randomly partitioned presence and pseudo-absence (in the case of Maxent) data of each species into 75% for calibration (or training) and 25% for evaluation (or test); repeating this process 10 times by cross-validation for all models. For each ENM, we converted the continuous predictions of suitability into a binary vector of 1/0 (presence and absence in each cell), finding the threshold that maximizes sensitivity and specificity values in the receiver operating characteristic (ROC). The ROC curve is generated by plotting the fraction of true positives vs. the fraction of false positives at various threshold settings. The distribution areas were estimated obtaining 280 predictions (7 models x 10 randomizations x 4 AOGCMs) for each species and time-period of climatic conditions (i.e. present and future). This allowed us to generate a frequency of projections in the ensemble. Then, we generated the frequency of projections weighted by the total sum of squares (TSS) statistics for the present and future (the best models according to this metric have more weight in our consensus projections). The TSS range from -1 to +1, where values equal to +1 is a perfect prediction and values equal to or less than zero is a prediction no better than random (Allouche *et al.*, 2006). We considered the species present only in cells where at least

50% of models retained in the ensemble point out the species as present. In our analyses we obtained the CONS for each AOGCM and time period (present and 2080). Thus, we obtained the final maps of richness for present, future and turnover through the average of values projected by CONS for each grid cell – considering the different GCMs. We run all models using the computational platform Bioensembles (Diniz-Filho *et al.*, 2009), and mapped results using the software SAM v.4.0 (Rangel *et al.*, 2010). To determine the species patterns of amphibians of Atlantic Forest, we employed the modeling strategy at the community level of “predict first, assemble later” (sensu Overton *et al.*, 2002), where the ranges of individual species are modeled one at a time as a function of environmental predictors and then overlapped for obtaining the species richness.

In addition, we used principal components analyses (PCA) (Legendre & Legendre, 2012) to compare species richness patterns derived from alternative ENMs and their consensus. This analyses allowed us to evaluate the degree of which different ENMs converge in estimates of regional species richness and determine which model reflects the main direction of variation among richness maps (Diniz-Filho *et al.*, 2010b). Thus, in our study, we interpreted only the results of the model reflected by the main direction of variation among suitability maps. We calculated species turnover between current and future species distributions in each cell according to formula  $(G+L/S)/S+G$  (Thuiller, 2005), where “G” refers to the number of species gained, “L” the number of species lost and “S” the contemporary species richness found in the cell.

### 3.2.4 Functional Diversity and Phylogenetic Diversity

We used the following traits according to Haddad *et al.* (2013), and added complements from our fieldwork: (i) body size; (ii) appendices (apodal and tetrapod); (iii) activity (nocturnal, diurnal, and both); (iv) poison (poisonous, unpalatable or bad odor, and no poison); (v) habitat (forested area, open area, and both); (vi) habit (arboreal, phytotelmate, terrestrial, cryptozoic, fossorial, rheophilic, semi-aquatic, and aquatic); (vii) calling site (bamboo grove, swamp or lake, bromeliad, forest floor, tree canopy, caves or burrows, rock wall, backwater river, stream, river, shrubs, grasslands and not sings); and (viii) reproductive mode (1 to 39 modes; see Haddad & Prado, 2005). See Appendix 2 Table S2 for details.

We followed the protocol proposed by Petchey & Gaston (2006) to calculate the functional diversity (FD): (i) construction of a species-trait matrix; (ii) conversion of species-trait matrix into a distance matrix; (iii) clustering distance matrix into a dendrogram (UPGMA); and (iv) calculating functional diversity by summing dendrogram branch lengths of species community. To create the distance matrices, we used the method proposed by Pavoine *et al.* (2009), through the Gower distance.

We used the phylogenetic diversity index (Faith, 1992) to quantify the phylogenetic diversity (PD), which comprises the sum of the lengths of the branches lengths of the phylogenetic tree of all species assessed and is often used in the assessment of phylogenetic diversity of co-occurrent species (e.g. Rodrigues & Gaston, 2002; Safi *et al.*, 2011; Trindade-Filho *et al.*, 2012). The PD index has appropriate ways of accounting for relatedness between taxa and evolutionary history in a conservation context (Pio *et al.*, 2011).

We based the phylogenetic distance through 208 species nucleotide sequences obtained from the GenBank (Benson *et al.*, 2013), provided by the National Center for Biotechnology Information (NCBI) (See Appendix 2, Table S3). Following the protocol proposed by Pyron and Wiens (2011) in an extant amphibian phylogeny, we used 12 genes to produce a novel estimate phylogeny for the Atlantic Forest amphibians (i.e. 11,906 bp for each species), through three mitochondrial genes were included: cytochrome b (cyt-b), and the large and small sub-units of the mitochondrial ribosome genes (12S/16S); and nine nuclear genes: C-X-C chemokine receptor type 4 (CXCR4), histone 3a (H3A), sodium–calcium exchanger (NCX1), pro-opiomelanocortin (POMC), recombination-activating gene 1 (RAG1), rhodopsin (RHOD), seventh-in-absentia (SIA), solute-carrier family 8 (SLC8A3), and tyrosinase (TYR). For the length-variable regions, we performed multiple pairwise comparisons by the online version of MAFFT v.6.8 and the G-INS-i algorithm (Katoh & Toh, 2008). After, we put together alignments of all genes in the same alignment using the software SequenceMatrix 1.7.7 (Vaidya *et al.*, 2011) to concatenate the supermatrix previously produced.

We conducted the phylogenetic relationships with Bayesian analyses in software BEAST 1.8 (Drummond & Rambaut, 2007), estimating the node supports with a fast bootstrapping mode. We performed the Bayesian analyses on the combined data matrix using a HKY model of sequence evolution for one partition for all genes, under a Yule speciation process as the tree prior and an uncorrelated relaxed clock. After removal of the burn-in, we run the Yule process for 100 million generations, ensuring that the number of generations after convergence were sufficient assessed with Tracer v1.6 (Drummond & Rambaut, 2007), combining the results with the use of Logcombiner 1.8 and Treeanotator 1.8 (Drummond & Rambaut, 2007). We considered the nodes strongly



supported if they received probability (pp) support values  $\geq 0.95$ . To solve the polytomies, we used the ‘multi2di’ function of the package ‘ape’ (Paradis, 2004) in the R software (R Development Core Team, 2010). Thus, we reconstructed a new phylogenetic tree using the Mesquite software, version 3.0 (Maddison & Maddison, 2015).

The functional diversity (FD) and phylogenetic diversity (PD) indices are often correlated with the species richness (SR) values (Devicter *et al.*, 2010). In order to overcome this problem, we used null models (Swenson, 2014) for current and future times. Hence, we tested if the functional and phylogenetic diversity were higher, equal or lower than expected by chance for each grid cell (random or non-random pattern), assuming a random distribution in which every species could occupy any grid cell in the biome. For each pruning event (present and future), we computed 1,000 replicates of random remaining PD and FD, allowing us to obtain a *P*-value of predicted PD and FD as compared to the distribution of the random replicates. All analyses were performed using the packages “ade4”, “picante”, “FD” and “vegan” through the R software (R Development Core Team, 2009).

### *3.2.5 Species Richness, Functional Diversity and Phylogenetic Diversity vs. Topography and Spatial References*

We used correlation matrices to compare the topographic patterns and spatial references (altitude and latitude) to the values obtained by the consensus model for SR, Turnover, FD and PD in each grid cell for current and future time (2080). Thus, we

correlated the values obtained for SR, FD and PD with altitude, and the turnover values with altitude and latitude, using simple linear regression models.

### 3.2.6 *Threat status of species facing the climate change*

From the individual range sizes (i.e. number of occupied cells) of each species in current and future time, we estimated the threat status of amphibian species in the Atlantic Forest at 2080, according to the Criterion B established by IUCN Red List (2015) and Maiorano *et al.*, (2011). We considered the following threat categories: (i) Extinct (EX), the species whose loss of distribution area is estimated at 100% for the next 50 or 80 years; (ii) Critically Endangered (CR), the species whose loss is estimated at 80% for the projected time interval; (iii) Endangered (EN), the species whose loss is estimated at 50% for the projected time interval; (iv) Vulnerable (VU), the species whose loss is estimated at 30% for the projected time interval; and (v) Least Concern (LC), the species whose loss is estimated at below 30% for the projected time interval. It is noteworthy that this approach only considers the effects of climate change on species distribution, thus providing a synthetic view of the specific threats imposed on the species by the alteration in climatic conditions.

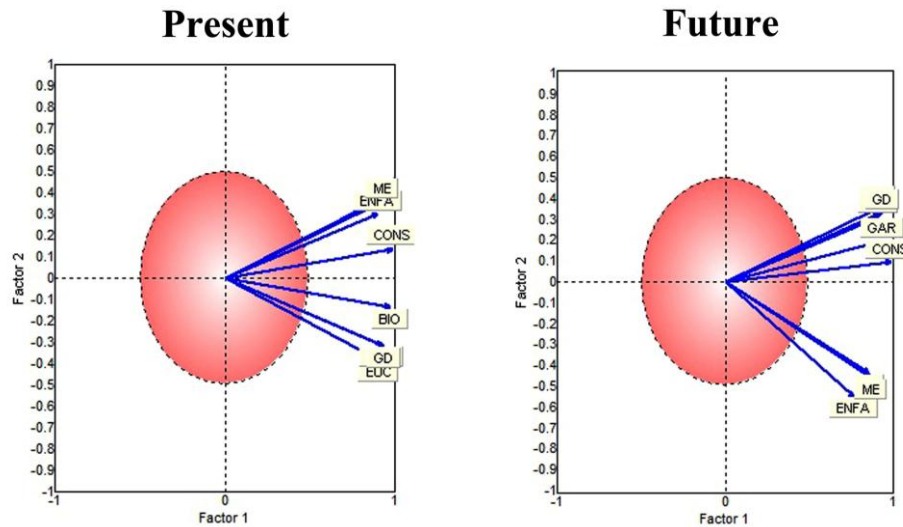
## 3.3 RESULTS

The TSS values for most species was  $0.61 \pm 0.11$ , indicating relatively high good fit model (Eskildsen *et al.*, 2013). The PCA axes explained a large proportion of variation between species richness maps generated by different ENMs. The accumulated proportion of variation explained by the two axes ranged from 97.1% to current time

and 95.8% for species in 2080 (Table 1). Similar results were produced in both time periods by GD and EUD (Fig. 2). The predictions of ENFA produced low values: similar results between GD and GARP for the future. The values showed by BIO were similar to CONS values in the current time. However, the CONS model had the highest loading for the first PCA axis, reflecting the main direction of variation among suitability maps (Table 1). Because of this, only the outputs derived from the consensus of multiple ENMs (CONS) were retained for interpretation.

**Table 1.** Values of PCA loadings of the different ecological niche models and consensus for all amphibians species in the current time and future.

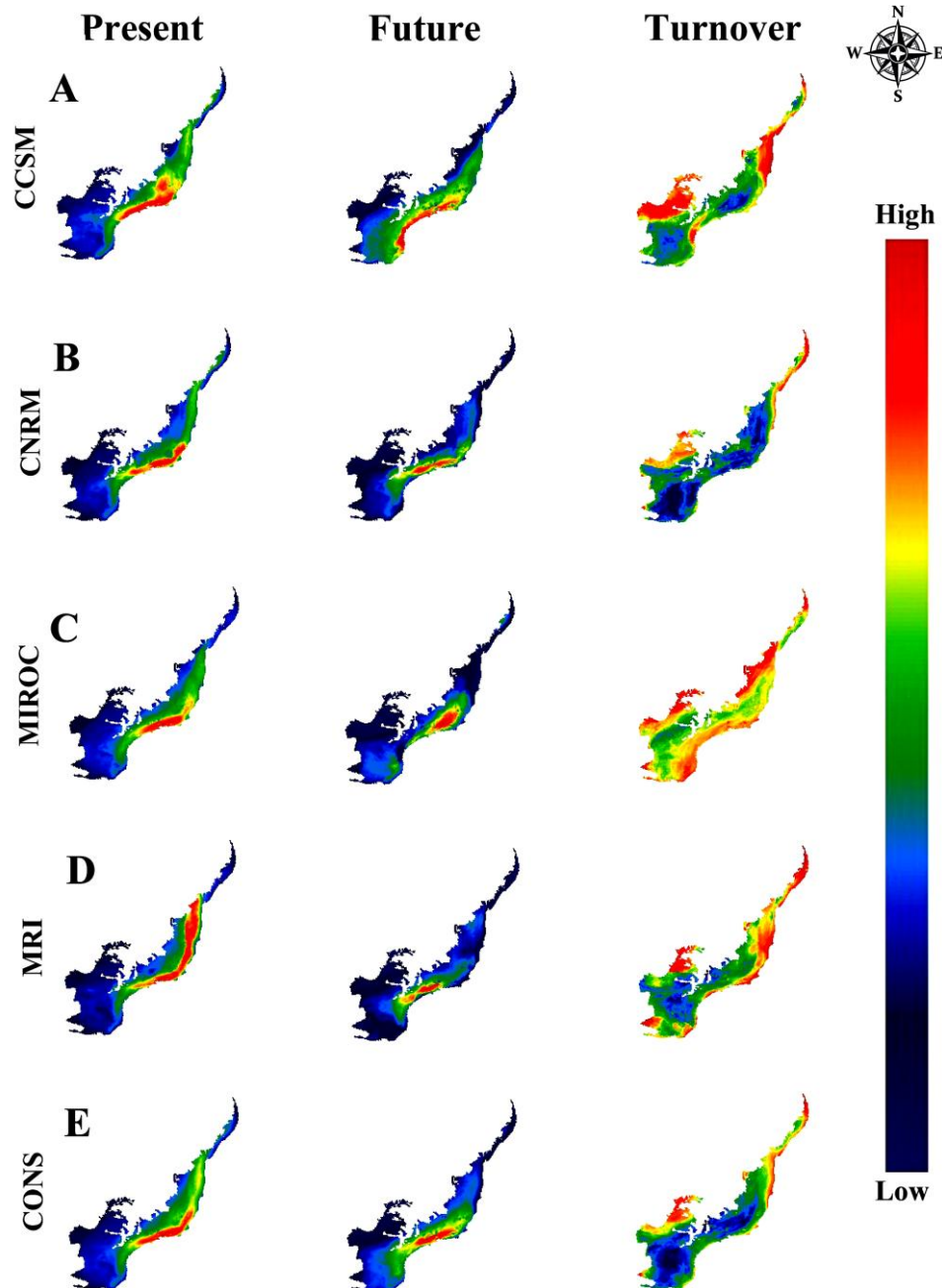
Ecological Niche Models		Present		Future	
		PC1	PC2	PC1	PC2
Distance Methods	ENFA	0.894	0.298	0.767	-0.548
	GD	0.937	-0.319	0.930	0.332
	EUD	0.902	-0.380	0.909	0.311
Machine Learning	BIO	0.972	-0.134	0.906	0.351
	ME	0.924	0.356	0.867	-0.457
	RF	0.906	0.365	0.856	-0.433
	GAR	0.918	-0.314	0.936	0.102
	CONS	<b>0.983</b>	0.137	<b>0.986</b>	0.095
	Axes explanation (%)	0.865	0.091	0.804	0.134
	Accumulated explanation (%)	0.971		0.958	



**Figure 2.** Loadings of the first two axes of the PCA performed considering the amphibians species richness predictions provided by different ENMs (ENFA = Ecological Niche Factorial Analysis; GD = Gower distance; EUD = Euclidean Distance; BIO = Bioclim; ME = Maximum Entropy; RF = Random Forest; GAR = Genetic Algorithm for Rule set Production and CONS = Consensus Model) for amphibians in the current (A) and future (B) times.

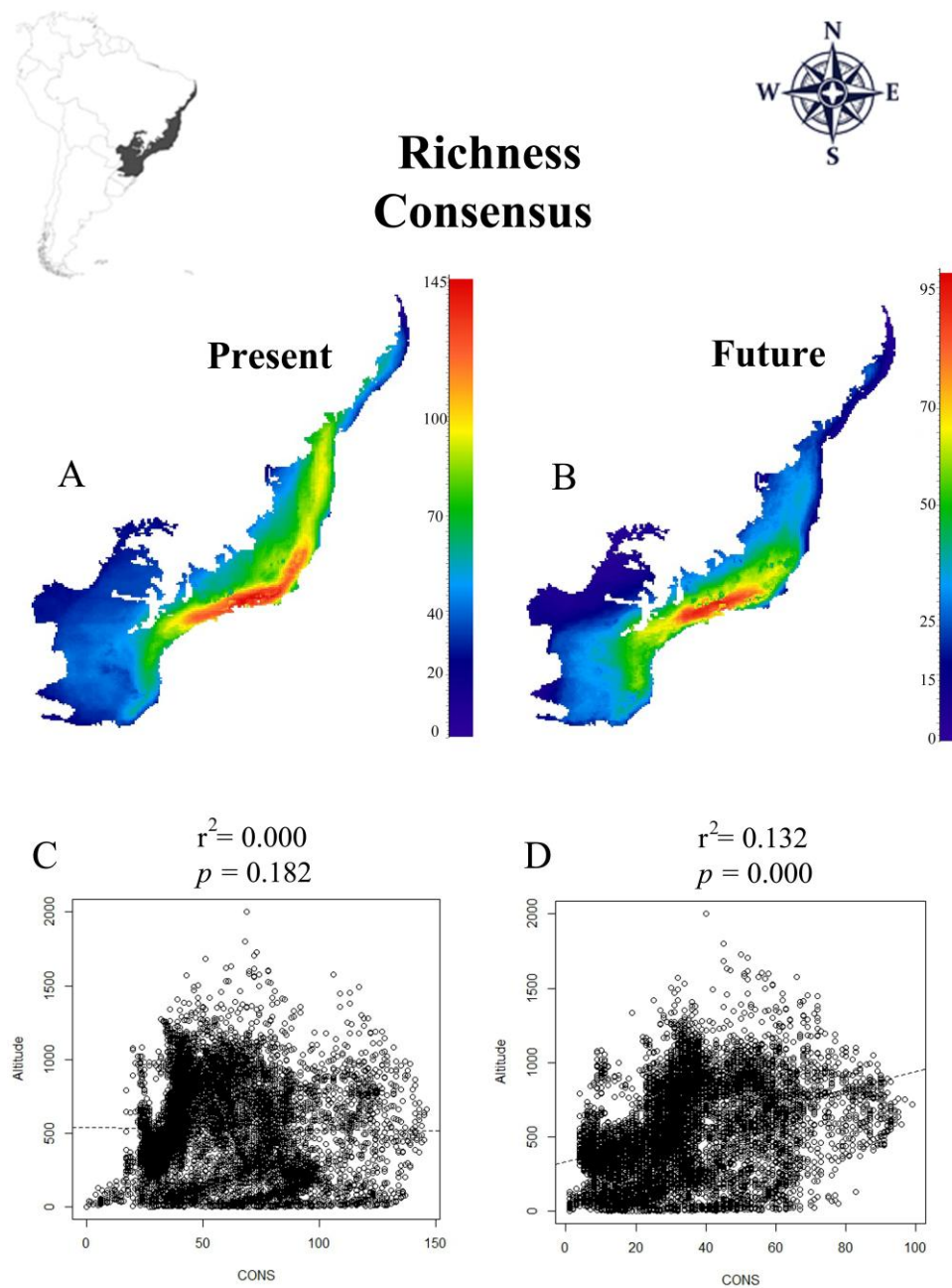
The overlap of individual species ranges generated by the CONS model evidenced that highest species richness values were restricted for almost all eastern-central portion of the Atlantic Forest in the current time (Fig. 3 and 4). The species richness pattern showed no significant relationships with the altitude ( $r = 0.000$ ,  $p = 0.182$ ) (Fig. 4c). The future predictions produced by CONS from different AOGCMs pointed out the losses of climatically suitable areas in this region by 2080, with the species richness directed to the east-central portion of the Atlantic Forest. In this case, the species richness increases toward higher altitude ( $r = 0.132$ ,  $p < 0.001$  Fig. 4d). In general, CCSM and MRI showed two distinct species-rich areas and CNRM and MIROC produced more homogeneous results, with the latter more restrictive (Fig. 3a-d). By combining the results of AOGCMs in a full ensemble model, we found proof that

in the future the species richness peaks will be restricted to a reduced portion of the central-eastern region of the biome, in locations closer to the mountains region of the Atlantic Forest (Fig. 2).



**Figure 3.** Species richness and turnover derived from different Global Circulation Models (CCSM - A, CNRM - B, MIROC - C and MRI - D) and consensus model (CONS - E) for amphibian species in the current time and for 2080.

Consequently, by combining the results of AOGCMs in a full ensemble model, we observed that the temporal turnover was high. Thus changes in the species composition are predicted to be greater on the western edge and on the northeastern edge of the biome. We found significantly higher turnover rate from high to low latitudes ( $r = 0.308, p < 0.001$ ) and areas from low to high altitudes ( $r = 0.307, p < 0.001$ ) (see Fig. 5a-c).

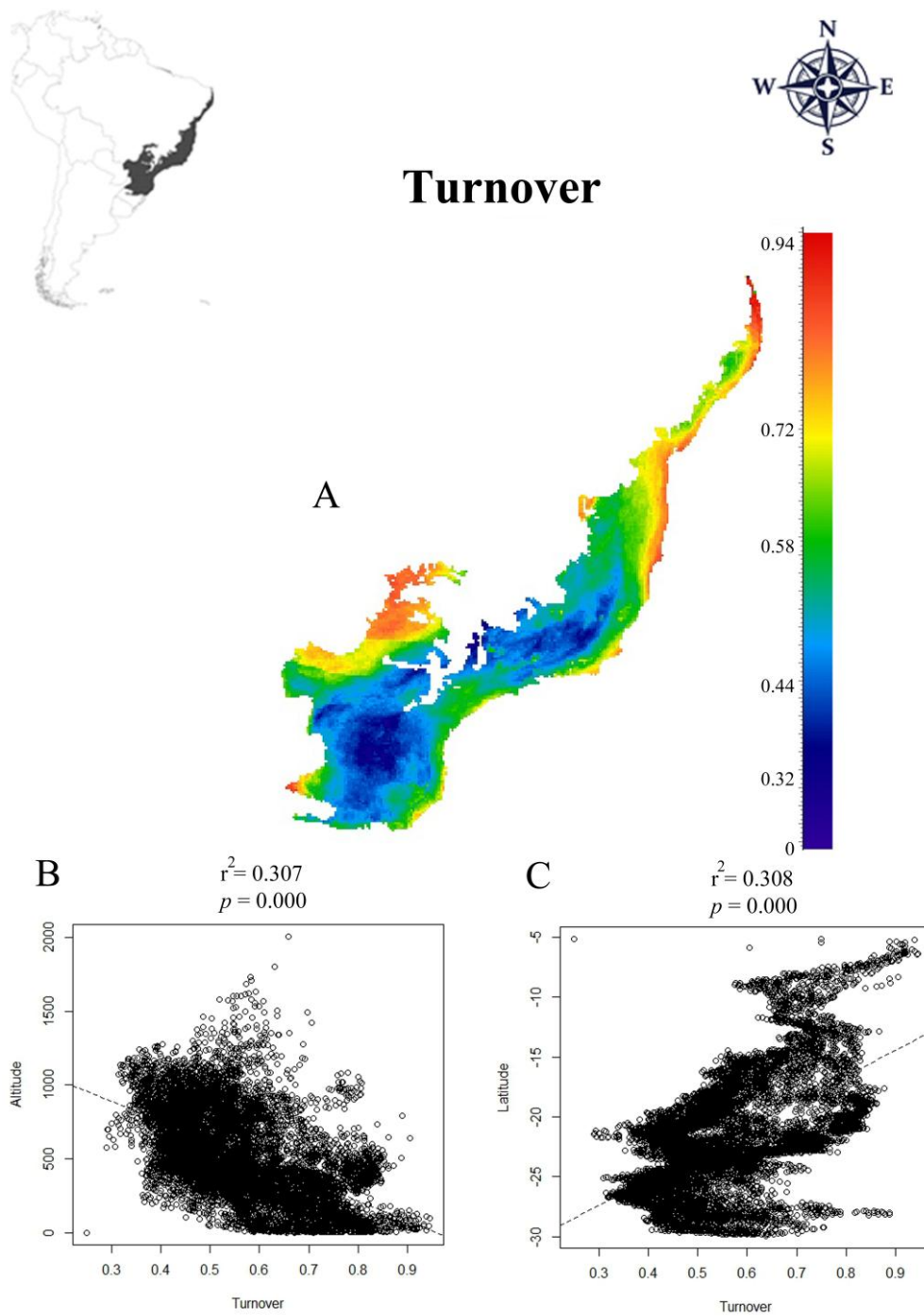


**Figure 4.** Species richness derived from consensus model Map for current time (A) and for 2080 (B). Relation between amphibians species richness consensus model (CONS) and altitude for current time (C) and for 2080 (D)

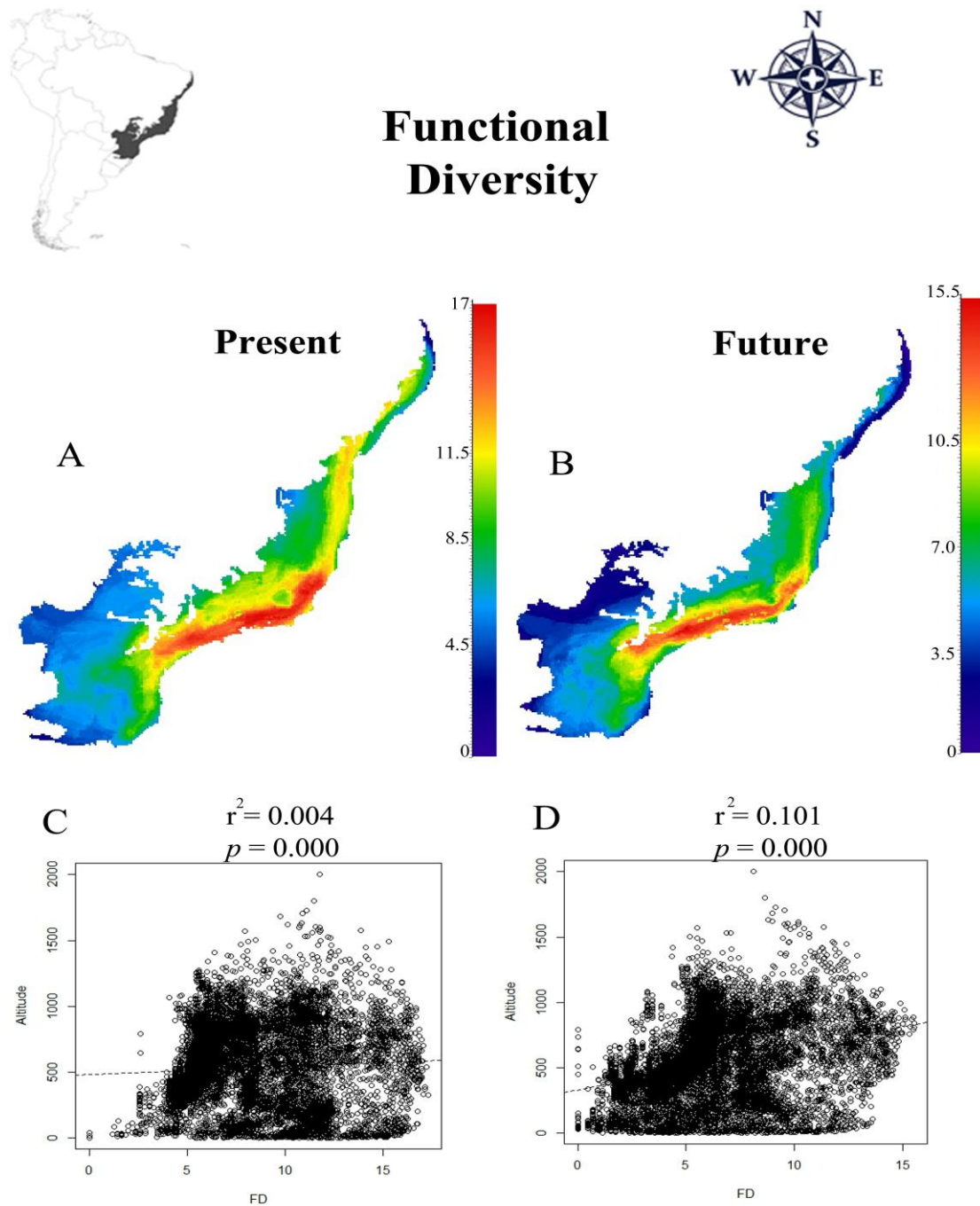
Our results showed high functional diversity (FD) in the regions of the eastern Atlantic Forest with the highest rates in the east-central region rising to the northeast in the current time (Fig. 6a, c). In 2080, these values will decrease from 17.296 to 15.532 at its maximum value (Fig. 6b), and will have a significantly higher loss in areas of lower altitudes ( $r = 0.101$ ,  $p < 0.001$ , Fig. 6d). High rates were found mainly in the south of Bahia to the south of São Paulo states.

The highest values of phylogenetic diversity (PD) in the current time are distributed in the in the east-central region, mainly in the region of Serra do Mar rising to Central Corridor and a small part of Pernambuco (in high altitude areas) (Fig. 7a, c). For 2080, these values will decrease from 5.752 to 5.440 at its maximum value (Fig. 7b), which will dramatically decrease in the south and north of Bahia, increasing towards the south of the Serra do Mar portion. The regions of highest altitude will be replaced by significantly values in the future ( $r = 0.121$ ,  $p < 0.001$ ) (Fig. 7d). The built null models for the FD and PD in current and future time showed different values than expected by chance ( $p < 0.001$ ), indicating a pattern of non-random of functional and phylogenetic diversity.

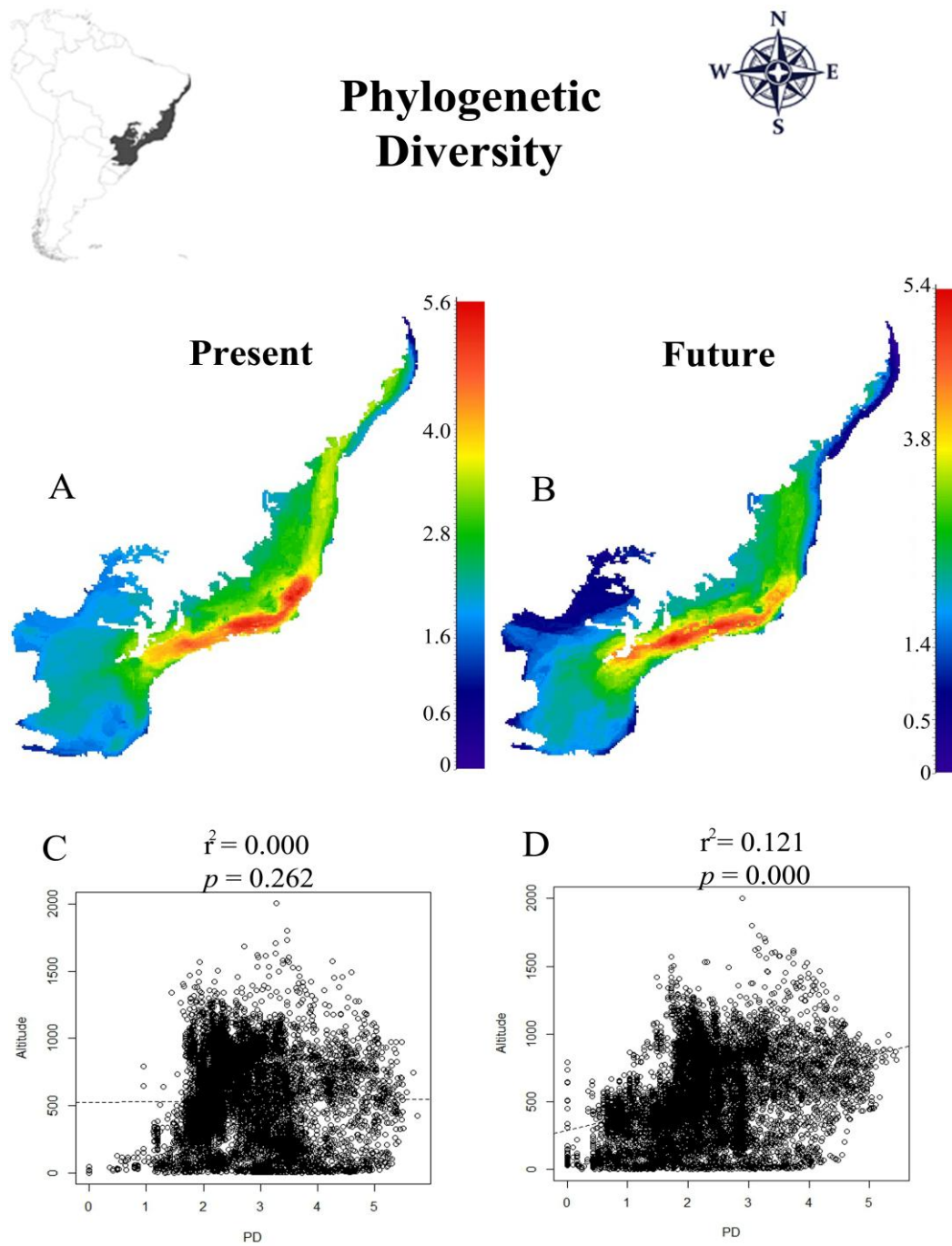




**Figure 5.** Turnover derived from consensus model Map (A). Relation between turnover consensus model and altitude (B) and for latitude (C) for amphibians species.

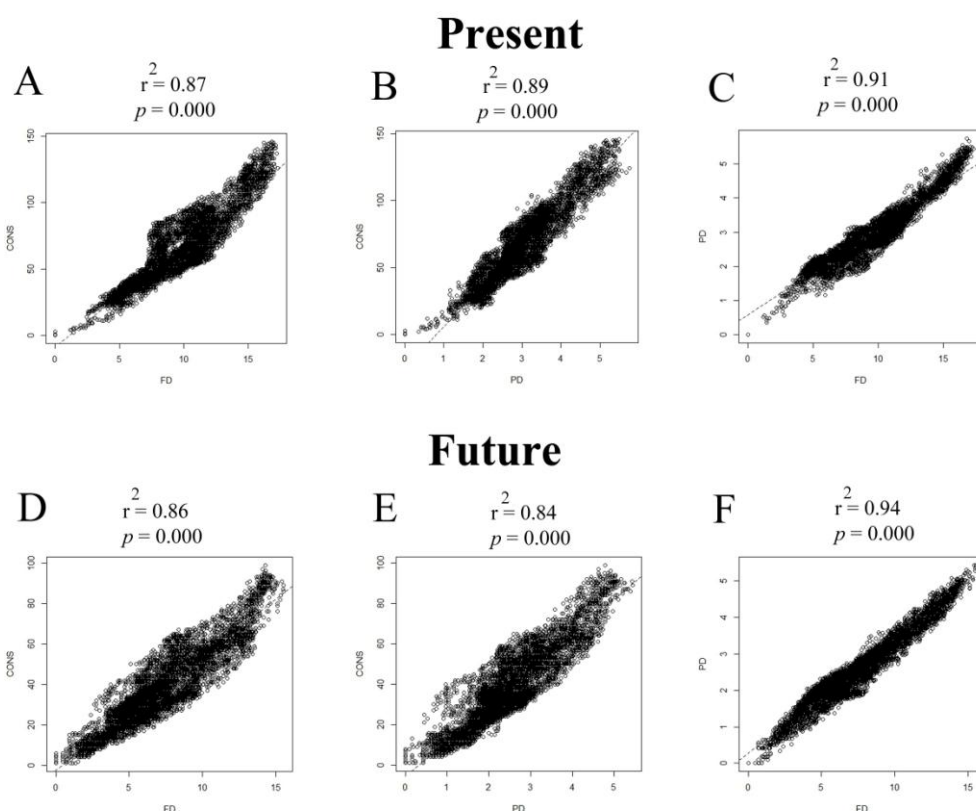


**Figure 6.** Spatial distribution of functional diversity (FD) derived from amphibians species richness consensus model Map for current time (A) and for 2080 (B). Relation between functional diversity (FD) consensus model and altitude for current time (C) and for 2080 (D).



**Figure 7.** Spatial distribution of phylogenetic diversity (PD) derived from amphibians species richness consensus model Map for current time (A) and for 2080 (B). Relation between phylogenetic diversity (PD) consensus model and altitude for current time (C) and for 2080 (D).

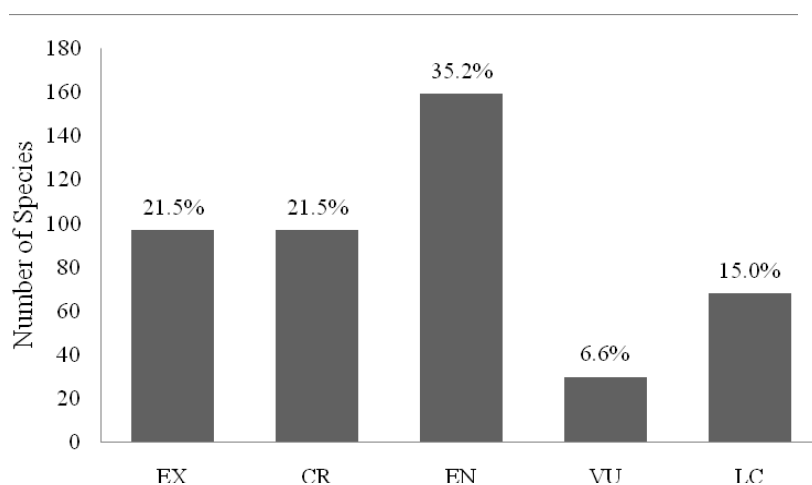
Our results pointed that FD and PD were highly correlated (present:  $r = 0.91$ ,  $p < 0.001$ ; Future:  $r = 0.94$ ,  $p < 0.001$ ), which was also observed when compared with the SR consensus model (present: FD –  $r = 0.87$ ,  $p < 0.001$ , PD –  $r = 0.89$ ,  $p < 0.001$ ; future: FD –  $r = 0.86$ ,  $p < 0.001$ , PD –  $r = 0.84$ ,  $p < 0.001$ ). Therefore, the loss of species richness in the future may be accompanied by the loss of FD and PD (Fig. 8).



**Figure 8.** Relationships between Consensus model of richness species (CONS), Functional diversity (FD), Phylogenetic diversity (PD) of amphibians in the Brazilian Atlantic Forest. Present time: A (CONS vs FD), B (CONS vs PD) and C (FD vs. PD); Future time: D (CONS vs FD), E (CONS vs PD) and F (FD vs PD).

Our simulations revealed that due to the massive habitat suitability losses facing climate change, the vast majority of amphibians of the Atlantic Forest (62.0%) will be subject to some level of threat by 2080 (Fig. 9). Considering the projections for a moderate carbon emission scenario, our results showed that only 15% (N= 62 spp) of the species should present a low risk of extinction (i.e. less than 30% reduction from the original distribution area), given the restrictions imposed by future climatic conditions.

However, from these 15% of the species, 3.5 % even with an increase in its distribution still get a low ranges. Thus, our results also indicated that 84.9% (N= 383 spp) of the species will be threatened, and 21.5 % (N= 97 spp) of the species are expected to be extinct from the biome by 2080. The summary of the impacts of future climatic alterations on each individual species is given in Appendix 2, Table S1.



**Figure 9.** Status of threat project to 2080 in the face of climate change for amphibians of Atlantic Forest hotspot. EX = Extinct; CR = Critically Endangered; EN = Endangered; VU = Vulnerable; and LC = Least Concern.

### 3.4 DISCUSSION

Our data showed a dramatic reduction of species range (SR, PD and FD) for 2080. High rates of turnover in the lower latitudes and lower rates in the altitudes can strengthen the mountains of Atlantic Forest as climatic refuges (Randin *et al.*, 2009; Araújo *et al.*, 2011). The persistence of the species in their original ranges will be dependent on their degree of physiological and phenotypic plasticity, antipredator mechanisms, reproductive mode, or evolutionary adaptation of each species to changing environmental conditions, as well as the combination of these processes (Holt, 1990; Visser, 2008; Toledo *et al.*, 2012; Urban *et al.*, 2014; Ferreira *et al.*, 2016; Lourenço-de-Moraes *et al.*, 2016). As a result, some species have increased their ranges; although even features a low range in the future (lower 30%). For instance, opportunistic species adapted to warmer and drier environments also had increase of range (e.g. *Rhinella crucifer*, *Dendropsophus braneri*, *Leptodactylus troglodytes* and *Siphonops annulatus*) (Haddad *et al.*, 2013; present study). However, species that live in high altitudes and high latitudes have their ranges little affected. *Brachycephalus brunneus*, a species that due to their morphology and great dependence on abiotic factors (i.e. temperature and rainfall – 1,300 m elev.) (Ribeiro *et al.*, 2005), had low range changes in our model, which can enable this species to remain in the future. On the other hand, species from this genus that live more to north of Atlantic Forest and with similar features as *B. pulex* (Napoli *et al.*, 2011) may be extinct according to the same model.

Our results showed that FD and PD have high correlation among themselves and a nonrandom pattern of species composition for both present and future times. This pattern suggests that environmental factors may act as filters that does not allow the coexistence of similar species (Diamond, 1975; Weiher & Keddy, 1999). In addition,

historical factors provided biogeographic barriers acting to determine the regional bank of some species assemblages (Ricklefs & Schluter, 1993). Our results revealed three major areas of FD and PD for the current time, which are located in the mountains of the Serra do Mar, the Central Corridor of the Atlantic Forest and the altitude areas in the Pernambuco state. Carnaval *et al.* (2009) pointed out three climatic refuges for amphibians during the Pleistocene: (i) Southern Bahia state (located in the Central Corridor of the Atlantic Forest); (ii) Pernambuco state; and (iii) east-north region of São Paulo (i.e. Serra do Mar). Our results also showed higher rates of phylogenetic and functional diversity in these areas, supporting Carnaval *et al.* (2009) hypothesis. Moreover, our data also suggest climatic refuges in the Espírito Santo state, the Serra da Mantiqueira region that corresponds to the South of Minas Gerais state, and the South of Serra do Mar that corresponds to the east of Paraná state. In the future (i.e. 2080), these same areas will continue as climatic refuges, but changing the scenery for the areas of higher altitudes as new climatic refuges. Our model clearly showed the species will disperse from lower latitudes to higher latitudes and altitudes (Fig. 2, 3). Species with access to mountainous regions may migrate to highest altitudes areas, which have lower temperatures (Colwell *et al.*, 2008), and in the case of the Atlantic Forest, should retain greater humidity by better-preserved forests cover (Ribeiro *et al.*, 2009).

Loyola *et al.* (2013) indicated high values of phylogenetic diversity in the Atlantic Forest for the future (i.e. 2080). Our data contradict these values, indicating low PD values and significant correlations with low SR consensus model for the future. Nevertheless, our results corroborate the data obtained by Thuiller *et al.* (2011), which showed that the loss of species richness may be accompanied by the loss of

phylogenetic diversity. Furthermore, our data also indicates that the loss of species richness for the future will follow the loss of functional diversity.

The historical loss and their relationships do not reflect the actual proportion of loss in relation to the conservation of species. Moreover, FD reflected better the lost of ecological functions and evolutionary perspectives, because similar species from similar phylogenetic clades may not coexist due to a possible competitive exclusion (Diamond, 1975). However, species phylogenetically similar may have different effects on the ecosystem (Webb *et al.*, 2002; Gomez *et al.*, 2010). In these cases, the loss of phylogenetically similar species may coexist and may have greater losses in processes and ecosystem services.

Our results showed high species extinction rate in 2080 (21.5%) and other 63.3% species will be threatened. The Atlantic Forest was severely destroyed and fragmented, resulting on 9% remaining of its original formation (Ribeiro *et al.*, 2009). It is possible that these values can be even greater. According to our results, 84.9% of the studied species will be endangered or extinct by 2080. It is highly concerning that 94% of the species of our study are not included in any threatened status by the Brazilian Red list (Appendix 2, Table S1).

Studies in the Atlantic Forest has been warning about the need to invest in Protected Areas at altitude areas (Lemes *et al.*, 2013; Loyola *et al.*, 2013), mainly in the areas of Serra do Mar and south of Bahia (Carnaval *et al.*, 2009). Therefore, considering the dramatic evolutionary and ecological loss showed in this study, we suggest four large areas as priority for conservation efforts, the Serra do Mar, the Serra da



Mantiqueira, the Central Corridor and areas of northeastern altitude of the Atlantic Forest.

### 3.5 CONCLUSION

Many biotic and abiotic factors can influence the richness and composition of species in an environment (Diamond, 1975). Amphibians are particularly sensitive to environmental changes, and depending on the species, the dispersion may be difficult because of its specializations as diminutive size (Crump, 2010; Early & Sax, 2011; Lourenço-de-Moraes *et al.*, 2012, 2014). The use of various algorithms to reach a better consensus have been demonstrated as an effective strategy to reach outcomes closer to reality, and is one of the keys to understanding how communities can respond to climate changes (Araújo & New, 2006; Marmion *et al.*, 2009). Therefore, our consensus model showed consistent results and reflected the main direction in relation to species richness, as well as their refuges in the Antropocene age, ensured their evolutionary and ecosystem values.

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#### **4 HISTÓRIA EVOLUCIONÁRIA DOS MECANISMOS ANTIPREDAÇÃO DOS ANFÍBIOS DA FLORESTA ATLÂNTICA BRASILEIRA**

##### **Resumo**

Compreender os mecanismos de antipredação é uma das principais questões para entender a diversidade e dispersão das espécies. A origem dos mecanismos de antipredação pode ter origem evolucionária (de forma aleatória determinada geneticamente sem controle), por pressões ecológicas/etológicas (fatores bióticos e abióticos que levam o animal a aprender e repetir um determinado comportamento) ou pela união de ambos. Os anfíbios possuem alta plasticidade adaptativa e sofrem forte pressão ambiental, principalmente em relação ao seus diversos predadores. Aqui, nós investigamos a origem dos mecanismos de antipredação de 115 espécies de anfíbios da Floresta Atlântica. Analisamos as suas origens evolutivas com base nas características genéticas e suas relações entre a diversidade de mecanismos e características ecológicas funcionais. Os resultados mostraram que muitos mecanismos têm origem de forma independente em diferentes famílias e subfamílias. O mecanismo de antipredação de produzir secreções é puramente filogenético e segue um modelo de evolução Browniano e pode dirigir o ajustamento ou desenvolvimento de novos mecanismos. Características funcionais que expõem anfíbios suscetíveis à predação, agem como uma forte pressão evolutiva e em resposta as espécies tendem a ter uma maior diversidade de mecanismos, que estão em vantagem por sua vez, e aumentam a sua distribuição geográfica. Os resultados apresentam novas perspectivas e orientações para o sucesso da diversidade de anfíbios na Floresta Atlântica brasileira.

**Palavras chaves:** características funcionais; dispersão; homologia; sinal filogenético.

#### **EVOLUTIONARY HISTORY OF AMPHIBIANS ANTIPREDATION MECHANISMS OF THE BRAZILIAN ATLANTIC FOREST**

##### **Abstract**

Understanding antipredation mechanisms is one of the key issues to understand the diversity and dispersal of species. The origin of antipredator mechanisms may be evolutionary (randomly, genetically determined without control), for ecological/ethological pressures (biotic and abiotic factors that cause the animal to learn and repeat a certain behavior) or the union of both. Amphibians have high adaptive plasticity and suffering strong environmental pressure, especially in relation to their various predators. Here, we investigate the origin of antipredator mechanisms of 115 amphibians species of the Atlantic Forest. We analyze their evolutionary origins based on genetic characteristics and relationships between the diversity of mechanisms and functional ecological characteristics. Our results showed that many mechanisms originate independently in different families and subfamilies. The antipredator mechanism to produce secretions is purely phylogenetic and follows a model of Brownian evolution and directs the adjustment or development of new mechanisms. Functional traits that exposes amphibians susceptible to predation, acts as a strong evolutionary pressure is in response to species tend to have a greater diversity of antipredator mechanisms and because this, increase their geographical distribution. Our results showed new perspectives and directions for the success of diversity of amphibians in the Brazilian Atlantic Forest.

**Key words:** functional traits; dispersion; homology; phylogenetic signal

#### 4.1 INTRODUÇÃO

Uma das principais pressões evolutivas sobre as espécies é a corrida predador-presa. Este processo evolutivo opera em conjunto com mudanças genéticas aleatórias selecionadas, gerando fenótipos especializados através de um processo de seleção natural (Darwin 1859). Predadores afetam as presas diretamente pelo consumo e, indiretamente, pela indução de respostas antipredação energeticamente caras (Alcock 2011). Estas respostas podem incluir alterações nos traços comportamentais, morfológicos, ou no desenvolvimento das presas (Smith e Van Bus-kirk 1995; Preisser et al. 2005; Verheggen et al. 2009). Mecanismos antipredação são essenciais para a sobrevivência da presa (Lima 1998; Ruxton et al. 2004). No entanto, ambos os lados (predador e presa) podem não ter o controle dessas adaptações (Petranka et al. 1987; Podjasek et al. 2005; Alckook 2011), e às vezes o predador sai na frente desta corrida evolucionária (Brodie e Brodie 1999). Devido a isso, a evolução e a regulação de um determinado comportamento em resposta a predação pode ser rápida (Stillfried et al. 2015) e em poucas gerações (Juliano e Cascalho 2002; Zimmer et al. 2011).

O sucesso de um mecanismo antipredação é o que permite a evolução (Vermeij 1982). Correlacionar os mecanismos antipredação são particularmente importantes para a compreensão da seleção natural sobre um único mecanismo (Brodie1992). Presas que apresentam fracas respostas comportamentais a predadores, ou aparentemente abaixo do ideal, podem não sofrer predação pesada se elas apresentarem defesas morfológicas e químicas eficazes (Dewit et al. 1999; Willians 2000; Yadete et al. 2012). Neste sentido, a evolução de um mecanismo requer que as espécies possuam características filogenéticas que direcionem a ocorrência de um mecanismo eficiente para a sua sobrevivência.



A origem evolutiva do mecanismo antipredação está diretamente relacionada ao sucesso adaptativo das espécies, e pode refletir em novas estratégias defensivas ou simplesmente o ajuste de um mecanismo existente (Dewit et al. 1999; Juliano e Gravel 2002; Zimmer et al. 2011; Stillfried et al. 2015 ). Alternativamente, o comportamento e a morfologia podem ser mecanicamente independentes, e essas características muitas vezes devem ser utilizadas em combinações particulares para produzir uma resposta eficaz de antipredação (Dewit et al. 1999). Espécies filogeneticamente próximas tendem a ter características comportamentais similares herdadas de um ancestral comum (Harvey e Pagel 1991), levando-as a explorar e se estabelecer em ambientes semelhantes. Esta semelhança fenotípica entre espécies relacionadas é conhecida como sinal filogenético (Harvey e Pagel 1991; Blomberg e Garland 2002) e descreve a tendência de uma característica particular em ser conservada. No entanto, a diversidade de características fenotípicas, podem apresentar diferentes direções que irão depender da sua história evolutiva. Esta característica evolutiva pode ter origem na raiz de uma árvore filogenética, ou podem convergir e as características serem encontradas nas pontas da árvore (Pavoine et al. 2010). Além disso, a evolução destas características pode ser explicadas pelo modelo Browniano de evolução, que é um processo de deriva genética aleatória a uma taxa constante de evolução não direcional (Felsenstein 1985; Lynch 1990; Martins 1994; Pagel 1997; Diniz-Filho e Vieira 1998 ).

Características funcionais podem influenciar a área de distribuição das espécies (Brown e Maurer 1989; Díaz et al 2007; Jimenez-Val Verde et al 2015). Estas características podem conduzir a presa para ajustar sua estratégia antipredação, dependendo da atual ameaça imposta por predadores potenciais (Smith e Van Bus-kirk 1995; Relyea 2001; Ferrari et al. 2010; Higginson et al. 2012; Brown et al. 2013). Como

consequência, em mecanismos evasivos (fuga) e defensivos (contato) é esperado que haja flexibilidade fenotípica em resposta ao nível do risco da predação (Kats e Dill 1998; Lass e Spaak 2003; Ferrari et al. 2010; Higginson et al. 2012; Brown et al. 2013) .

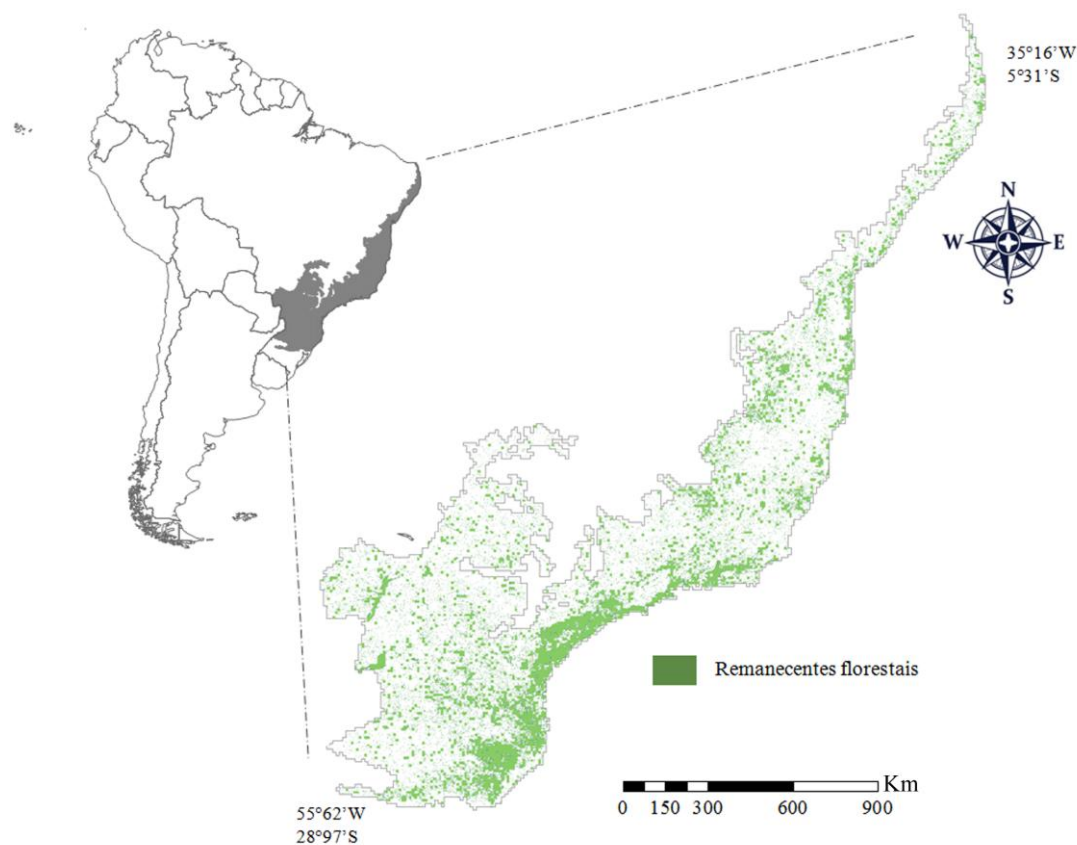
Anfíbios são predados por várias espécies de vertebrados e invertebrados em todos os seus estágios de vida (desde adultos, bem como larvas e juvenis) (Toledo 2005; Wells 2007). Esta forte pressão promovida pela predação atua como força de seleção evolucionária (Wells 2007). Vários mecanismos de antipredação foram desenvolvidos pelas espécies de anfíbios. Toledo e colaboradores (2011) listaram mais de 30 mecanismos diferentes em anuros. Muitas espécies de anuros apresentam estes comportamentos em sinergia (Lourenço-de-Moraes et al. 2014a, Figueiredo et al. no prelo) e de forma escalonada (Lourenço-de-Moraes et al. 2014b, 2016). A origem filogenética da maioria dos mecanismos de antipredação ainda são desconhecidas para os anfíbios. Correlacionar os traços destes mecanismos e, em particular, a evolução conjunta dos traços de cada mecanismo é importante para se determinar a sua história evolutiva (Lande & Arnold 1983; Price & Langen 1992), e pode responder a perguntas sobre as vantagens e desvantagens de um traço particular em relação ao seu potencial de distribuição geográfica.

Nesse sentido, nós testamos a hipótese de que: i) mecanismos de antipredação ancestrais são puramente filogenéticos explicados pelo modelo de evolução Browniano; ii) a especialização ecológica (funcional) das espécies influencia a diversidade de mecanismos antipredação; iii) espécies com maior diversidade de mecanismos antipredação têm maior área de distribuição geográfica. Para isso, avaliamos os mecanismos ancestrais e analisamos sua origem evolutiva com base no repertório de mecanismos de antipredação das espécies de anfíbios da Floresta Atlântica.

## 4.2 MÉTODOS

### 4.2.1 Área de estudo

As análises foram focadas nas espécies que ocorrem na Floresta Atlântica Brasileira, considerada um dos hotspots da biodiversidade (Myers et al., 2000), que originalmente cobria cerca de 150 milhões de hectares de condições ambientais heterogêneas fornecidas por uma grande variedade de zonas climáticas e formações florestais (Tabarelli et al. 2005; Ribeiro et al. 2009) (Figura 1).



**Figura 1.** Mapa mostrando a área de estudo no hotspot Floresta Atlântica brasileira (em cinza) no território Sulamericano.

#### 4.2.2 *Dados das espécies*

Foram obtidos dados espaciais de espécies de anfíbios em uma pesquisa conjunta através de quatro tipos de procedimentos: (a) foi construído um conjunto de dados com todas as espécies distribuídas na Floresta Atlântica de acordo com o banco de dados de mecanismos antipredação proposto por Ferreira e colaboradores (em preparação) e o banco de dados com as características funcionais de Haddad e colaboradores (2013); (b) selecionamos as espécies baseados no banco de dados GenBank (Benson et al. 2013) para a construção da árvore filogenética; (c) realizamos trabalhos de campos complementares nos principais remanescentes da Floresta Atlântica Brasileira, para complementar o conjunto de dados de mecanismos antipredação e os atributos funcionais observados (Apêndice 3, Tabela S1 e S2); e (d) determinamos a área geográfica de cada espécie baseados na Lista Vermelha da IUCN pelo banco de dados de Espécies Ameaçadas (IUCN, 2016).

No total, foram avaliados os limites geográficos (IUCN 2016) de 115 espécies de anfíbios cobertos por nosso sistema de grid. Usamos o software ArcGIS10 (ESRI, 2010) para construir matrizes de presença/ausência de dados de distribuição de espécies por meio da sobreposição de um sistema de grade com células de 0,1 graus de latitude/longitude (10 x 10 km), criando uma rede com 10,359 células da grade. Cada célula de grade corresponde a 1 range de ocorrência.

#### 4.2.3 *Mecanismos antipredação*

Para cada espécie encontrada durante os levantamentos de campo (105 das 115 espécies avaliadas, veja Apêndice 3, Tabela S2) nos aplicamos os estímulos de

mecanismos antipredação, utilizando a metodologia I (usando o humano como predador utilizando somente o dedo para aplicar os estímulos) proposta por Lourenço-de-Moraes e colaboradores (2016), mas adaptamos o tempo de cada aplicação em relação ao tamanho da espécie (para espécies  $\leq 3$  cm o tempo de cada aplicação foi de 15s, para espécies maiores o tempo foi de 30s). Os mecanismos antipredação foram classificados em 13 categorias de acordo com o sistema de classificação proposto por Ferreira, Lourenço-de-Moraes e Brodie Jr (em preparação). Estes mecanismos foram divididos nas seguintes categorias: produção de secreções, camuflagem, imobilidade, contração, inflar o corpo, elevação do corpo, manter as pernas esticadas, fuga, aposematismo, descarga cloacal, abrir a boca, vocalizações defensivas e lutar. A descrição e as variações dos mecanismos estão na Tabela 1.

**Tabela 1.** Mecanismos antipredação avaliados e suas variações dentro de cada mecanismo.

<b>Mecanismos de Antipredação</b>	<b>Variações dos Mecanismos</b>
Produzir secreções	0= Ausência; 1=Odorífera; 2=Adesiva; 3=Tóxica; 4=Escorregadia; 5 = Desagradavel
Camuflagem	0= Ausência; 1= Presença
Imobilidade	1= Imóvel agachada; 2= Imóvel alerta; 3= Tanatose
Contração	0= Ausência; 1= Contração; 2= Pragmose; 3= Agachando-se; 4= Dobrando o queixo; 5=Protegendo o olho
Inflar o corpo	0= Ausência; 1= Presença
Elevar o corpo	0= Ausência; 1= Pernas esticadas verticalmente; 2= Pernas esticadas lateralmente; 3= Elevação; 4= Elevação lateral
Manter as pernas esticadas	0= Ausência; 1= Presença
Escapar	1= Único pulo; 2= Varios pulos; 3= Esconder-se
Aposematismo	0= Ausência; 1= Total; 2= Parcial
Descarga	
Cloacal	0= Ausência; 1= Líquido; 2= Sólido
Abrir a boca	0= Ausência; 1= Presença
Vocalização defensiva	0= Ausência;1= Canto de agônia; 2= Canto de alarme
Lutar contra o predador	0= Ausência; 1= Movimentos Erráticos; 2=Lutar; 3=Agressão com espinho; 4=Avançar; 5= Cabeçada; 6=Morder

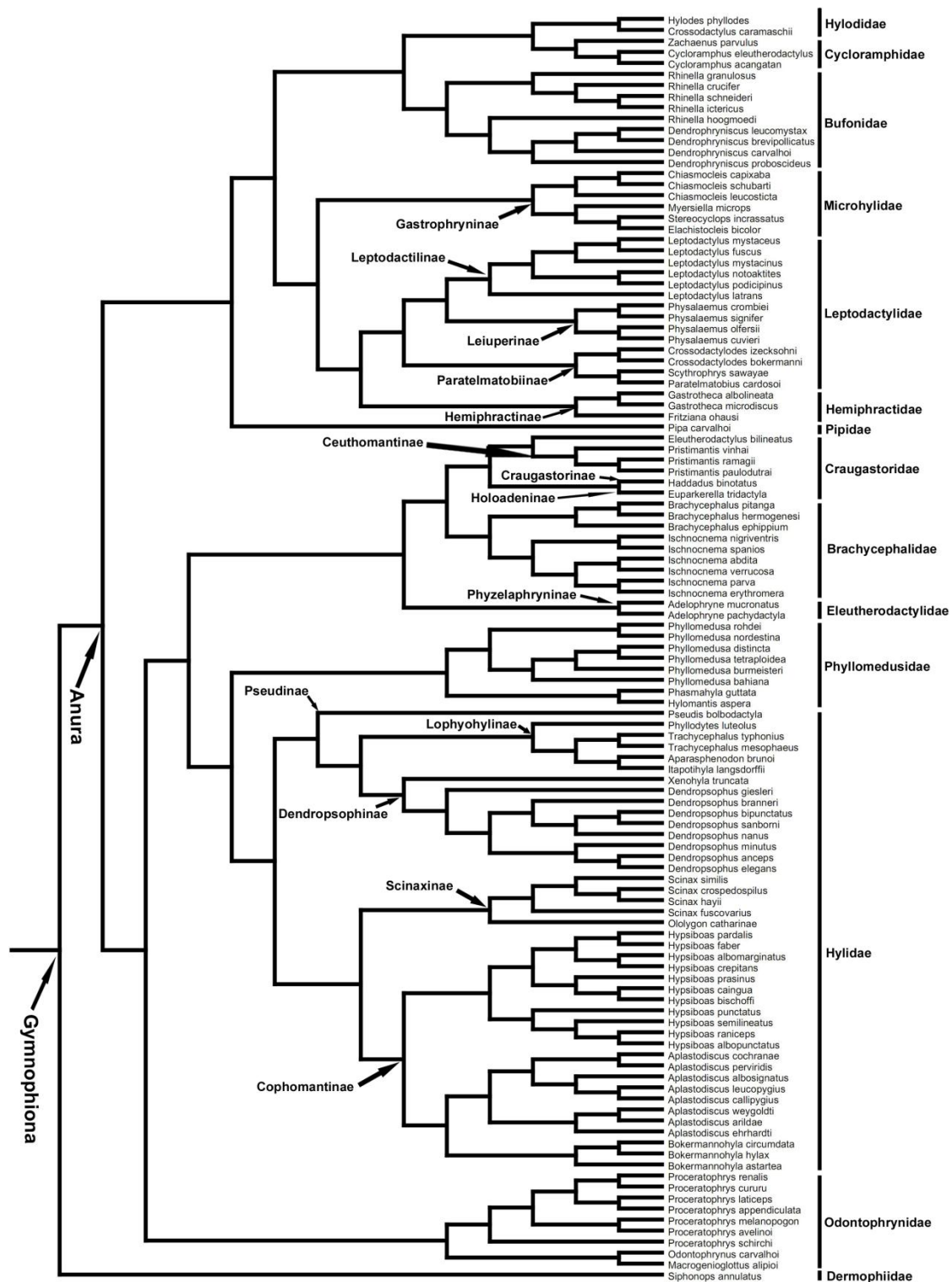
#### 4.2.4 *Dados filogenéticos*

Nós baseamos a distância filogenética das 115 espécies estudadas (Apêndice 3, Tabela S3) através de sequências de nucleótidos obtidas no GenBank (Benson et al. 2013), fornecidas pelo Centro Nacional de Informações sobre Biotecnologia (NCBI). Seguindo o protocolo proposto por Pyron e Wiens (2011) baseado em uma filogenia de anfíbios existentes. Foram utilizados 12 genes para gerar a filogenia para os anfíbios da Floresta Atlântica (ou seja, 11, 906 pb para cada espécie), foram incluídos três genes mitocondriais: cytochromo b (cyt-b), e as grandes e pequenas sub-unidades dos genes mitocondriais ribossômicos (12S/16S); e nove genes nucleares: tipo C-X-C receptor de quimiocinas 4 (CXCR4), histona 3a (H3A), alternador de sódio-cálcio (NCX1), pró-opiomelanocortin (POMC), gene ativador de recombinação 1 (RAG1), rodopsina (Rhod), sétimo ausente (SIA), família soluto-carrier 8 (SLC8A3) e tirosinase (TYR). Para as regiões de comprimento variável, realizamos várias comparações de pares pela versão online do MAFFT v.6.8 e G-INS-i algoritmo (Kato e Toh 2008). Em seguida, alinhamos todos os genes em um mesmo conjunto alinhado usando o SequenceMatrix software 1.7.7 (Vaidya 2011) para concatenar a supermatrix gerada.

Nós conduzimos as relações filogenéticas com análises Bayesianas no software BEAST 1,8 (Drummond e Rambaut 2007), estimando-se os nós suportados com uma análise de bootstrapping. Depois, foram realizadas análises Bayesianas em uma matriz de dados combinados usando um modelo de sequência evolutiva HKY que particiona todos os genes, com um processo de especiação (Yule) e um relógio molecular relaxado não correlacionado. Após a remoção dos genes considerados indispensáveis, partimos

para o processo de especiação (Yule) com 100 milhões de gerações, garantindo que o número de gerações depois de convergência fosse suficientemente avaliado utilizando o uso do Tracer v1.6 (Drummond e Rambaut 2007), combinando os resultados com o uso de Logcombiner 1.8 e o Tree Anotator 1.8 (Drummond e Rambaut 2007). Consideramos que cada nó é fortemente suportado com valores de probabilidade (pp) suporte de  $\geq 0.95$ . Para resolver as politomias, usamos a função 'multi2di' do pacote 'ape' (Paradis 2012) no software R (R Development Core Team 2010). Assim, reconstruímos uma nova árvore filogenética (Figura 2) usando o software Mesquite, versão 3.0 (Maddison e Maddison 2014).





**Figura 2.** Árvore original. A nomenclatura das Famílias, Subfamílias e espécies segue Frost (2016). A árvore mostra as relações de parentesco entre as espécies. Barras laterais indicam as Famílias, flechas indicam os nós das Ordens e Subfamílias.

#### 4.2.5 *Características funcionais*

Nós caracterizamos as características funcionais das 115 espécies de anfíbios de acordo com Haddad e colaboradores (2013) e com as nossas observações de campo. As características funcionais avaliadas foram: tamanho do corpo (Pequeno <3 cm, Médio  $\geq 3$  e <10cm, Grande  $\geq 10$  cm), o modo de desenvolvimento reprodutivo (direto ou indireto), *Habitat* (áreas florestais, áreas abertas e áreas abertas e florestais), atividade (noturna, diurna, e ambos), e o hábito (arbóreo, fitotelmata, terrestre, criptozoico, fossorial, reofílicos, e aquáticos). Veja detalhes e características das espécies no Apêndice 3 Tabela S2.

### 4.3 Análise dos dados

#### 4.3.1 *Reconstrução dos estados ancestrais*

Para determinar os mecanismos antipredação ancestrais, criamos uma árvore parcimoniosa entre os mecanismos antipredação e a árvore filogenética original. As análises das árvores ancestrais foram desenvolvidas com o auxílio do software Mesquite, versão 3.0 (Maddison e Maddison, 2014). O método de reconstrução de parcimônia para encontrar os estados ancestrais, minimiza o número de etapas da mudança de um caráter dadas pela árvore original e a distribuição do traço observado. Para isso, nós utilizamos o método de "História de Rastreamento do Caractere", que pinta os ramos da árvore para mostrar a reconstrução, e o traço ancestral é definido para cada nó na árvore.

Foram avaliados os traços ancestrais dos seguintes 'nós' da árvore original: Ordem Anura e Gymnophiona; Famílias Brachycephalidae, Bufonidae, Craugastoridae, Cycloramphidae, Dermophiidae, Eleutherodactylidae, Hemiphractidae, Hylidae, Hyloidae, Leptodactylidae, Microhylidae, Odontophrynidae, Phyllomedusidae, e Pipidae; e as subfamílias Ceuthomantinae, Cophomantinae, Craugastorinae, Dendropsophinae, Holoadeninae, Leiuperinae, Leptodactilinae, Lophyohylinae, Paratelmatobiinae, Pseudinae e Scinaxinae (ver Figura 2). Seguimos Frost (2016) para a nomenclatura das Famílias, Subfamílias e espécies.

#### 4.3.2 Sinal filogenético

Para estimar o sinal filogenético de cada mecanismo antipredação, usamos uma medida robusta proposta por Abouheif (1999) para detectar sinal filogenético nos mecanismos antipredação. O teste de Abouheif utiliza o índice de I de Moran, mas a matriz de proximidades filogenéticas de Abouheif tem uma diagonal diferente de zero. Estas aproximações são baseadas usando tanto os testes de C de Geary e o I de Moran (Legendry e Fortin 1989). O teste foi realizado com 999 aleatorizações utilizando o pacote "adephylo" (Pavoine et al. 2008) no software R (R Development Core Team 2010).

Para testar quais os mecanismos antipredação seguem o modelo de evolução Browniano (evolução aleatória), utilizamos uma medida baseada no máximo de probabilidade de sinal filogenético, chamado de lambda ( $\lambda$ ), desenvolvido por Pagel (1997). Esta métrica corresponde a um parâmetro de transformação da árvore que elimina gradualmente a estrutura filogenética variando de 1 a 0. Lambda transformação

é realizado através da multiplicação dos elementos fora da diagonal da matriz de variância/covariância descrevendo os comprimentos da topologia da árvore e ramos. Valores de lambda igual a 1 correspondem a evolução Browniano, enquanto que para os valores de lambda igual a 0 corresponde à ausência completa da estrutura filogenética (filogenia star-like). A estimativa de lambda pode ser comparada a zero pelo cálculo de uma razão de probabilidade (likelihood) e comparando-a com uma distribuição de Qui-quadrado com um grau de liberdade. Para as análises nós usamos a função 'phylosig' do pacote 'phytools' (Revell 2012) no software R (R Development Core Team 2010).

#### 4.3.3 *Mecanismos antipredação vs características funcionais*

Afim de avaliar a resposta da diversidade de mecanismos antipredação de cada espécie relacionados ao tamanho do corpo, modo de desenvolvimento reprodutivo, hábito, *habitat* e atividade, nós utilizamos uma análise de permutação multivariada de variância (PERMANOVA), com 1,000 permutações com base numa matriz de distância euclidiana através da função "adonis". Além disso, usamos Boxplots para visualizar as características que melhor explicaram a diversidade de mecanismos antipredação geradas pela PERMANOVA. Realizamos as análises usando o pacote 'vegan' (Oksanen et al. 2013), em R (Team R Development Core 2012).

#### 4.3.4 *Distribuição geográfica vs. mecanismos antipredação*

Para avaliar se as espécies com diversidade de mecanismos antipredação influenciam a área de ocorrência das espécies, usamos matrizes de correlação para

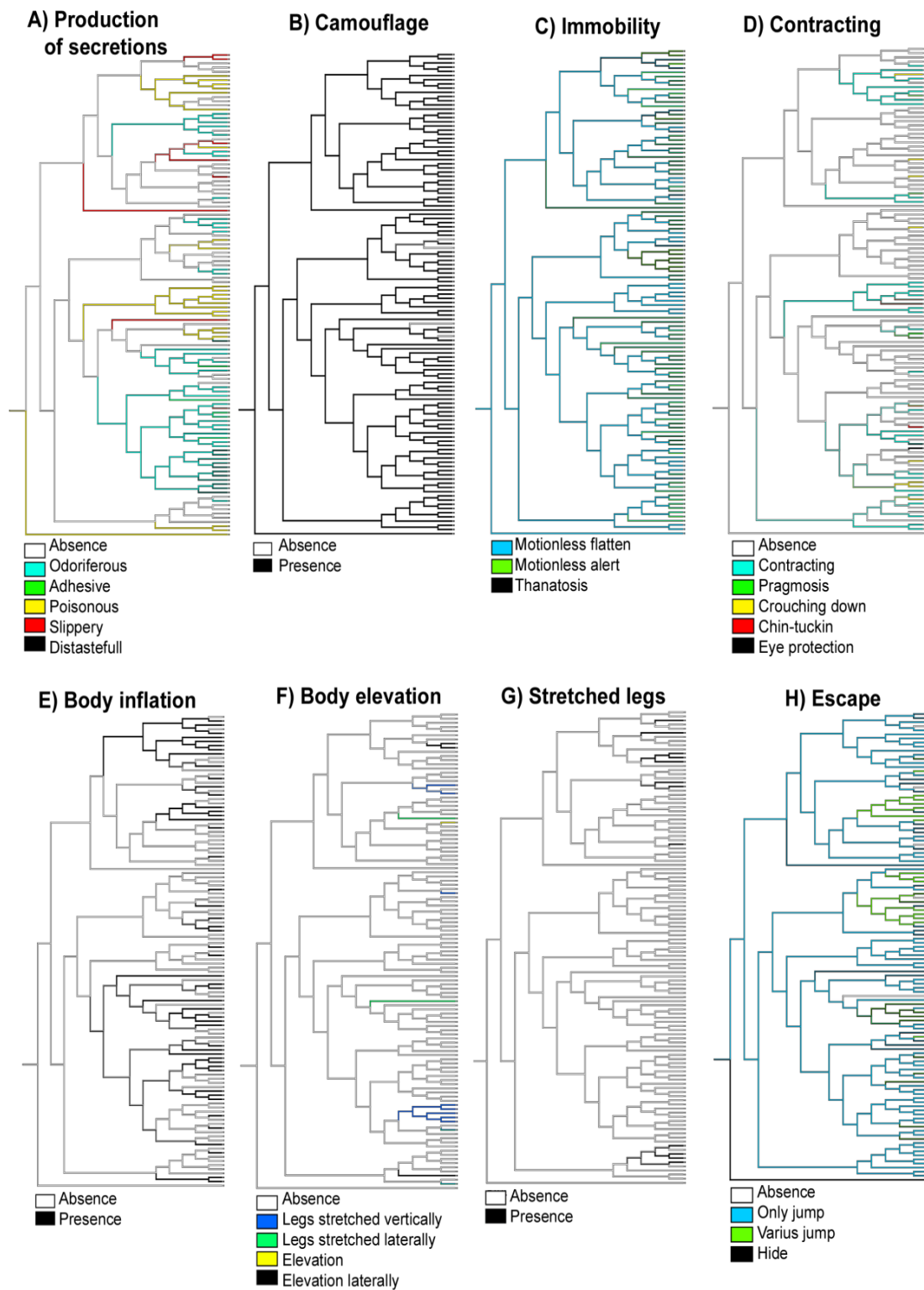
comparar a diversidade de mecanismos de cada espécie e suas referências espaciais (intervalos de distribuição), utilizando modelos de regressão linear simples. Estas correlações foram realizadas para todos os mecanismos e suas variações (ver Tabela 1). As análises foram realizadas utilizando o pacote 'vegan' (Oksanen et al. 2007) em R (Team R Development Core 2012).

#### 4.4 Resultados

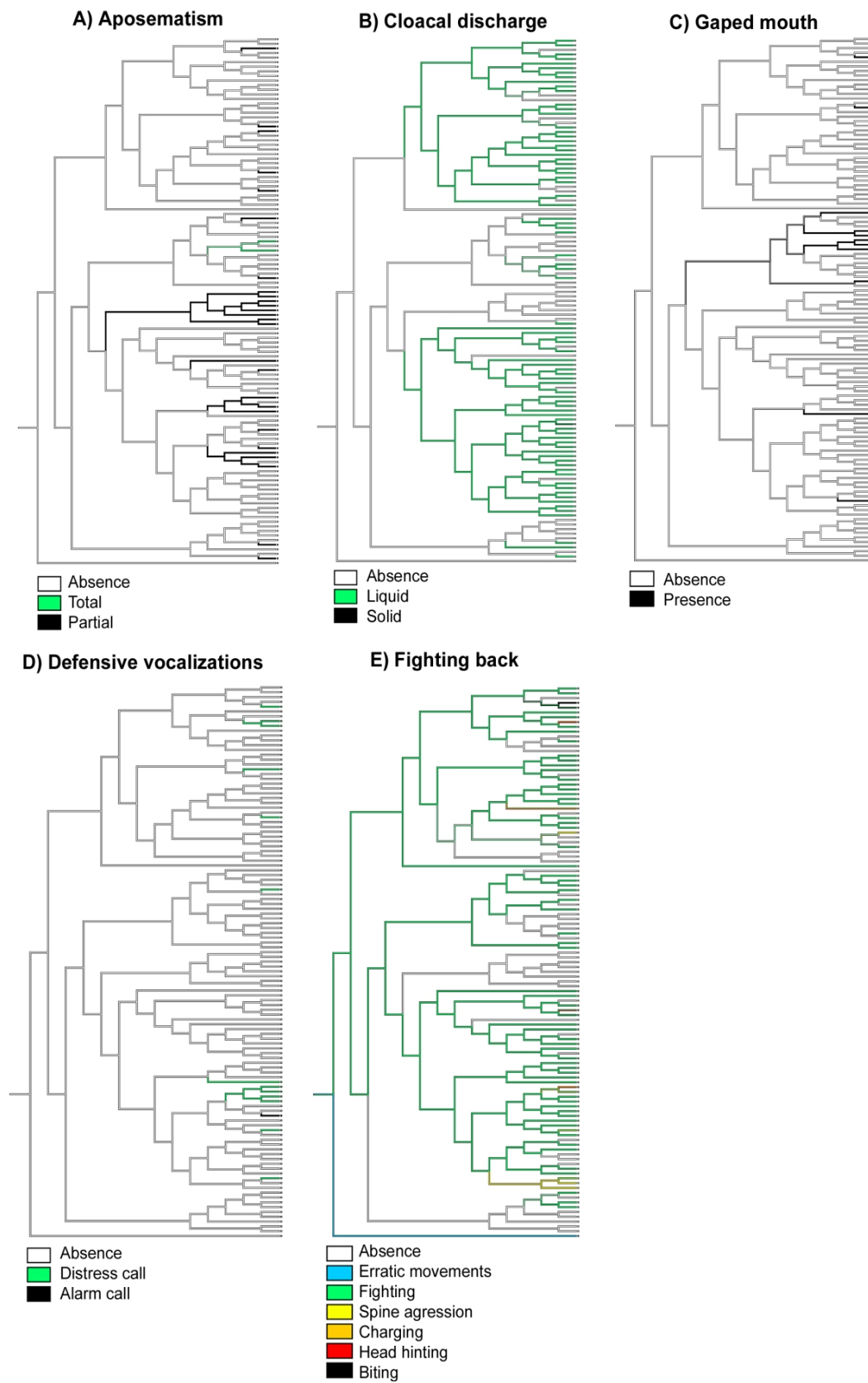
##### 4.4.1 *Estados ancestrais*

Os resultados mostraram que os mecanismos antipredação de camuflagem, imobilidade, lutar, e fuga são os estados ancestrais de Anura e Gymnophiona (simplesiomórficos) com base em 115 espécies (duas ordens, 14 famílias e 12 subfamílias, ver a Figura 2, 3 e 4). O mecanismo de lutar não aparece como estado ancestral apenas para as famílias Dermophiidae, Hemiphractidae, Phyllomedusidae e Odontophrynidae. O mecanismo de fuga apresenta duas variações apomórficas em anuros "único pulo" e "vários pulos" sendo que "vários pulos" é um estado homólogo para as famílias Brachycephalidae e Eleutherodactilidae (subfamília Ceuthomantinae), tendo evoluído de forma independente na subfamília Leptodactilinae (homoplasia) (Figura 4E).

O mecanismo de inflar o corpo é um estado apomórfico que surgiu independentemente através de diferentes ancestrais, sendo considerado homoplástico em Bufonidae, Microhylidae (sinapomorfia de Gastrophryninae), Leptodactylidae (sinapomorfia de Leptodactiilinae e Leiuperinae), Craugastoridae (sinapomorfia de Holoadeninae), Hylidae (sinapomorfia de Cophomantinae) e Cycloramphidae (Figura 3E).



**Figura 3.** Reconstrução filogenética parcimoniosa entre os mecanismos antipredação e a árvore original.



**Figura 4.** Reconstrução filogenética parcimoniosa entre os mecanismos antipredação e a árvore original.

**Tabela 4.** Lista de mecanismos antipredação ancestrais para cada Ordem, Família e Subfamília das 115 espécies de anfíbios na Floresta Atlântica geradas pelas árvores parcimoniosas.

Mecanismos de Antipredação Ancestrais													
Ordem/Família/ Subfamília	Produzir secreções	Camuflagem	Imobilidade	Contração	Inflar o Corpo	Elevar o Corpo	Manter as pernas esticadas	Escapar	Aposematismo	Descarga Cloacal	Abrir a Boca	Vocalização defensiva	Lutar contra o predador
<b>Gymnophiona</b>	<b>Ausência e Tóxico</b>	<b>Presença</b>	<b>Imovel agachada</b>	Ausência	Ausência	Ausência	Ausência	<b>Esconder</b>	Ausência	Ausência	Ausência	Ausência	<b>Movimentos erráticos &amp; Lutar</b>
<b>Dermophiidae</b>	Ausência	<b>Presença</b>	<b>Imovel agachada</b>	Ausência	Ausência	Ausência	Ausência	Ausência	Ausência	Ausência	Ausência	Ausência	Ausência
<b>Anura</b>	Ausência	<b>Presença</b>	<b>Imovel agachada</b>	Ausência	Ausência	Ausência	Ausência	<b>Único pulo</b>	Ausência	Ausência	Ausência	Ausência	<b>Lutar</b>
<b>Bufoidea</b>	<b>Tóxico</b>	<b>Presença</b>	<b>Imovel agachada</b>	<b>Contração</b>	<b>Presença</b>	Ausência	Ausência	<b>Único pulo</b>	Ausência	<b>Líquido</b>	Ausência	Ausência	<b>Lutar</b>
<b>Brachycephalidae</b>	Ausência	<b>Presença</b>	<b>Imovel agachada</b>	Ausência	Ausência	Ausência	Ausência	<b>Vários pulos</b>	Ausência	Ausência	<b>Ausência &amp; Presença</b>	Ausência	<b>Lutar</b>
<b>Craugastoridae</b>	Ausência	<b>Presença</b>	<b>Imovel agachada</b>	Ausência	Ausência	Ausência	Ausência	<b>Único pulo</b>	Ausência	Ausência	<b>Ausência &amp; Presença</b>	Ausência	<b>Lutar</b>
Ceuthomantinae	<b>Odorífero</b>	<b>Presença</b>	<b>Imovel agachada</b>	Ausência	Ausência	Ausência	Ausência	<b>Vários pulos</b>	Ausência	<b>Líquido</b>	Ausência	Ausência	<b>Lutar</b>
Craugastorinae e Holoadeninae	Ausência	<b>Presença</b>	<b>Imovel agachada</b>	Ausência	Ausência	Ausência	Ausência	<b>Único pulo</b>	Ausência	Ausência	<b>Presença</b>	Ausência	<b>Lutar</b>



<b>Cycloramphidae</b>	Ausência	<b>Presença</b>	<b>Imóvel agachada &amp; Thanatosis</b>	Ausência	<b>Presença</b>	Ausência	Ausência	<b>Único pulo</b>	Ausência	<b>Líquido</b>	Ausência	Ausência	<b>Lutar &amp; Morder</b>
<b>Eleutherodactylidae</b>	Ausência	<b>Presença</b>	<b>Imóvel agachada</b>	Ausência	Ausência	Ausência	Ausência	<b>Único pulo</b>	Ausência	Ausência	<b>Ausência &amp; Presença</b>	Ausência	<b>Lutar</b>
Phyzelaphryninae	Ausência	<b>Presença</b>	<b>Imóvel agachada</b>	Ausência	Ausência	Ausência	Ausência	<b>Único pulo</b>	Ausência	Ausência	<b>Ausência &amp; Presença</b>	Ausência	<b>Lutar</b>
<b>Hemiphractidae</b>	Ausência	<b>Presença</b>	<b>Imóvel agachada</b>	<b>Contração</b>	Ausência	Ausência	Ausência	<b>Único pulo</b>	Ausência	<b>Líquido</b>	Ausência	Ausência	Ausência
<b>Hylidae</b>	Ausência	<b>Presença</b>	<b>Imóvel agachada</b>	Ausência	<b>Ausência &amp; Presença</b>	Ausência	Ausência	<b>Único pulo</b>	Ausência	<b>Líquido</b>	Ausência	Ausência	<b>Lutar</b>
Comophantinae	<b>Odorifero</b>	<b>Presença</b>	<b>Imóvel agachada</b>	<b>Ausência &amp; Contração</b>	<b>Ausência &amp; Presença</b>	Ausência	Ausência	<b>Único pulo</b>	Ausência	<b>Líquido</b>	Ausência	Ausência	<b>Lutar</b>
Dendropsodinae	Ausência	<b>Presença</b>	<b>Imóvel agachada</b>	Ausência	<b>Ausência &amp; Presença</b>	Ausência	Ausência	<b>Único pulo</b>	Ausência	<b>Líquido</b>	Ausência	Ausência	<b>Lutar</b>
Lophyohylineae	Ausência	<b>Presença</b>	<b>Imóvel agachada</b>	Ausência	<b>Ausência &amp; Presença</b>	Ausência	Ausência	<b>Único pulo</b>	Ausência	<b>Líquido</b>	Ausência	Ausência	<b>Lutar</b>
Pseudinae	Ausência	<b>Presença</b>	<b>Imóvel agachada</b>	Ausência	Ausência	Ausência	Ausência	<b>Único pulo</b>	Ausência	Ausência	Ausência	Ausência	<b>Lutar</b>
Scinaxinae	<b>Odorifero</b>	<b>Presença</b>	<b>Imóvel agachada</b>	Ausência	<b>Ausência &amp; Presença</b>	Ausência	Ausência	<b>Único pulo</b>	Ausência	<b>Líquido</b>	Ausência	Ausência	<b>Lutar</b>
<b>Hylodidae</b>	<b>Escorregadia</b>	<b>Presença</b>	<b>Imóvel agachada &amp; Thanatosis</b>	Ausência	<b>Presença</b>	Ausência	Ausência	<b>Único pulo</b>	Ausência	<b>Líquido</b>	Ausência	Ausência	<b>Lutar</b>
<b>Leptodactylidae</b>	Ausência	<b>Presença</b>	<b>Imóvel agachada</b>	Ausência	Ausência	Ausência	Ausência	<b>Único pulo</b>	Ausência	<b>Líquido</b>	Ausência	Ausência	<b>Ausência &amp; Lutar</b>

Leiuiperinae	Ausência	<b>Presença</b>	<b>Imóvel agachada</b>	Ausência	Ausência	Ausência	Ausência	<b>Único pulo</b>	Ausência	<b>Liquido</b>	Ausência	Ausência	<b>Lutar</b>
Leptodactylinae	<b>Ausência e Escorregadia</b>	<b>Presença</b>	<b>Imóvel agachada</b>	Ausência	<b>Presença</b>	Ausência	Ausência	<b>Vários pulos</b>	Ausência	<b>Liquido</b>	Ausência	Ausência	<b>Lutar</b>
Paratelmatoibiinae	Ausência	<b>Presença</b>	<b>Imóvel agachada</b>	Ausência	Ausência	Ausência	Ausência	<b>Único pulo</b>	Ausência	<b>Liquido</b>	Ausência	Ausência	<b>Ausência &amp; Lutar</b>
<b>Microhylidae</b>	<b>Odorifero</b>	<b>Presença</b>	<b>Imóvel agachada</b>	Ausência	Ausência	Ausência	Ausência	<b>Único pulo</b>	Ausência	<b>Liquido</b>	Ausência	Ausência	<b>Lutar</b>
<b>Odontophryinae</b>	Ausência	<b>Presença</b>	<b>Imóvel agachada</b>	<b>Ausência &amp; Contração</b>	Ausência	Ausência	Ausência	<b>Único pulo</b>	Ausência	Ausência	Ausência	Ausência	Ausência
<b>Pipidae</b>	Ausência	<b>Presença</b>	<b>Imóvel agachada</b>	Ausência	Ausência	Ausência	Ausência	<b>Único pulo</b>	Ausência	Ausência	Ausência	Ausência	<b>Lutar</b>
<b>Phyllomedusidae</b>	<b>Tóxico</b>	<b>Presença</b>	<b>Imóvel agachada</b>	<b>Contração</b>	Ausência	Ausência	Ausência	<b>Único pulo</b>	<b>Parcial</b>	Ausência	Ausência	Ausência	Ausência

O mecanismo de produzir secreções, apesar de ser um estado simplesiomórfico, apareceu em Anura de forma homoplástica com diferentes especializações ou ajustes ao longo de sua evolução. Duas subfamílias dentro de Hylidae (Scinaxinae e Cophomantinae) apresentam uma produção de secreção do tipo odorífera como característica homóloga. O mecanismo de produzir secreções do tipo escorregadia, é uma característica homoplástica presente na família Hylidae e na subfamília Leptodactylinae. O mecanismo de produzir secreções tóxica está presente em Bufonidae e Phyllomedusidae (homoplasia) (Figura 2A).

O mecanismo de contração é apomórfico e surgiu de forma independente em Anura; ele é considerado uma homoplasia em Bufonidae, Hemipractidae, Phyllomedusidae, Odontophrynidae e na subfamília Comophantinae. O mecanismo de abrir a boca é um estado apomórfico, homólogo em Terrarana (que corresponde as Famílias Craugastoridae, Brachycephalidae e Eleutherodactylidae) e pode ser uma sinapomorfia para o grupo Terrarana (Figura 3D). O mecanismo de elevar o corpo, manter as pernas esticadas e vocalizações defensivas não foram indicados como mecanismos ancestrais em nenhum dos “nós” avaliados (ver Tabela 4).

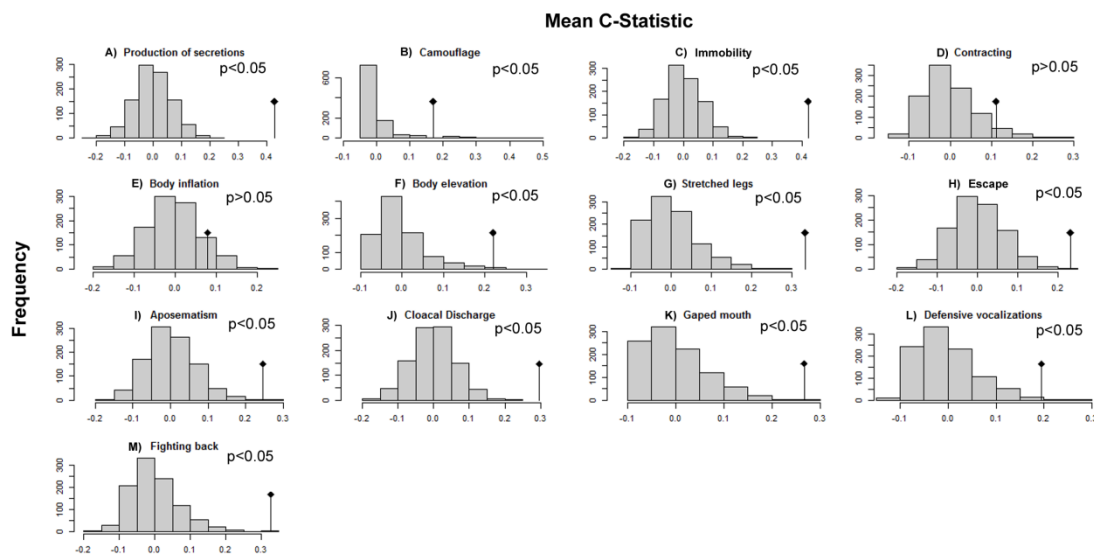
#### 4.4.2 Sinal filogenético

Os resultados indicaram que por meio do índice de Abouheif que a maioria dos mecanismos antipredação apresenta um sinal filogenético significativo. Os mecanismos que mostraram sinais filogenéticos fortes foram o de produzir secreções (média  $C = 0,43$ ,  $p < 0,05$ ) e imobilidade (média  $C = 0,42$ ,  $p < 0,05$ ), seguido por lutar (média  $C = 0,33$ ,  $p < 0,05$ ) e manter as pernas esticadas (média  $C = 0,33$ ,  $p < 0,05$ ) com valores

ligeiramente baixos. Os mecanismos de contração (média  $C = 0,11$ ,  $p > 0,05$ ) e de inflar o corpo (média  $C = 0,08$ ,  $p > 0,05$ ), não mostraram sinais filogenéticos significativos (Tabela 2 Figura 5).

**Tabela 2.** Resultados dos índices de Pagel e Abouheif

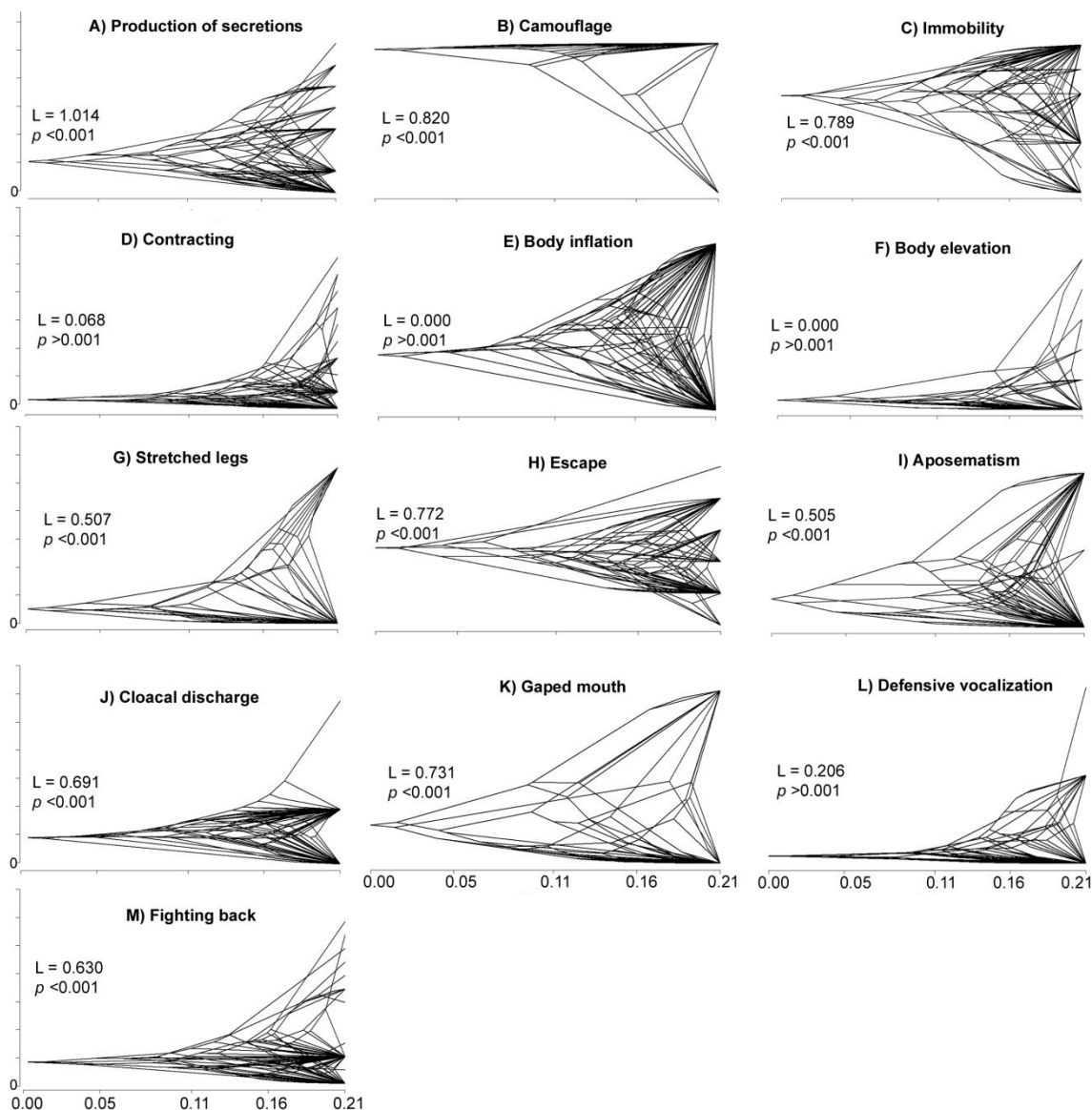
Mecanismos Antipredação	Pagel's $\lambda$	$P$	Abouheif's	
			C-mean	$P$
Produzir secreções	1.0145	0.0000	0.4273	0.001
Camuflagem	0.8200	0.0008	0.1707	0.031
Imobilidade	0.7897	0.0000	0.4204	0.001
Contração	0.0680	0.3834	0.0800	0.063
Inflar o corpo	0.0001	1.0000	0.2192	0.099
Elevar o corpo	0.0001	1.0000	0.3337	0.011
Manter as pernas esticadas	0.5073	0.0001	0.2323	0.001
Escapar	0.7729	0.0001	0.2465	0.001
Aposematismo	0.5052	0.0001	0.2973	0.003
Descarga cloacal	0.6911	0.0000	0.2667	0.001
Abrir a boca	0.7310	0.0000	0.1950	0.001
Vocalização defensiva	0.2065	0.3236	0.3278	0.006
Lutar contra o predador	0.6304	0.0086	0.4273	0.001



**Figura 5.** Os círculos negros indicam a posição da média observada em relação à distribuição da média estatística-C randomizados. O P-valor da estatística C média observada é significativa em um alfa de 0,05. As simulações representam a média estatística-C calculada a partir dos mecanismos antipredação ao longo das pontas da filogenia.

#### 4.4.3 Valores de lambda ( $\lambda$ ): evolução Browniano

Os valores de Pagel lambda indicaram que os mecanismos de produzir secreção ( $\lambda = 1,01$ ,  $p < 0,001$ ) valor acima de 1, camuflagem ( $\lambda = 0,82$ ,  $p < 0,001$ ) e imobilidade ( $\lambda = 0,79$ ,  $p < 0,001$ ) mostraram valores próximos de 1. Os mecanismos de contração ( $\lambda = 0,07$ ,  $p > 0,001$ ), inflar o corpo ( $\lambda = 0,00$ ,  $p > 0,001$ ) e elevação do corpo ( $\lambda = 0,00$ ,  $p > 0,001$ ) não apresentaram valores significativos de evolução Browniano (Tabela 2; Figura 6).



**Figura 6.** Os fenogramas mostra as simulações e os valores lambda Pagel ( $\lambda$ ). Eixo 'x' indica o número de tipos de mecanismos antipredação, o eixo 'y' a escala de tempo em milhões de anos. O P-valor do teste lambda (L) é estatisticamente significativo em um alfa de 0,001.

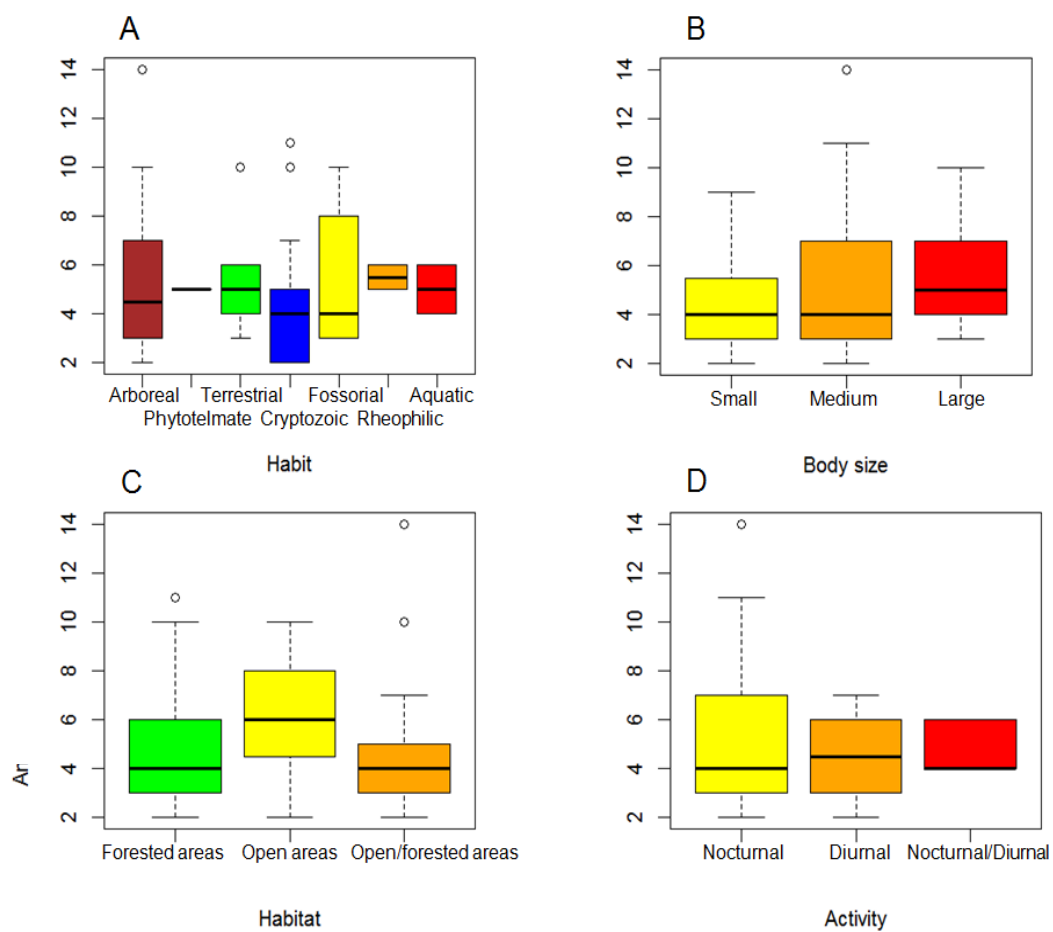
#### 4.4.4 Mecanismos antipredação vs. características funcionais

Os resultados mostraram que as características funcionais explicaram 40,4% da diversidade de mecanismos antipredação de acordo com as análises de PERMANOVA.

O hábito foi responsável por cerca de 25,1%, o tamanho do corpo de 5,2%, a atividade de 5,0% e *habitat* 4,9%. O modo de desenvolvimento reprodutivo não mostrou qualquer relação significativa com a diversidade de mecanismos antipredação (Tabela 3). O Boxplot mostrou que as espécies arbóreas e fossoriais, espécies médias e grandes, espécies noturnas e de áreas abertas possuem maior diversidade de mecanismos antipredação (Figura 7).

**Tabela 3** Associação da diversidade de mecanismos antipredação as características funcionais dos anfíbios da Floresta Atlântica Brasileira pela PERMANOVA.

Variáveis	df	<i>F</i> model	<i>R</i> <sup>2</sup>	<i>P</i> value
Atividade	1	9.463	0.05060	0.002
Tamanho do corpo	1	9.857	0.05271	0.001
Habito	1	47.100	0.25187	0.001
<i>Habitat</i>	1	9.306	0.04976	0.001
Desenvolvimento reprodutivo	1	2.275	0.01217	0.108
Resíduos	109		0.58289	–
Total	114	–	1.00000	–

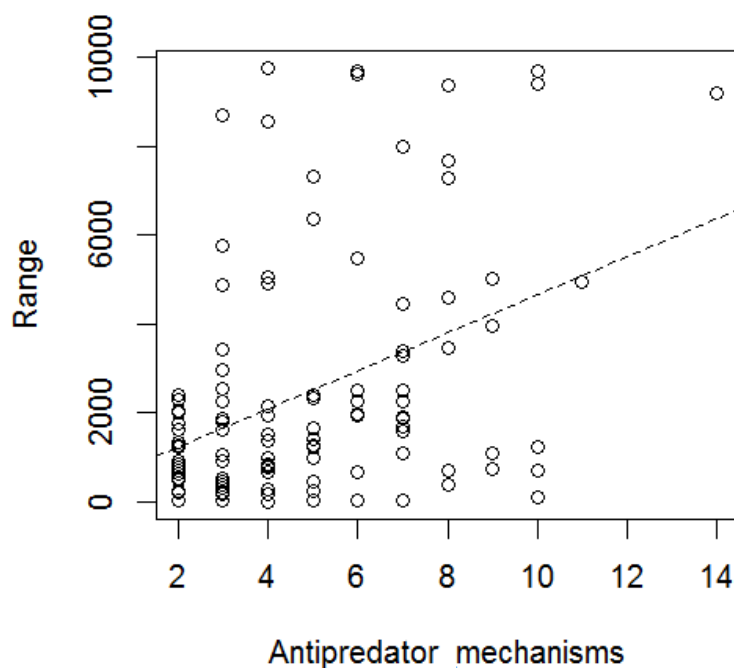


**Figura 7.** O Boxplot mostra a relação entre as características funcionais e a diversidade de mecanismos antipredação.



#### 4.4.5 Distribuição geográfica vs mecanismos antipredação

Os resultados do modelo linear mostraram valores significativos em relação à diversidade de mecanismos antipredação ( $r^2 = 0,15$ ,  $p < 0,001$ ). Espécies que tem mais mecanismos apresentam maior distribuição geográfica na Floresta Atlântica (Figura 8).



**Figura 8.** Relação entre os mecanismos antipredação e a distribuição geográfica de 115 espécies analisadas. Cada range representa uma grade de 10 x 10 km.

#### 4.5 DISCUSSÃO

Mecanismos ancestrais que possuem um forte sinal filogenético (produzir secreções e imobilidade) são simpliomórficos e sua evolução ocorreu de forma independente em diferentes grupos taxonômicos (homoplásticos). Os resultados mostraram que a

evolução destes mecanismos é aleatória, seguindo um modelo de evolução Browniano. Para o mecanismo de produzir secreções, os resultados mostraram que é puramente filogenética e também sugere um nível mais forte nas raízes das árvores do que nas pontas.

De acordo com Brodie (1977, 1983), o mecanismo de produzir secreções pode direcionar a diversidade de mecanismos antipredação. Dependendo da eficiência da secreção que a espécie libera, pode determinar uma redução na diversidade de mecanismos antipredação devido ao sucesso deste mecanismo (Meyers et al. 1978, 1986 Sih, Harvell 1990, 1992 Bronmark e Miner, Yadete et al. 2012). Os dados mostraram que as espécies que produzem secreções podem direcionar a evolução e surgimento de novos mecanismos em sinergia. De acordo com os resultados, famílias com espécies tóxicas geralmente têm mecanismos de contração atuando em sinergia. Por exemplo, as espécies da família Phyllomedusidae têm mecanismos antipredação mais passivos (ou seja, não apresentam contato agressivo com o predador). Para as espécies desta família a combinação dos mecanismos de produzir secreções tóxicas, aposematismo parcial, imobilidade do tipo encolhido, camuflagem e contração parecem ter chegado a um sucesso contra a predação. Entretanto, o mecanismo de produzir secreções do tipo escorregadia (Família Hylodidae), dirige espécies a desenvolverem estratégias sinérgicas pós-predador (lutar, fugir "vários pulos", camuflagem e imobilidade "alerta").

O mecanismo de camuflagem curiosamente não apresenta um forte sinal filogenético, mas uma forte influência sobre o modelo Browniano de evolução. Mecanismos comuns entre Gymnophiona e Anura (como camuflagem e imobilidade) são simplesiomórficos e, possivelmente, os primeiros mecanismos de defesa em

anfíbios. O mecanismo de camuflagem apresenta pouca mudança evolutiva, com poucas espécies totalmente aposemáticas na Floresta Atlântica, enquanto que a imobilidade sofreu alterações mais recentes (como imobilidade do tipo “tanatose”). Ambos os mecanismos (ou seja, camuflagem e imobilidade) têm forte influência a uma evolução aleatória, mas não com um forte sinal, sugerindo influências ecológicas como pressões evolutivas seletoras.

O mecanismo de contração é usado em sinergia com algum tipo de mecanismo de produzir secreções que pode causar ou fornecer a regurgitação da presa pelo predador (Sazima 1974, Toledo et al 2010). Contração é um mecanismo ancestral para as famílias Bufonidae, Phyllomedusidae, Odontophrynidae e Hemipractidae e Subfamília Cophamantinae (sinapomorfia para esta subfamília) e sua evolução é homoplástica (ver Tabela 4 e Figura 3D). No entanto, não apresenta sinais de evolução Browniano, e suas origens são concentradas nas pontas das árvores (Figura 6D). Isto sugere que a evolução deste mecanismo pode ter surgido através da ocorrência na espécie ancestral de um tipo de mecanismo de produzir secreção dos tipos tóxicos ou desagradáveis.

O mecanismo de manter as pernas esticadas apresenta sinal filogenético, mas um fraco sinal de evolução aleatória (Browniano). Este mecanismo é homoplástico para Cycloramphidae, Bufonidae, Microhylidae (sinapomorfia em Gastrophryninae), Leptodactylidae (sinapomorfia em Paratelmatobinae) e Odontophrynidae. Sua evolução parece ser recente surgindo de forma independente (homoplásticos) nas diferentes famílias e subfamílias de anuros (mecanismo apomórfico).

De acordo com Gingras e Fitch (2013) o canto de anúncio tem forte relação filogenética entre os gêneros de anuros. Entretanto, os dados não mostram esse padrão para o mecanismo de vocalização defensiva. Este mecanismo é um estado apomórfico com origem homoplástica em Cycloramphidae, Bufonidae, Microhylidae (Gastrophryninae), Leptodactylidae (Leiuperinae) e Hylidae (Scinaxinae e Cophomantinae). O mecanismo de vocalização defensiva "alarme" foi registrado em estudos recentes (Toledo et al. 2015 e Lourenço-de-Moraes et al. 2016), sugerindo que os mecanismos de vocalização defensiva tenham origens etológicas. Este mecanismo é uma sinapomorfia nas subfamílias Comophantinae (Figura 4D) e Hemiphractinae (veja Toledo et al. 2015; Lourenço-de-Moraes et al. 2016).

O mecanismo de abrir a boca é um estado apomórfico e homólogo em Terrarana (Brachycephalidae, Craugastoridae e Eleutherodactylidae). Este grupo tem espécies diminutas e seus principais predadores são os artrópodes (Toledo 2005; Lourenço-de-Moraes e Pertel 2011). Este mecanismo deve desorientar estes pequenos predadores; com um forte sinal filogenético apresenta alta tendência a um modelo de evolução Browniano concentrado nas raízes da árvore em Terrarana (Figura 4C e 6K). A miniaturização das espécies deste grupo pode ter influenciado origem deste mecanismo (Hedges et al. 2008; Carvalho et al. 2011). No entanto, esse comportamento também foi observado (mas não foi incluído na nossa árvore) em famílias de espécies maiores como Hemiphractidae e Ceratophryidae (Duellman e Trueb 1994; Toledo et al. 2011). Mas esse mecanismo geralmente é exibido por espécies maiores aparecendo em sinergia com os mecanismos de inflar o corpo, lutar (do tipo morder) e vocalização defensiva (ver Toledo et al. 2011; Lourenço-de-Moraes et al. 2016). Em espécies diminutas de Terrarana estes mecanismos não aparecem em sinergia com outros comportamentos

(veja Lourenço-de-Moraes et al. 2014b) que sugerem que este mecanismo evoluiu de forma homoplástica nos diferentes grupos.

Os mecanismos sem sinais filogenéticos (inflar o corpo e contração) também não seguem um modelo de evolução aleatória, estes mecanismos evoluíram de forma independente. Estes mecanismos parecem ser recentes em termos evolutivos (ver Figura 6 D, F) principalmente o mecanismo de contração. No entanto, aparecem como mecanismos ancestrais surgidos de forma homoplástica em algumas famílias e subfamílias. O surgimento de mecanismos com origens filogenéticas recentes sugere origens etológicas. A capacidade de memorizar um determinado recurso defensivo que obteve sucesso é que ajuda o animal a manter este comportamento, está presente mesmo em invertebrados (Punzo 2004). Demonstrando a capacidade de aprendizagem em resposta a um estímulo de ameaça, essa característica seletora é necessária para a sobrevivência da presa. Espécies que vivem próximas apresentam repertório similar de mecanismos antipredação (presente estudo), mas ainda são necessários mais testes.

Os resultados revelam que o mecanismo de lutar apresenta um sinal filogenético fraco e um valor baixo para uma evolução Browniano. Toledo e colaboradores (2011) sugeriram que o mecanismo de lutar do tipo morder pode estar relacionado ao cuidado parental. Lourenço-de-Moraes e colaboradores (2016) encontraram evidências de que morder pode realmente ser de cuidado parental. Figueiredo e colaboradores (no prelo) registraram uma fêmea grávida de *Ischnocnema henseli* realizando o mecanismo de lutar do tipo “morder”. Esta espécie apresenta desenvolvimento reprodutivo direto, espécies deste grupo foram encontradas desempenhando cuidado parental (Townsend et al. 1996; Ryan et al. 2010). É possível que este mecanismo tenha origem etológica com base em pressões ecológicas devido às suas características filogenéticas ancestrais. A

família Cycloramphidae tem o mecanismo de lutar do tipo “morder” com origem homoplástica.

As especializações ecológicas das espécies de anfíbios apresentam de um modo geral, uma forte pressão evolutiva em relação aos seus mecanismos antipredação. Os resultados mostraram que características funcionais explicam 40,4% da diversidade de mecanismos. A característica funcional "hábito" foi a que mais explicou a diversidade de mecanismos antipredação. As características funcionais podem ajudar a explicar a distribuição das espécies (Gaston 1990; Lawton 1993; Diaz et al. 2007; Gomes-Rodrigues 2015), portanto, as espécies que estão mais expostas aos predadores, como as espécies arbóreas (Figura 7A), conseqüentemente apresentam maior diversidade de predadores, como aves, répteis, mamíferos e invertebrados (Toledo 2005, 2007; Pombal Jr 2007), devido a isso espera-se que esta pressão ecológica influencie as respostas rápidas e eficazes para a sobrevivência da espécie selecionando os mais aptos. Espécies fossoriais apresentam redução dos membros posteriores que foram adaptados a escavação (Haddad et al. 2013), e por causa disso, perderam uma das principais defesas das espécies terrestres, como o mecanismo de fugir com vários saltos. Devido a isso, eles desenvolveram uma maior diversidade de mecanismos antipredação (Figura 7A). Os resultados mostram que as espécies com tamanho corporal médio e grande têm maior diversidade de mecanismos antipredação. Isto sugere que estas espécies, devido ao tamanho, se tornam mais expostas a predação que as espécies pequenas e portanto, desenvolveram uma maior diversidade de mecanismos antipredação, bem como espécies de áreas abertas.

É amplamente documentada a influência das características funcionais na dispersão de espécies, tais como o tamanho do corpo e especializações ecológicas

(Brown e Maurer 1989; Bell 2001). Os resultados confirmam a hipótese sugerida por Lourenço-de-Moraes et al. (2016) de que as espécies com uma diversidade de mecanismos antipredação apresentam maior distribuição geográfica. Uma das principais barreiras bióticas para dispersão dos anfíbios são o número de predadores (Toledo et al. 2007) uma maior diversidade de mecanismos antipredação fornece a uma presa em potencial maior capacidade de sobreviver a diversidade de predadores que consomem anfíbios.

Apesar das diferenças de mecanismos antipredação que podem ocorrer entre populações, e alguns indivíduos podem apresentar mais mecanismos do que outros (com. pessoal), as características filogenéticas das espécies podem influenciar ou direcionar espécies de diferentes populações, mas sob pressões ecológicas semelhantes a apresentarem o mesmo comportamento, isso devido à alta plasticidade dos anfíbios (Lawler 1989; Horat e Semlitsch 1994; Relyea 2001; Urban et al. 2014).

Os resultados mostram que a origem dos mecanismos antipredação e sua diversificação é altamente plástico e rápido. As diferentes estratégias entre os anfíbios proporcionam maior sobrevivência aos predadores e, conseqüentemente, a dispersão das espécies. Especializações as várias características funcionais, agem como uma forte pressão seletiva e adaptativa em anfíbios. Mecanismos antipredação podem ter origem puramente filogenética ou origens etológicas/ecológicas selecionados por um processo de “seleção natural”, através da aprendizagem em resposta a possíveis predadores no passado. Sendo assim fornecemos novas perspectivas sobre os mecanismos antipredação e a plasticidade dos anfíbios.

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## 5 CONCLUSÕES GERAIS

Neste trabalho confirmamos os efeitos das alterações antrópicas na distribuição das espécies de anfíbios na Floresta Atlântica. A riqueza de espécies está associada a características abióticas ideais, devido às suas características fisiológicas dependentes de umidade e temperatura amenas e relacionada com eventos históricos como o Pleistoceno glacial. Espécies menores estão relacionadas às suas sensibilidades fisiológicas às alterações abióticas, e espécies maiores e mais adaptadas têm maior dispersão do que as espécies menores. Em média, as áreas com maiores altitudes possuem menores espécies e endêmicas, ao passo que áreas com menores latitudes possuem média de espécies maiores. As análises de diversidade filogenética e funcional mostraram os principais *hotspots* são a Serra do Mar e o Corredor Central da Floresta Atlântica e confirmaram que as áreas de altitude podem servir como um refúgio climático para as espécies em um cenário de aquecimento global durante o Antropoceno. Além disso, mostramos alta relação entre diversidade filogenética e funcional respondendo da mesma forma quanto à riqueza de espécies e distribuição. Apresentamos também, a origem dos mecanismos antipredação, bem como a influência das adaptações ecológicas na diversidade de mecanismos bem como a diversidade destes mecanismos influenciando na distribuição das espécies de anfíbios na ameaçada Floresta Atlântica brasileira. Esta tese demonstra principalmente os diversos fatores bióticos e abióticos que trabalhando em sinergia explicam a riqueza, origem e permanência das espécies nas florestas do Bioma Floresta Atlântica.

APÊNDICE 1- **Table S1.** List of species studied in alphabetical order with their respective distribution area (range = grid of 10 x 10 km) and functional traits.

Species	Range	Activity	Body size (cm)	Poisonous	Habit	Habitat	Development mode	References
<i>Adelophryne glandulata</i>	3	Nocturnal	1.2	Non-toxic	Cryptozoic	Forest areas	Direct	Lourenço-de-Moraes <i>et al.</i> , 2014; present study
<i>Adelophryne meridionalis</i>	2	Nocturnal	1.1	Non-toxic	Cryptozoic	Forest areas	Direct	Santana <i>et al.</i> , 2012; present study
<i>Adelophryne mucronatus</i>	38	Nocturnal	1.4	Non-toxic	Cryptozoic	Forest areas	Direct	Lourenço-de-Moraes <i>et al.</i> , 2012a; Haddad <i>et al.</i> , 2013; present study
<i>Adelophryne pachydactyla</i>	301	Nocturnal	1.4	Non-toxic	Cryptozoic	Forest areas	Direct	Lourenço-de-Moraes <i>et al.</i> , 2012; Haddad <i>et al.</i> , 2013; present study
<i>Adenomera ajurauna</i>	27	Nocturnal	1.9	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Adenomera araucaria</i>	66	Nocturnal	1.9	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Adenomera bokermanni</i>	2624	Nocturnal	2.4	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Adenomera engelsi</i>	45	Diurnal/nocturnal	2.2	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Adenomera marmorata</i>	2378	Diurnal/nocturnal	2.1	Non-toxic	Cryptozoic	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Adenomera nana</i>	224	Nocturnal	1.8	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Adenomera thomei</i>	299	Nocturnal	2.4	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Allobates alagoanus</i>	6	Diurnal	1.5	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Allobates</i>	578	Diurnal	1.6	Non-toxic	Rheophilic	Open and	Indirect	Haddad <i>et al.</i> , 2013; present study

<i>capixaba</i>						Forest areas		
<i>Allobates olfersioides</i>	2322	Diurnal	1.6	Non-toxic	Rheophilic	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013; Lourenço-de-Moraes <i>et al.</i> , 2013; present study
<i>Aparasphenodon arapapa</i>	842	Nocturnal	4.5	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; Lourenço-de-Moraes <i>et al.</i> , 2013; present study
<i>Aparasphenodon bokermanni</i>	2	Nocturnal	4.6	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Aparasphenodon brunoi</i>	2532	Nocturnal	8	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Aplastodiscus albofrenatus</i>	435	Nocturnal	3.9	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Aplastodiscus albosignatus</i>	1077	Nocturnal	4.3	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Aplastodiscus arildae</i>	1903	Nocturnal	3.7	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Aplastodiscus callipygius</i>	404	Nocturnal	4.6	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Aplastodiscus cavicola</i>	587	Nocturnal	3.7	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Aplastodiscus cochranæ</i>	173	Nocturnal	4.5	Unpalatable or bad odour	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Aplastodiscus ehrhardti</i>	395	Nocturnal	3.1	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Aplastodiscus eugenioi</i>	269	Nocturnal	3.5	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Aplastodiscus flumineus</i>	9	Nocturnal	4.5	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Aplastodiscus ibirabitanga</i>	519	Nocturnal	4.1	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study

<i>Aplastodiscus leucoeygius</i>	1588	Nocturnal	4.5	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Aplastodiscus musicus</i>	13	Nocturnal	5	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Lutz 1949; Cruz <i>et al.</i> , .2004; present study
<i>Aplastodiscus perviridis</i>	5781	Nocturnal	4.5	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Aplastodiscus sibilatus</i>	271	Nocturnal	3.2	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Aplastodiscus weygoldti</i>	34	Nocturnal	3.8	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Arcovomer passarellii</i>	779	Nocturnal	2.4	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Bokermannohyla ahenea</i>	2	Nocturnal	5.2	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Bokermannohyla astartea</i>	1101	Nocturnal	4.4	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Bokermannohyla capra</i>	32	Nocturnal	6.1	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Bokermannohyla caramaschii</i>	358	Nocturnal	6.2	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Bokermannohyla carvalhoi</i>	168	Nocturnal	6.4	Unpalatable or bad odour	Terrestrial	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Bokermannohyla circumdata</i>	1109	Nocturnal	5.4	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Bokermannohyla claresignata</i>	101	Nocturnal	6.1	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Lutz and Lutz 1939; Carvalho-e-Silva and Caramaschi 2004; present study
<i>Bokermannohyla clepsydra</i>	67	Nocturnal	4	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Lutz 1925; Carvalho-e-Silva and Telles 2004a; present study
<i>Bokermannohyla diamantina</i>	6	Nocturnal	4.9	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013

<i>Bokermannohyla gouveai</i>	41	Nocturnal	6.9	Unpalatable or bad odour	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Bokermannohyla hylax</i>	979	Nocturnal	6.6	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Bokermannohyla ibitipoca</i>	2	Nocturnal	3.9	Unpalatable or bad odour	Terrestrial	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Bokermannohyla itapoty</i>	25	Nocturnal	4.1	Unpalatable or bad odour	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Bokermannohyla izecksohni</i>	2	Nocturnal	3.1	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Bokermannohyla langei</i>	4	Nocturnal	6.6	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Bokermann 1965; Segalla and Silvano 2004; present study
<i>Bokermannohyla luciana</i>	1	Nocturnal	4.7	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Bokermannohyla luctuosa</i>	450	Nocturnal	6.7	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Bokermannohyla martinsi</i>	224	Nocturnal	7.3	Unpalatable or bad odour	Terrestrial	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Bokermannohyla nanuzae</i>	121	Nocturnal	4.4	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Bokermannohyla oxente</i>	1	Nocturnal	4.9	Unpalatable or bad odour	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Brachycephalus alipioi</i>	9	Diurnal	1.4	Toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Brachycephalus auroguttatus</i>	4	Diurnal	1.3	Toxic	Cryptozoic	Forest areas	Direct	Ribeiro <i>et al.</i> , 2015
<i>Brachycephalus boticario</i>	2	Diurnal	1.2	Toxic	Cryptozoic	Forest areas	Direct	Ribeiro <i>et al.</i> , 2015
<i>Brachycephalus brunneus</i>	1	Diurnal	1.2	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013

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<i>Brachycephalus didactylus</i>	474	Diurnal	0.8	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Brachycephalus ephippium</i>	1863	Diurnal	1.9	Toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013; present study
<i>Brachycephalus ferruginus</i>	16	Diurnal	1.4	Toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Brachycephalus fuscolineatus</i>	2	Diurnal	1.2	Non-toxic	Cryptozoic	Forest areas	Direct	Ribeiro <i>et al.</i> , 2015
<i>Brachycephalus garbeanus</i>	9	Diurnal	1.7	Toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Brachycephalus guarani</i>	23	Diurnal	1.3	Toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Brachycephalus hermogenesi</i>	653	Diurnal	1.1	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Brachycephalus izecksohni</i>	4	Diurnal	1.3	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Brachycephalus leopardus</i>	2	Diurnal	1.2	Toxic	Cryptozoic	Forest areas	Direct	Ribeiro <i>et al.</i> , 2015
<i>Brachycephalus margaritatus</i>	10	Diurnal	1.7	Toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Brachycephalus mariaterezae</i>	2	Diurnal	1.3	Toxic	Cryptozoic	Forest areas	Direct	Ribeiro <i>et al.</i> , 2015
<i>Brachycephalus nodoterga</i>	6	Diurnal	1.3	Toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Brachycephalus olivaceus</i>	2	Diurnal	1.3	Toxic	Cryptozoic	Forest areas	Direct	Ribeiro <i>et al.</i> , 2015
<i>Brachycephalus pernix</i>	2	Diurnal	1.5	Toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Brachycephalus pitanga</i>	8	Diurnal	1.3	Toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013

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<i>Brachycephalus pombali</i>	10	Diurnal	1.5	Toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Brachycephalus pulex</i>	10	—	0.8	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Brachycephalus toby</i>	7	Diurnal	1.4	Toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Brachycephalus tridactylus</i>	14	Diurnal	1.4	Toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Brachycephalus verrucosus</i>	2	Diurnal	1.3	Toxic	Cryptozoic	Forest areas	Direct	Ribeiro <i>et al.</i> , 2015
<i>Brachycephalus vertebralis</i>	66	Diurnal	1.4	Toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Ceratophrys aurita</i>	5003	Nocturnal	14.9	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Chiasmocleis alagoana</i>	5	Nocturnal	2.7	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Chiasmocleis atlantica</i>	562	Nocturnal	3.2	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Chiasmocleis capixaba</i>	495	Nocturnal	1.5	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Chiasmocleis carvalhoi</i>	741	Nocturnal	2.3	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Chiasmocleis cordeiroi</i>	1	Nocturnal	2.2	Non-toxic	Cryptozoic	Forest areas	Indirect	Caramaschi and Pimenta 2003, 2004
<i>Chiasmocleis crucis</i>	1	Nocturnal	2	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Chiasmocleis gnoma</i>	1	Nocturnal	1.6	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Chiasmocleis leucosticta</i>	1533	Diurnal/nocturnal	2.5	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013

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<i>Chiasmocleis mantiqueira</i>	2	Nocturnal	2.2	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Chiasmocleis sapiranga</i>	3	Nocturnal	2.5	Non-toxic	Cryptozoic	Forest areas	Indirect	Cruz <i>et al.</i> , 2007; Ângulo 2008
<i>Chiasmocleis schubarti</i>	1398	Nocturnal	3.3	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Chthonerpeton indistinctum</i>	528	—	43.5	—	Aquatic	Open and forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Chthonerpeton noctinectes</i>	3	—	34.5	—	Aquatic	Open and forest areas	Direct	Silva <i>et al.</i> , 2003; Wilkinson 2004
<i>Chthonerpeton viviparum</i>	12	—	55.8	—	Aquatic	Open and forest areas	Direct	Taylor 1968; Measey <i>et al.</i> , 2004
<i>Crossodactylodes bokermanni</i>	36	Nocturnal	1.4	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Crossodactylodes izecksohni</i>	4	Nocturnal	1.5	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Crossodactylodes pinto</i>	4	Nocturnal	1.7	Non-toxic	Arboreal	Forest areas	Indirect	Cochran 1938; Peixoto <i>et al.</i> , 2004
<i>Crossodactylus aeneus</i>	1212	—	2.3	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Crossodactylus bokermanni</i>	2	—	2.7	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Crossodactylus caramaschii</i>	672	Diurnal/nocturnal	2.3	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Crossodactylus cyclopinus</i>	8	—	2.9	Non-toxic	Rheophilic	Forest areas	Indirect	Nascimento and Cruz 2005; Ângulo 2008
<i>Crossodactylus dantei</i>	2	—	2.2	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Crossodactylus dispar</i>	1187	Diurnal	3.1	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013

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<i>Crossodactylus gaudichaudii</i>	783	Diurnal/nocturnal	3.1	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Crossodactylus grandis</i>	133	Diurnal	4.2	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Crossodactylus lutzorum</i>	4	—	2.6	Non-toxic	Rheophilic	Forest areas	Indirect	Carcereali and Caramaschii 1993; Silvano and 2004b; Pimenta <i>et al.</i> , 2014
<i>Crossodactylus schmidti</i>	255	Nocturnal	2.8	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Crossodactylus trachystomus</i>	25	—	2.5	Non-toxic	Rheophilic	Forest areas	Indirect	Cochran 1955; Pimenta and Nascimento 2010
<i>Cycloramphus acangatan</i>	125	Nocturnal	4.3	—	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Cycloramphus asper</i>	188	Nocturnal	3.5	—	Rheophilic	Forest areas	Indirect	Cochran 1955; Heyer <i>et al.</i> , 2004
<i>Cycloramphus bandeirensis</i>	5	—	4.5	—	Rheophilic	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Cycloramphus bolitoglossus</i>	306	Nocturnal	3.8	—	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Cycloramphus boraceiensis</i>	571	Nocturnal	4.7	—	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Cycloramphus brasiliensis</i>	340	Nocturnal	6.2	—	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Cycloramphus carvalhoi</i>	5	—	5.6	—	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Cycloramphus catarinensis</i>	2	Nocturnal	3.5	—	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Cycloramphus cedrensis</i>	9	—	4.9	—	Rheophilic	Forest areas	Indirect	Heyer 1983; Heyer and Silvano 2004a
<i>Cycloramphus diringshofeni</i>	53	—	3.1	—	Rheophilic	Forest areas	Indirect	Bokermann 1965; Heyer <i>et al.</i> , 2004

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<i>Cycloramphus dubius</i>	32	Nocturnal	4.9	—	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Cycloramphus duseni</i>	9	—	3.7	—	Rheophilic	Forest areas	Indirect	Heyer 1983; Verdade and Segalla 2004
<i>Cycloramphus eleutherodactylus</i>	1891	Nocturnal	4.8	—	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Cycloramphus fuliginosus</i>	1052	Nocturnal	3.3	—	Rheophilic	Forest areas	Indirect	Cochran 1955; Verdade and Carvalho-e-Silva 2004
<i>Cycloramphus granulosis</i>	104	—	4.2	—	Rheophilic	Forest areas	Indirect	Bokermann 1951; Heyer and Verdade 2004
<i>Cycloramphus izecksohni</i>	226	Nocturnal	4	—	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Cycloramphus juimirim</i>	5	Nocturnal	3	—	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Cycloramphus lithomimeticus</i>	62	Nocturnal	3.1	—	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Cycloramphus lutzorum</i>	634	Nocturnal	4.1	—	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Cycloramphus migueli</i>	24	—	4.2	—	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Cycloramphus mirandaribeiroi</i>	4	—	6.9	—	Rheophilic	Forest areas	Indirect	Heyer 1983; Heyer and Silvano 2004b
<i>Cycloramphus ohausi</i>	17	—	3.8	—	Rheophilic	Forest areas	Indirect	Cochran 1955; Rodrigues and Carvalho-e-Silva 2004b
<i>Cycloramphus organensis</i>	31	Nocturnal	3.2	—	Rheophilic	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Cycloramphus rhyakonastes</i>	17	Nocturnal	4.2	—	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Cycloramphus semipalmatus</i>	108	—	5	—	Rheophilic	Forest areas	Indirect	Heyer 1983; Verdade and Heyer 2004a

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<i>Cycloramphus stejneri</i>	17	—	5.6	—	Cryptozoic	Forest areas	Indirect	Heyer 1983; Verdade and Heyer 2004b
<i>Cycloramphus valae</i>	96	—	3.6	—	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Dasylops schirchi</i>	186	Nocturnal	5.2	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Dendrophryniscus berthaltutzae</i>	374	Nocturnal	2.4	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Dendrophryniscus brevipollicatus</i>	758	Nocturnal	2	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Dendrophryniscus carvalhoi</i>	31	—	1.9	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Dendrophryniscus krausae</i>	8	—	2.3	Non-toxic	Fossorial	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Dendrophryniscus leucomystax</i>	905	Nocturnal	2.5	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Dendrophryniscus oreites</i>	29	—	2.5	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Dendrophryniscus proboscideus</i>	1353	Nocturnal	4.1	Toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Dendrophryniscus stawiarskyi</i>	2	—	2.2	—	—	Forest areas	Indirect	Izecksohn 1994; Silvano <i>et al.</i> , 2004
<i>Dendropsophus anceps</i>	2294	Nocturnal	4.2	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Dendropsophus berthaltutzae</i>	998	Nocturnal	2.4	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Dendropsophus bipunctatus</i>	1622	Nocturnal	2.6	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Dendropsophus branneri</i>	3285	Nocturnal	2.3	Unpalatable or bad odour	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study

<i>Dendropsophus decipiens</i>	4178	Nocturnal	1.9	Non-toxic	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Dendropsophus dutrai</i>	44	Nocturnal	3.8	—	Arboreal	Forest areas	Indirect	Gomes and Peixoto 1996; Silvano and Peixoto 2004a
<i>Dendropsophus elegans</i>	5047	Nocturnal	3.1	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Dendropsophus giesleri</i>	855	Nocturnal	2.7	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Dendropsophus haddadi</i>	1029	Nocturnal	2.3	Non-toxic	Arboreal	Open and Forest areas	Indirect	Lourenço-de-Moraes <i>et al.</i> , 2012b; Haddad <i>et al.</i> , 2013; present study
<i>Dendropsophus limai</i>	4	Nocturnal	1.9	—	Arboreal	Forest areas	Indirect	Bokermann 1962; Carvalho-e-Silva 2004
<i>Dendropsophus meridianus</i>	248	Nocturnal	1.8	Non-toxic	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Dendropsophus microps</i>	3122	Nocturnal	2.9	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Dendropsophus minutus</i>	9676	Nocturnal	2.5	Unpalatable or bad odour	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Dendropsophus nahderere</i>	288	Nocturnal	4.6	Non-toxic	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Dendropsophus nanus</i>	7282	Nocturnal	2.3	Unpalatable or bad odour	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Dendropsophus novaisi</i>	40	Nocturnal	3.4	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Dendropsophus oliveirai</i>	811	Nocturnal	1.9	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Dendropsophus pseudomeridianus</i>	200	Nocturnal	1.7	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Dendropsophus ruschii</i>	26	Nocturnal	2.8	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013

<i>Dendropsophus sanorni</i>	3410	Nocturnal	2	Unpalatable or bad odour	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Dendropsophus seniculus</i>	2681	Nocturnal	3.8	Non-toxic	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Dendropsophus soaresi</i>	315	Nocturnal	3.5	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Dendropsophus studerae</i>	2	Nocturnal	2.8	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Dendropsophus weneri</i>	923	Nocturnal	1.9	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Elaschitocleis bicolor</i>	3457	Nocturnal	3.8	Non-toxic	Fossorial	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Elaschitocleis cesarii</i>	7121	Nocturnal	3.2	Non-toxic	Fossorial	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Elaschitocleis erythrogaster</i>	37	Nocturnal	3.5	Non-toxic	Fossorial	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Eleutherodactylus bilineatus</i>	550	Nocturnal	2.6	Non-toxic	Fossorial	Forest areas	Direct	Haddad <i>et al.</i> , 2013; present study
<i>Euparkerella brasiliensis</i>	369	Diurnal/nocturnal	1.7	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Euparkerella cochranae</i>	186	Diurnal/nocturnal	1.6	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013; present study
<i>Euparkerella cryptica</i>	2	Diurnal/nocturnal	1.9	Non-toxic	Cryptozoic	Forest areas	Direct	Izecksohn 1998
<i>Euparkerella robusta</i>	2	Diurnal/nocturnal	2.3	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Euparkerella tridactyla</i>	2	Diurnal/nocturnal	1.5	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Fritziana fissilis</i>	934	Nocturnal	2.8	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study

<i>Fritziana goeldii</i>	1825	Nocturnal	3.7	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Fritziana ohausi</i>	977	Nocturnal	3.1	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Frostius erythropthalmus</i>	435	Nocturnal	2.3	Toxic	Fossorial	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Frostius pernambucensis</i>	1008	Nocturnal	2.1	Toxic	Fossorial	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Gastrotheca albolineata</i>	703	Nocturnal	6	Non-toxic	Arboreal	Forest areas	Direct	Haddad <i>et al.</i> , 2013; present study
<i>Gastrotheca ernestoi</i>	321	Nocturnal	6.9	Non-toxic	Arboreal	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Gastrotheca fissipes</i>	1550	Nocturnal	7.2	Non-toxic	Arboreal	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Gastrotheca flamma</i>	1	Nocturnal	5.5	Non-toxic	Arboreal	Forest areas	Direct	Juncá and Nunes 2008
<i>Gastrotheca fulvorufa</i>	495	Nocturnal	6.9	Non-toxic	Arboreal	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Gastrotheca megacephala</i>	895	Nocturnal	10.1	Non-toxic	Arboreal	Forest areas	Direct	Haddad <i>et al.</i> , 2013; Lourenço-de-Moraes <i>et al.</i> , 2016; present study
<i>Gastrotheca microdiscus</i>	1645	Nocturnal	5.9	Non-toxic	Arboreal	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Gastrotheca prasina</i>	12	Nocturnal	9.7	Non-toxic	Arboreal	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Gastrotheca pulchra</i>	5	Nocturnal	3.3	Non-toxic	Arboreal	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Gastrotheca recava</i>	6	Nocturnal	7.6	Non-toxic	Arboreal	Forest areas	Direct	Haddad <i>et al.</i> , 2013; Lourenço-de-Moraes <i>et al.</i> , 2016; present study
<i>Haddadus aramunha</i>	2	Nocturnal	6.2	Non-toxic	Cryptozoic	Open areas	Direct	Haddad <i>et al.</i> , 2013
<i>Haddadus</i>	4965	Nocturnal	6.5	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013; present study

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<i>binotatus</i>								
<i>Haddadus plicifer</i>	1	Nocturnal	1.7	—	—	—	Direct	Hedges <i>et al.</i> , 2008
<i>Holoaden bradei</i>	26	—	2.5	Toxic	Cryptozoic	Open and Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Holoaden luederwaldti</i>	473	Nocturnal	4.6	Toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Holoaden pholeter</i>	14	Nocturnal	4.6	Toxic	Cryptozoic	Open and Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Hylodes amnicola</i>	8	Diurnal	2.7	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hylodes asper</i>	967	Diurnal	4.7	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Hylodes babax</i>	32	Diurnal	3.2	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hylodes cardosoi</i>	65	Diurnal	4	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hylodes charadranaetes</i>	73	Diurnal	4.3	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hylodes dactylocinus</i>	19	Diurnal	2.7	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Hylodes fredii</i>	1	Diurnal	3.7	Non-toxic	Rheophilic	Forest areas	Indirect	Canedo and Pombal 2007
<i>Hylodes glaber</i>	7	Diurnal	2.9	Non-toxic	Rheophilic	Forest areas	Indirect	Miranda-Ribeiro 1926; Rocha <i>et al.</i> , 2004 <sup>a</sup>
<i>Hylodes heyeri</i>	291	Diurnal	4.3	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hylodes lateristrigatus</i>	989	Diurnal	3.5	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hylodes magalhaesi</i>	46	Diurnal	3.4	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hylodes meridionalis</i>	102	Diurnal	4.8	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Hylodes mertensi</i>	8	Diurnal	5.4	Non-toxic	Rheophilic	Forest areas	Indirect	Bokermann 1956; Rocha <i>et al.</i> , 2004b
<i>Hylodes nasus</i>	1510	Diurnal	3.8	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hylodes ornatus</i>	318	Diurnal	2.7	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013

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<i>Hylodes otavioi</i>	7	Diurnal	3.4	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hylodes perere</i>	30	Diurnal	2.7	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hylodes perplicatus</i>	83	Diurnal	4.4	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hylodes phyllodes</i>	468	Diurnal	3	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hylodes pipilans</i>	5	Diurnal	2.8	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Hylodes regius</i>	61	Diurnal	3.5	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hylodes sazimai</i>	228	Diurnal	2.8	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hylodes uai</i>	24	Diurnal	3.8	Non-toxic	Rheophilic	Forest areas	Indirect	Nascimento <i>et al.</i> , 2001; Nascimento and Pimenta 2010
<i>Hylodes vanzolinii</i>	4	Diurnal	2.9	Non-toxic	Rheophilic	Forest areas	Indirect	Heyer 1982; Heyer and Peixoto 2004
<i>Hylomantis aspera</i>	234	Nocturnal	4.4	Toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hylomantis granulosa</i>	383	Nocturnal	4.8	Toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hyophryne histrio</i>	9	Nocturnal	4.8	—	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hypsiboas albomarginatus</i>	4594	Nocturnal	5.8	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Hypsiboas albopunctatus</i>	7663	Nocturnal	6.5	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Hypsiboas atlanticus</i>	1007	Nocturnal	4	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Hypsiboas bischoffi</i>	2422	Nocturnal	5.9	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Hypsiboas caingua</i>	1241	Nocturnal	3.9	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Hypsiboas caipora</i>	14	Nocturnal	4	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hypsiboas crepitans</i>	7978	Nocturnal	6.2	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study

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<i>Hypsiboas curupi</i>	121	Nocturnal	4.5	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hypsiboas cymbalum</i>	8	Nocturnal	4.9	Non-toxic	Arboreal	Open areas	Indirect	Bokermann 1963; Garcia and Rodrigues 2004
<i>Hypsiboas exastis</i>	47	Nocturnal	9.2	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Hypsiboas faber</i>	9183	Nocturnal	8.9	Non-toxic	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Hypsiboas freicanecae</i>	1	Nocturnal	4	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hypsiboas guentheri</i>	373	Nocturnal	4.6	Non-toxic	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hypsiboas joaquina</i>	158	Nocturnal	5.4	Non-toxic	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hypsiboas latistriatus</i>	24	Nocturnal	4.7	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hypsiboas leptolineatus</i>	1499	Nocturnal	3.3	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hypsiboas lundii</i>	3083	Nocturnal	6.9	Non-toxic	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hypsiboas marginatus</i>	205	Nocturnal	5.3	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hypsiboas pardalis</i>	2279	Nocturnal	6.8	Non-toxic	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hypsiboas poaju</i>	7	Nocturnal	3.9	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hypsiboas polytaenius</i>	1685	Nocturnal	3.6	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hypsiboas pombali</i>	1055	Nocturnal	5.7	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Hypsiboas prasinus</i>	2974	Nocturnal	4.8	Non-toxic	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study

<i>Hypsiboas pulchellus</i>	2241	Nocturnal	4.5	Non-toxic	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hypsiboas punctatus</i>	6355	Nocturnal	3.9	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hypsiboas raniceps</i>	3978	Nocturnal	7.1	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Hypsiboas secedens</i>	129	Nocturnal	4.9	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hypsiboas semiguttatus</i>	222	Nocturnal	4.1	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hypsiboas semilineatus</i>	5012	Nocturnal	5.6	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Hypsiboas stellae</i>	164	Nocturnal	4.6	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Hypsiboas stenocephalus</i>	7	Nocturnal	2.7	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Ischnocnema lactea</i>	2020	Nocturnal	3.2	Non-toxic	Cryptozoic	Forest areas	Direct	Miranda-Ribeiro 1923; Hedges <i>et al.</i> , 2008; present study
<i>Ischnocnema paranaensis</i>	1	—	1.8	Non-toxic	Cryptozoic	Forest areas	Direct	Langone and Segalla 1996; Hedges <i>et al.</i> , 2008
<i>Ischnocnema abdita</i>	23	—	1.6	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Ischnocnema bolbodactyla</i>	410	—	1.9	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Ischnocnema concolor</i>	4	Nocturnal	2	Non-toxic	Cryptozoic	Open and Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Ischnocnema epipeda</i>	2	—	2.9	Non-toxic	Cryptozoic	Forest areas	Direct	Heyer 1984; Silvano and Peixoto 2004b
<i>Ischnocnema erythromera</i>	35	Nocturnal	3	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013; present study

<i>Ischnocnema gehrti</i>	1	—	1.5	Non-toxic	Cryptozoic	Forest areas	Direct	Pombal and Cruz 1999; Van Luiz and Cruz 2004; Hedges <i>et al.</i> , 2008
<i>Ischnocnema gualteri</i>	19	—	4	Non-toxic	Cryptozoic	Forest areas	Direct	Lutz 1974; Carvalho-e-Silva <i>et al.</i> , 2004; present study
<i>Ischnocnema guentheri</i>	3635	Nocturnal	4	Non-toxic	Fossorial	Forest areas	Direct	Haddad <i>et al.</i> , 2013; present study
<i>Ischnocnema henselii</i>	1233	Nocturnal	3.6	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013; present study
<i>Ischnocnema hoehnei</i>	422	—	2.9	Non-toxic	Fossorial	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Ischnocnema holti</i>	6	Nocturnal	1.7	Non-toxic	Fossorial	Open and Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Ischnocnema izecksohni</i>	82	—	4.7	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Ischnocnema juipoca</i>	2010	—	2.3	Non-toxic	Cryptozoic	Open and Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Ischnocnema karst</i>	7	—	2.3	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Ischnocnema manezinho</i>	247	Nocturnal	3.3	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Ischnocnema melanopygia</i>	1	—	2	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Ischnocnema nasuta</i>	3936	—	3.9	Non-toxic	Fossorial	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Ischnocnema nigriventris</i>	4	—	1.9	Non-toxic	Fossorial	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Ischnocnema octavioi</i>	342	—	3.2	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Ischnocnema oea</i>	2	Nocturnal	2.2	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Ischnocnema</i>	1423	Nocturnal	2	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013; present study

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<i>parva</i>								
<i>Ischnocnema pusilla</i>	11	Nocturnal	1.6	Non-toxic	Cryptozoic	Forest areas	Direct	Bokermann 1967; Cruz and Carvalho-e-Silva 2004a; Hedges <i>et al.</i> , 2008
<i>Ischnocnema randorum</i>	5	Nocturnal	1.8	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Ischnocnema sambaqui</i>	34	Nocturnal	3.7	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Ischnocnema spanios</i>	190	—	2.1	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Ischnocnema venancioi</i>	76	—	2	Non-toxic	Cryptozoic	Forest areas	Direct	Lutz 1958; Carvalho-e-Silva 2004 <sup>a</sup>
<i>Ischnocnema verrucosa</i>	855	Nocturnal	2.4	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Ischnocnema vizottoi</i>	40	Nocturnal	2	Non-toxic	Fossorial	Forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Itapotihyla langsdorffii</i>	4454	Nocturnal	10.3	Unpalatable or bad odour	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Leptodactylus cupreus</i>	12	Nocturnal	5.7	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Leptodactylus flavopictus</i>	1014	Nocturnal	13.2	Toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Leptodactylus furnarius</i>	3093	Nocturnal	4.2	Non-toxic	Aquatic	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Leptodactylus fuscus</i>	9759	Nocturnal	4.4	Non-toxic	Terrestrial	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Leptodactylus gracilis</i>	3623	Nocturnal	4.4	Non-toxic	Cryptozoic	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Leptodactylus hylodes</i>	2	—	2.5	—	—	—	—	Heyer 2000

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<i>Leptodactylus jolyi</i>	7	Nocturnal	5.4	Non-toxic	Terrestrial	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Leptodactylus labyrinthicus</i>	4539	Nocturnal	15.5	Toxic	Terrestrial	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Leptodactylus latrans</i>	9670	Nocturnal	9.6	Toxic	Terrestrial	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Leptodactylus macrosternum</i>	8302	Nocturnal	9	Toxic	Terrestrial	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Leptodactylus marambaiae</i>	4	—	4.1	—	—	—	Indirect	Izecksohn 1976; Carvalho-e-Silva and Cruz 2004b
<i>Leptodactylus mystaceus</i>	830	Nocturnal	5.2	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Leptodactylus mystacinus</i>	9605	Nocturnal	5.5	Toxic	Terrestrial	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Leptodactylus natalensis</i>	2132	Nocturnal	5.2	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Leptodactylus notoaktites</i>	2253	Nocturnal	5	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Leptodactylus plaumanni</i>	2141	Nocturnal	3.8	Non-toxic	Cryptozoic	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Leptodactylus podicipinus</i>	2529	Nocturnal	4.2	Non-toxic	Cryptozoic	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Leptodactylus spixii</i>	3808	Nocturnal	5.2	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Leptodactylus troglodytes</i>	1011	Nocturnal	5.1	Non-toxic	Terrestrial	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Leptodactylus vastus</i>	886	Nocturnal	15	Toxic	Terrestrial	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Leptodactylus viridis</i>	234	Nocturnal	7.2	Toxic	Terrestrial	Open areas	Indirect	Haddad <i>et al.</i> , 2013

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<i>Limnomedusa macroglossa</i>	1590	Nocturnal	5.4	—	Terrestrial	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Lithobates catesbeianus</i>	1198	Nocturnal	8.9	Non-toxic	Semi-aquatic	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Lithobates palmipes</i>	346	Nocturnal	11.3	Non-toxic	Semi-aquatic	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Luetkenotyphlus brasiliensis</i>	3313	—	27	—	Fossorial	Open and forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Macrogenioglottus alipioi</i>	2512	Nocturnal	9.1	Toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Megaelosia apuana</i>	2	Diurnal	8.1	Non-toxic	Rheophilic	Forest areas	Indirect	Pombal <i>et al.</i> , 2003; Pombal 2004
<i>Megaelosia bocainensis</i>	5	Diurnal	10.1	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Megaelosia boticariana</i>	3	—	7.8	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Megaelosia goeldii</i>	451	Diurnal	10	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Megaelosia lutzae</i>	7	Diurnal	9.6	Non-toxic	Rheophilic	Forest areas	Indirect	Izecksohn and Gouvêa 1987; Carvalho-e-Silva and Telles 2004b
<i>Megaelosia massarti</i>	38	Diurnal	11.5	Non-toxic	Rheophilic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Melanophryniscus admirabilis</i>	7	Diurnal	3.8	Toxic	Terrestrial	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Melanophryniscus alipioi</i>	5	Diurnal	2.5	Toxic	Phytotelmata	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Melanophryniscus cambaraensis</i>	22	Diurnal/nocturnal	3.5	Toxic	Terrestrial	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Melanophryniscus dorsalis</i>	110	Diurnal	2.5	Toxic	Terrestrial	Open areas	Indirect	Haddad <i>et al.</i> , 2013

<i>Melanophryniscus macrogranulosus</i>	2	—	3.5	Toxic	Terrestrial	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Melanophryniscus moreirae</i>	17	Diurnal	2.9	Toxic	Terrestrial	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Melanophryniscus peritus</i>	2	Diurnal	4	Toxic	—	—	Indirect	Caramaschi and Cruz 2011;
<i>Melanophryniscus setiba</i>	67	Diurnal	1.5	Toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Melanophryniscus simplex</i>	9	Nocturnal	2.8	Toxic	Terrestrial	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Melanophryniscus spectabilis</i>	1	Diurnal	3.1	Toxic	Terrestrial	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Melanophryniscus tumifrons</i>	1480	—	3.1	Toxic	Terrestrial	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Melanophryniscus vilavelhensis</i>	13	Nocturnal	1.7	Toxic	Phytotelmate	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Mimosiphonops vermiculatus</i>	4	—	18.7	—	Cryptozoic	Forest areas	Direct	Taylor 1968; Silvano <i>et al.</i> , 2004 <sup>a</sup>
<i>Myersiella microps</i>	1236	Nocturnal	3.1	Non-toxic	Fossorial	Forest areas	Direct	Haddad <i>et al.</i> , 2013; present study
<i>Odontophrynus americanus</i>	8662	Nocturnal	4.9	Toxic	Terrestrial	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Odontophrynus carvalhoi</i>	657	Nocturnal	6.7	Toxic	Terrestrial	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Odontophrynus maisuma</i>	8390	Nocturnal	4.1	Toxic	Terrestrial	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Ololygon agilis</i>	825	Diurnal/nocturnal	1.8	Non-toxic	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Ololygon albicans</i>	464	Nocturnal	3	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study



<i>Ololygon angrensis</i>	102	Diurnal/nocturnal	3.4	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Ololygon arduous</i>	2	Diurnal/nocturnal	2.1	Non-toxic	Phytotelmate	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Ololygon argyromatus</i>	2218	Nocturnal	2.1	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Ololygon ariadne</i>	60	Nocturnal	4.1	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Ololygon aromothyella</i>	10	Nocturnal	2.6	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Ololygon atrata</i>	31	—	2	Non-toxic	Arboreal	—	Indirect	Peixoto 1989
<i>Ololygon belloni</i>	5	Nocturnal	3.9	Non-toxic	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Ololygon brieni</i>	805	Nocturnal	3.5	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Ololygon carnevallii</i>	960	Nocturnal	3.2	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Ololygon catharinae</i>	716	Nocturnal	4.5	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Ololygon cosenzai</i>	10	Nocturnal	2.3	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Ololygon flavoguttata</i>	1911	Nocturnal	4.5	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Ololygon heyeri</i>	30	Nocturnal	3.9	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Ololygon hiemalis</i>	964	Nocturnal	3.5	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Ololygon humilis</i>	851	Nocturnal	2.6	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Ololygon insperata</i>	5	Nocturnal	2.3	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Ololygon jureia</i>	5	Nocturnal	3	Non-toxic	Arboreal	Forest areas	Indirect	Pombal and Gordo 1991; Rodrigues and Cruz 2004a
<i>Ololygon kautskyi</i>	1	—	2.8	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Ololygon littoralis</i>	300	Nocturnal	3.8	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study

<i>Ololygon littoreus</i>	113	Nocturnal	2.2	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Ololygon longilinea</i>	861	Nocturnal	4.2	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Ololygon luizotavioi</i>	1178	Nocturnal	2.5	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Ololygon machadoi</i>	908	Nocturnal	2.4	Non-toxic	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Ololygon melloi</i>	19	—	1.9	Non-toxic	Arboreal	—	Indirect	Peixoto 1989; Cruz and Telles 2004
<i>Ololygon obtriangulata</i>	1574	Nocturnal	3.5	Non-toxic	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Ololygon peixotoi</i>	34	Nocturnal	2.3	Non-toxic	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Ololygon perpusilla</i>	861	Nocturnal	2	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Ololygon ranki</i>	76	Nocturnal	2.3	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Ololygon rizibilis</i>	1817	Diurnal/nocturnal	3	Non-toxic	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Ololygon strigilata</i>	310	Nocturnal	3.8	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Ololygon signata</i>	x- 7889	Nocturnal	4.1	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Oscaecilia hypereumeces</i>	1	—	64	—	Cryptozoic	Forest areas	Direct	Taylor 1968; Silvano <i>et al.</i> , 2004b
<i>Paratelmatoobius cardosoi</i>	28	Nocturnal	1.7	—	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Paratelmatoobius gaigeae</i>	5	Nocturnal	1.8	—	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Paratelmatoobius lutzii</i>	2	—	2.4	—	Cryptozoic	Open areas	Indirect	Haddad <i>et al.</i> , 2013

<i>Paratelmatoebius mantiquera</i>	4	—	1.6	—	Terrestrial	Forest areas	Indirect	Pombal and Haddad 1999
<i>Paratelmatoebius poecilogaster</i>	34	Nocturnal	2.7	—	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Paratelmatoebius yepiranga</i>	16	Nocturnal	2.1	—	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Phasmahyla cochranæ</i>	1126	Nocturnal	3.2	Toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Phasmahyla exilis</i>	438	Nocturnal	2.9	Toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Phasmahyla guttata</i>	917	Nocturnal	3.8	Toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Phasmahyla jandaia</i>	230	Nocturnal	3.6	Toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Phasmahyla spectabilis</i>	127	Nocturnal	4.6	Toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Phasmahyla timbo</i>	5	Nocturnal	3.5	Toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Phrynomedusa appendiculata</i>	20	—	3.5	Toxic	Arboreal	Forest areas	Indirect	Cruz 1991; Garcia <i>et al.</i> , 2004
<i>Phrynomedusa bokermanni</i>	4	—	4.6	Toxic	Arboreal	Forest areas	Indirect	Cruz ruz and Caramaschi 2004
<i>Phrynomedusa fimbriata</i>	4	—	4.5	Toxic	Arboreal	Forest areas	Indirect	Cruz 1991; Cruz and Pimenta 2004
<i>Phrynomedusa marginata</i>	747	Nocturnal	3.1	Toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Phrynomedusa vanzolinii</i>	229	—	3.6	Toxic	Arboreal	Forest areas	Indirect	Cruz 1991; Cruz and Carvalho-e-Silva 2004b
<i>Phyllodytes acuminatus</i>	996	—	2.4	Non-toxic	Phytotelmate	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Phyllodytes</i>	4	—	2.4	Non-toxic	Phytotelmate	Forest areas	Indirect	Peixoto and Cruz 1998; Skuk <i>et al.</i> , 2004

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<i>brevirostris</i>									
<i>Phyllodytes edelmoi</i>	196	—	2.6	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013	
<i>Phyllodytes gyrinaethes</i>	105	—	2.6	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013	
<i>Phyllodytes kautskyi</i>	657	Diurnal/nocturnal	4	Non-toxic	Phytotelmate	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study	
<i>Phyllodytes luteolus</i>	1671	Nocturnal	2.5	Non-toxic	Phytotelmate	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study	
<i>Phyllodytes maculosus</i>	435	Nocturnal	4.9	Non-toxic	Phytotelmate	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; Ferreira <i>et al.</i> , 2016; present study	
<i>Phyllodytes melanomystax</i>	547	Nocturnal	2.3	Non-toxic	Phytotelmate	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study	
<i>Phyllodytes punctatus</i>	1	—	2.1	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013	
<i>Phyllodytes tuberculosis</i>	42	Nocturnal	2.4	Non-toxic	Phytotelmate	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study	
<i>Phyllodytes wuchereri</i>	615	Nocturnal	2.7	Non-toxic	Phytotelmate	Forest areas	Indirect	Haddad <i>et al.</i> , 2013	
<i>Phyllomedusa bahiana</i>	337	Nocturnal	7.5	Toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013	
<i>Phyllomedusa burmeisteri</i>	4892	Nocturnal	6	Toxic	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study	
<i>Phyllomedusa distincta</i>	1766	Nocturnal	6	Toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study	
<i>Phyllomedusa iheringii</i>	47	Nocturnal	6.8	Toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013	
<i>Phyllomedusa nordestina</i>	1805	Nocturnal	3.7	Toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study	

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<i>Phyllomedusa rohdei</i>	2021	Nocturnal	4.6	Toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Phyllomedusa tetraploidea</i>	3440	Nocturnal	6.6	Toxic	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Physalaemus aguirrei</i>	976	Nocturnal	2.7	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Physalaemus angrensis</i>	2	—	2	Non-toxic	Cryptozoic	Forest areas	Indirect	Weber <i>et al.</i> , 2006
<i>Physalaemus atlanticus</i>	1	Nocturnal	2.1	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Physalaemus barrioi</i>	6	Nocturnal	2.9	Non-toxic	Terrestrial	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Physalaemus bokermanni</i>	4	Nocturnal	1.7	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Physalaemus caete</i>	37	—	2.5	Non-toxic	Cryptozoic	Forest areas	Indirect	Pombal and Madureira 1997; Freire and Silvano 2004
<i>Physalaemus camacan</i>	1	Nocturnal	2.4	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Physalaemus crombiei</i>	1267	Nocturnal	2.3	Non-toxic	Terrestrial	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Physalaemus cuvieri</i>	9704	Nocturnal	3.1	Non-toxic	Terrestrial	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Physalaemus erikae</i>	222	Nocturnal	2.6	Non-toxic	Terrestrial	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Physalaemus feioi</i>	400	Nocturnal	2.8	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Physalaemus insperatus</i>	2	—	2.6	Non-toxic	Cryptozoic	Forest areas	Indirect	Cruz <i>et al.</i> , 2008; Ângulo 2009
<i>Physalaemus irroratus</i>	6	—	2.3	Non-toxic	Cryptozoic	Forest areas	Indirect	Cruz <i>et al.</i> , 2007

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<i>Physalaemus jordanensis</i>	363	Nocturnal	2.8	Non-toxic	Cryptozoic	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Physalaemus kroyeri</i>	769	Nocturnal	2.9	Non-toxic	Cryptozoic	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Physalaemus lateristriga</i>	185	Nocturnal	3.5	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Physalaemus lisei</i>	681	Nocturnal	3	Non-toxic	Cryptozoic	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Physalaemus maculiventris</i>	1481	Nocturnal	2.3	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Physalaemus maximus</i>	114	Nocturnal	4.8	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Physalaemus moreirae</i>	86	Nocturnal	2.6	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Physalaemus nanus</i>	573	Nocturnal	2.1	Non-toxic	Cryptozoic	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Physalaemus obtectus</i>	482	Nocturnal	2.8	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Physalaemus olfersii</i>	2346	Nocturnal	3.8	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Physalaemus signifer</i>	1688	Nocturnal	2.2	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Physalaemus soaresi</i>	6	Nocturnal	2.1	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Physalaemus spiniger</i>	286	Nocturnal	2.2	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Pipa carvalhoi</i>	2152	Diurnal/nocturnal	5.5	Non-toxic	Aquatic	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Pristimantis paulodutra</i>	708	Nocturnal	3.6	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013; present study

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<i>Pristimantis ramagii</i>	1272	Nocturnal	2.4	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013; present study
<i>Pristimantis vinhai</i>	763	Nocturnal	2.5	Non-toxic	Cryptozoic	Forest areas	Direct	Haddad <i>et al.</i> , 2013; present study
<i>Proceratophrys appendiculata</i>	1226	Nocturnal	6.3	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Proceratophrys avelinoi</i>	2412	Nocturnal	2.6	Non-toxic	Cryptozoic	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Proceratophrys bigibbosa</i>	913	Nocturnal	5.3	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Proceratophrys boiei</i>	5334	Nocturnal	5.7	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Proceratophrys brauni</i>	410	Diurnal/nocturnal	3.9	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Proceratophrys cristiceps</i>	722	Nocturnal	6.4	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Proceratophrys cururu</i>	1	Nocturnal	5.4	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Proceratophrys laticeps</i>	1287	Nocturnal	6.7	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Proceratophrys melanopogon</i>	532	Nocturnal	5.1	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Proceratophrys minuta</i>	11	Nocturnal	3	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Proceratophrys moehringi</i>	66	Nocturnal	6.3	Non-toxic	Cryptozoic	Forest areas	Indirect	Weygoldt and Peixoto 1985; Peixoto and Silvano 2004a
<i>Proceratophrys paviotii</i>	2	Nocturnal	4.7	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Proceratophrys phyllostoma</i>	4	Nocturnal	5.6	Non-toxic	Cryptozoic	Forest areas	Indirect	Izecksohn <i>et al.</i> , 1999; Peixoto and Silvano 2004b

<i>Proceratophrys renalis</i>	1366	Nocturnal	5.5	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Proceratophrys sanctaritae</i>	10	Nocturnal	4.1	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Proceratophrys schirchi</i>	1531	Nocturnal	5	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Proceratophrys subguttata</i>	309	Nocturnal	4.8	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Pseudis bolbodactyla</i>	1951	Nocturnal	4.5	Non-toxic	Aquatic	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Pseudis cardosoi</i>	257	Nocturnal	4.5	Non-toxic	Aquatic	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Pseudis fusca</i>	221	Nocturnal	4.1	Non-toxic	Aquatic	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Pseudis minuta</i>	795	Nocturnal	3.7	Non-toxic	Aquatic	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Pseudis paradoxa</i>	1464	Nocturnal	5.3	Non-toxic	Aquatic	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Pseudopaludicola falcipes</i>	5436	Diurnal/nocturnal	1.8	Non-toxic	Cryptozoic	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Rhinella abei</i>	531	Nocturnal	9.3	Toxic	Terrestrial	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Rhinella achavali</i>	306	Nocturnal	12	Toxic	Terrestrial	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Rhinella crucifer</i>	1943	Nocturnal	10.1	Toxic	Terrestrial	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Rhinella dorbignyi</i>	57	Nocturnal	5.6	Toxic	Terrestrial	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Rhinella fernandezae</i>	971	Nocturnal	5.7	Toxic	Terrestrial	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Rhinella granulosa</i>	8540	Nocturnal	5.3	Toxic	Terrestrial	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Rhinella henseli</i>	940	Nocturnal	7	Toxic	Terrestrial	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study



<i>Rhinella hoogmoedi</i>	2257	Nocturnal	5.6	Toxic	Terrestrial	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Rhinella icterica</i>	7322	Nocturnal	13.7	Toxic	Terrestrial	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Rhinella jimi</i>	2658	Nocturnal	14.5	Toxic	Terrestrial	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Rhinella ornata</i>	2158	Nocturnal	7.9	Toxic	Terrestrial	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Rhinella pygmaea</i>	305	Nocturnal	4.1	Toxic	Terrestrial	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Rhinella schneideri</i>	9416	Nocturnal	13	Toxic	Terrestrial	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Scinax alter</i>	3319	Nocturnal	2.8	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Scinax auratus</i>	956	—	2.6	Non-toxic	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Scinax caldarum</i>	1052	Nocturnal	2.9	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Scinax cardosoi</i>	282	—	2.6	Non-toxic	Arboreal	—	Indirect	Carvalho-e-Silva and Peixoto 1991; Carvalho-e-Silva and Caramaschi 2004
<i>Scinax cretatus</i>	66	Nocturnal	3.2	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Scinax crospeospilus</i>	777	Nocturnal	3.1	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Scinax cuspidatus</i>	2246	Nocturnal	3.1	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Scinax duartei</i>	1040	Nocturnal	3	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Scinax eurydice</i>	3145	Nocturnal	5.3	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Scinax fuscomarginatus</i>	6859	Nocturnal	2.2	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Scinax fuscovarios</i>	9363	Nocturnal	4.5	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Scinax granulatus</i>	2971	Nocturnal	4.1	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Scinax hayii</i>	1965	Nocturnal	5.1	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study

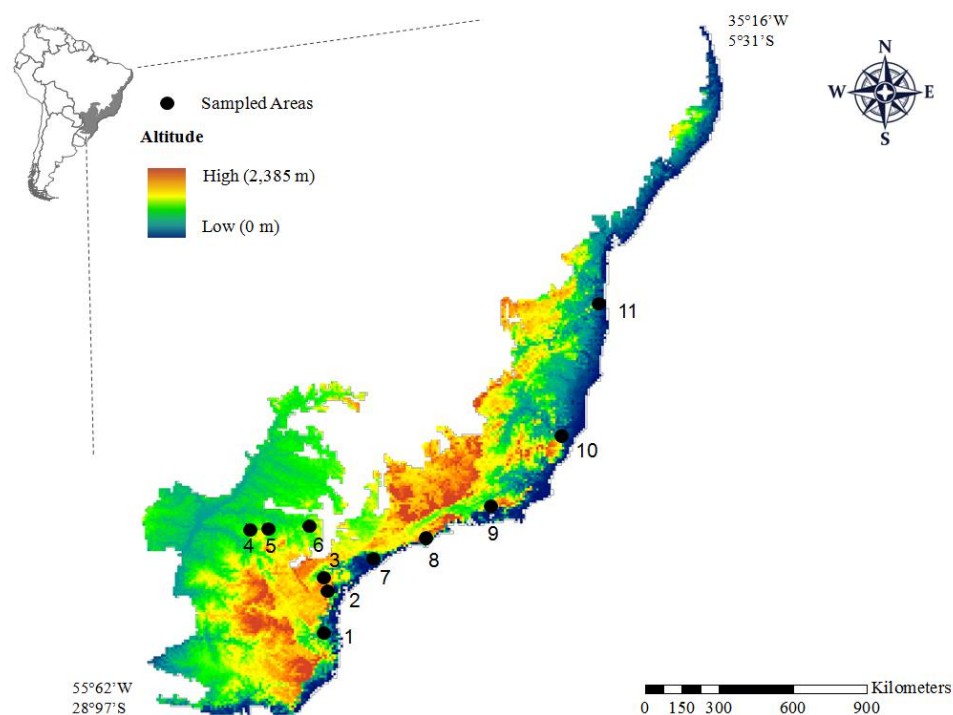
<i>Scinax imbegue</i>	16	Nocturnal	3.2	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Scinax juncae</i>	23	Nocturnal	2.4	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Scinax pachycrus</i>	820	Nocturnal	3.2	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Scinax perereca</i>	3047	Nocturnal	4.3	Non-toxic	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Scinax similis</i>	451	Nocturnal	3.8	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Scinax squalitostris</i>	8423	Nocturnal	2.7	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Scinax trapicheiroi</i>	239	Nocturnal	2.9	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Scinax tymbamirim</i>	12	Nocturnal	2.6	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Scinax uruguayus</i>	1520	Nocturnal	2.8	Non-toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Scinax v-signatus</i>	767	Nocturnal	2.7	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Scythroehrys sawayae</i>	226	Nocturnal	2.1	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Siphonops annulatus</i>	8685	—	45.4	Toxic	Fossorial	Open and forest areas	Direct	Haddad <i>et al.</i> , 2013; present study
<i>Siphonops hardyi</i>	288	—	17.8	—	Fossorial	Open and forest areas	Direct	Haddad <i>et al.</i> , 2013
<i>Siphonops insulanus</i>	19	—	19	—	Fossorial	Open and forest areas	Direct	Taylor 1968; Caramaschi <i>et al.</i> , 2004
<i>Siphonops paulensis</i>	8614	—	30	—	Fossorial	Open and forest areas	Inirect	Haddad <i>et al.</i> , 2013
<i>Sphaenorhynchus botocudo</i>	13	Nocturnal	2.7	Non-toxic	Aquatic	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Sphaenorhynchus bromelicola</i>	2	Nocturnal	2.8	Non-toxic	Phytotelmate	Forest areas	Indirect	Bokermann 1966; Silvano and Peixoto 2004c

<i>Sphaenorhynchus caramaschii</i>	846	Nocturnal	2.7	Non-toxic	Aquatic	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Sphaenorhynchus mirim</i>	9	Nocturnal	1.7	Non-toxic	Aquatic	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Sphaenorhynchus orophilus</i>	732	Nocturnal	3.2	Non-toxic	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Sphaenorhynchus palustris</i>	1398	Nocturnal	3.1	Non-toxic	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Sphaenorhynchus pauloalvini</i>	424	Nocturnal	1.9	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Sphaenorhynchus planicola</i>	1008	Nocturnal	2	Non-toxic	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Sphaenorhynchus prasinus</i>	2031	Nocturnal	2.8	Non-toxic	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Sphaenorhynchus surdus</i>	1046	Nocturnal	2.8	Non-toxic	Aquatic	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Stereocyclops incrassatus</i>	2051	Nocturnal	6.2	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Stereocyclops parkeri</i>	258	Nocturnal	6	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Thoropa lutzi</i>	703	Nocturnal	2.6	Non-toxic	Terrestrial	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Thoropa miliaris</i>	3966	Nocturnal	6.3	Non-toxic	Terrestrial	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study
<i>Thoropa petropolitana</i>	209	—	7.5	—	Terrestrial	Forest areas	Indirect	Cochrann 1955; Carvalho-e-Silva and Peixoto 2004
<i>Thoropa saxatilis</i>	145	Nocturnal	5.9	Non-toxic	Terrestrial	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Thoropa taophora</i>	499	Nocturnal	6.3	Non-toxic	Terrestrial	Forest areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Trachycephalus atlas</i>	323	Nocturnal	10.7	Toxic	Arboreal	Open areas	Indirect	Haddad <i>et al.</i> , 2013
<i>Trachycephalus</i>	809	Nocturnal	7.8	Toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013

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<i>dibernardoii</i>									
<i>Trachycephalus imitatrix</i>	2792	Nocturnal	7.1	Toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013	
<i>Trachycephalus lepidus</i>	4	Nocturnal	5.9	Toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013	
<i>Trachycephalus mesophaeus</i>	5477	Diurnal/nocturnal	6.9	Toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study	
<i>Trachycephalus nigromaculatus</i>	3490	Nocturnal	9.2	Toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013	
<i>Trachycephalus typhoni</i>	4913	Nocturnal	9.2	Toxic	Arboreal	Open and Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study	
<i>Vitreorana eurygnatha</i>	4334	Nocturnal	2.4	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study	
<i>Vitreorana uranoscopa</i>	5065	Nocturnal	2.7	Non-toxic	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study	
<i>Xenohyla eugenioi</i>	147	Nocturnal	4.5	—	Arboreal	Open and Forest areas	Indirect	Caramaschi 1998; Silvano and Pimenta 2004a	
<i>Xenohyla truncata</i>	218	Nocturnal	5	—	Arboreal	Forest areas	Indirect	Haddad <i>et al.</i> , 2013	
<i>Zachaenus carvalhoi</i>	2	Nocturnal	2.8	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013	
<i>Zachaenus parvulus</i>	268	Nocturnal	2.9	Non-toxic	Cryptozoic	Forest areas	Indirect	Haddad <i>et al.</i> , 2013; present study	

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**Figure S1.** Map showed the location of Brazilian Atlantic Forest hotspot (in gray) in South American territory. The black circle indicates the Protected areas visited as complementary field works. 1. Parque Ecológico Spitzkopf, SC; 2. Parque Estadual Pico do Marumbi, PR; 3. Parque estadual de Campinhos, PR; 4. Parque municipal das Perobas, PR; 5. Parque Estadual Mata dos Godoy, PR; 6. Refugio municipal da vida Silvestre, Horto Florestal de Jacarezinho, PR; 7. Estação Ecológica de Juréia-Itatins, SP; 8. Parque Estadual da Serra do Mar Núcleo Caraguatatuba, SP; 9. Parque Nacional da Serra dos Órgãos, RJ; 10. Reserva Biológica Augusto Ruschi, ES; 11. Reserva Biológica de Una, BA. From January of 2014 (PA 8 five days), July 2014 (PA 6 three days), November 2014 (PAs 4, 6 three days in each), December 2014 (PA 4 two days), January to March 2015 (PAs 1, 2, 7-11 seven days in each), May 2015 (PA 6 three days), November 2015 (PA 4 two days), December 2015 (PA 3 five days), February 2016 (PAs 5 two days, 6 three days), March 2016 (PA 4 one day, 5 two days). We sampled amphibians using acoustic and visual nocturnal and diurnal surveys (Crump & Scott Jr, 1994; Zimmerman, 1994). We searched at the margins of water bodies, in the streams, and along 2000 m forest transects. The species collected were euthanized with hydrochloride benzocaine  $\geq 250$  mg / L (ASH, 2004). Number of authorization / license SISBIO - 30344.

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APÊNDICE 2 - **Table S1.** List of studied amphibians species of Atlantic Forest with the summarization of the climate change effects on species geographical range.

Order/Family/Species	Brazilian Red List	Projections Future	Range Present	Range Future	% of lost (-) and gained (+) of cells
<b>Gymnophiona</b>					
<b>Typlonectidae</b>					
<i>Chthonerpeton indistinctum</i>	Not Evaluated	Critically Endangered	551,25	52	-90,57
<b>Siphonopidae</b>					
<i>Luetkenotyphlus brasiliensis</i>	Not Evaluated	Vulnerable	3337,25	1718	-48,52
<i>Siphonops annulatus</i>	Not Evaluated	Least Concern	8681,75	7889,25	-9,13
<i>Siphonops hardyi</i>	Not Evaluated	Endangered	562,5	124	-77,96
<i>Siphonops paulensis</i>	Not Evaluated	Least Concern	8264,75	6087,75	-26,34
<b>Anura</b>					
<b>Alsodidae</b>					
<i>Limnomedusa macroglossa</i>	Not Evaluated	Endangered	1553,25	570,5	-63,27
<b>Aromobatidae</b>					

<i>Allobates alagoanus</i>	Not Evaluated	Extinct	5,5	0	-100,00
<i>Allobates capixaba</i>	Not Evaluated	Critically Endangered	449,25	22,25	-95,05
<i>Allobates olfersioides</i>	Vulnerable	Vulnerable	2704,75	1816,75	-32,83
<b>Brachycephalidae</b>					
<i>Brachycephalus alipioi</i>	Not Evaluated	Critically Endangered	13,25	0,75	-94,34
<i>Brachycephalus brunneus</i>	Not Evaluated	Least Concern	3,75	25,25	573,33
<i>Brachycephalus didactylus</i>	Not Evaluated	Endangered	623,5	141	-77,39
<i>Brachycephalus ephippium</i>	Not Evaluated	Endangered	2200,25	830	-62,28
<i>Brachycephalus ferruginus</i>	Not Evaluated	Extinct	15,5	0	-100,00
<i>Brachycephalus garbeanus</i>	Not Evaluated	Critically Endangered	9,75	0,5	-94,87
<i>Brachycephalus guarani</i>	Not Evaluated	Critically Endangered	65,25	2	-96,93
<i>Brachycephalus hermogenesi</i>	Not Evaluated	Endangered	505	141	-72,08
<i>Brachycephalus izecksohni</i>	Not Evaluated	Extinct	4,25	0	-100,00
<i>Brachycephalus margaritatus</i>	Not Evaluated	Critically Endangered	11,75	0,5	-95,74
<i>Brachycephalus nodoterga</i>	Not Evaluated	Critically Endangered	22,75	0,25	-98,90
<i>Brachycephalus pernix</i>	Critically Endangered	Extinct	2	0	-100,00
<i>Brachycephalus pitanga</i>	Not Evaluated	Endangered	12,25	4,5	-63,27



<i>Brachycephalus pombali</i>	Not Evaluated	Extinct	12	0	-100,00
<i>Brachycephalus pulex</i>	Not Evaluated	Extinct	12,25	0	-100,00
<i>Brachycephalus toby</i>	Not Evaluated	Critically Endangered	8,5	0,25	-97,06
<i>Brachycephalus tridactylus</i>	Not Evaluated	Extinct	11,25	0	-100,00
<i>Brachycephalus vertebralis</i>	Not Evaluated	Critically Endangered	147	12,25	-91,67
<i>Ischnocnema abdita</i>	Not Evaluated	Extinct	14,25	0	-100,00
<i>Ischnocnema bolbodactyla</i>	Not Evaluated	Endangered	409,5	135,75	-66,85
<i>Ischnocnema concolor</i>	Not Evaluated	Extinct	5,25	0	-100,00
<i>Ischnocnema erythromera</i>	Not Evaluated	Critically Endangered	35,75	3,5	-90,21
<i>Ischnocnema guenteri</i>	Not Evaluated	Endangered	3776	1237,75	-67,22
<i>Ischnocnema henselli</i>	Not Evaluated	Endangered	1305,75	587	-55,04
<i>Ischnocnema hoehnei</i>	Not Evaluated	Critically Endangered	469,25	87,75	-81,30
<i>Ischnocnema holti</i>	Not Evaluated	Extinct	9,25	0	-100,00
<i>Ischnocnema izecksoni</i>	Not Evaluated	Extinct	55,5	0	-100,00
<i>Ischnocnema juipoca</i>	Not Evaluated	Endangered	1924,25	947,25	-50,77
<i>Ischnocnema karst</i>	Not Evaluated	Extinct	6,5	0	-100,00
<i>Ischnocnema manezinho</i>	Vulnerable	Endangered	217,25	99,75	-54,09

<i>Ischnocnema melonopygia</i>	Not Evaluated	Extinct	1	0	-100,00
<i>Ischnocnema nasuta</i>	Not Evaluated	Vulnerable	3837,25	1990	-48,14
<i>Ischnocnema nigriventris</i>	Not Evaluated	Extinct	5,25	0	-100,00
<i>Ischnocnema octavioi</i>	Not Evaluated	Critically Endangered	445,5	71,5	-83,95
<i>Ischnocnema oea</i>	Not Evaluated	Least Concern	4	81,5	1937,50
<i>Ischnocnema parva</i>	Not Evaluated	Endangered	1828,75	688,25	-62,37
<i>Ischnocnema randorum</i>	Not Evaluated	Extinct	4,75	0	-100,00
<i>Ischnocnema sambaqui</i>	Not Evaluated	Least Concern	29,25	22	-24,79
<i>Ischnocnema spanios</i>	Not Evaluated	Critically Endangered	178,75	14,5	-91,89
<i>Ischnocnema verrucosa</i>	Not Evaluated	Endangered	998,25	307,25	-69,22
<i>Ischnocnema vizottoi</i>	Not Evaluated	Extinct	62,75	0	-100,00

#### **Bufonidae**

<i>Dendrophryniscus berthalutzae</i>	Not Evaluated	Endangered	341,25	130,5	-61,76
<i>Dendrophryniscus brevipollicatus</i>	Not Evaluated	Endangered	939,75	426,75	-54,59
<i>Dendrophryniscus carvalhoi</i>	Not Evaluated	Extinct	40,75	0	-100,00
<i>Dendrophryniscus krause</i>	Not Evaluated	Extinct	8,5	0	-100,00
<i>Dendrophryniscus leucomystax</i>	Not Evaluated	Endangered	970,5	332,75	-65,71

<i>Dendrophryniscus oreites</i>	Not Evaluated	Extinct	24,75	0	-100,00
<i>Dendrophryniscus proboscideus</i>	Not Evaluated	Endangered	786,25	282	-64,13
<i>Frostius erythrophthalmus</i>	Not Evaluated	Critically Endangered	290,5	3	-98,97
<i>Frostius pernambucensis</i>	Not Evaluated	Endangered	911,5	398,5	-56,28
<i>Melanophryniscus admirabilis</i>	Critically Endangered	Extinct	7	0	-100,00
<i>Melanophryniscus alipioi</i>	Not Evaluated	Extinct	5,5	0	-100,00
<i>Melanophryniscus cambaraensis</i>	Vulnerable	Extinct	26,75	0	-100,00
<i>Melanophryniscus dorsalis</i>	Vulnerable	Endangered	97,5	23	-76,41
<i>Melanophryniscus macrogranulosus</i>	Endangered	Endangered	2	0,5	-75,00
<i>Melanophryniscus moreirae</i>	Not Evaluated	Extinct	47,25	0	-100,00
<i>Melanophryniscus setiba</i>	Critically Endangered	Endangered	66	24,75	-62,50
<i>Melanophryniscus simplex</i>	Not Evaluated	Least Concern	7,75	23,25	200,00
<i>Melanophryniscus spectabilis</i>	Not Evaluated	Extinct	1	0	-100,00
<i>Melanophryniscus tumifrons</i>	Not Evaluated	Critically Endangered	1569,75	117,25	-92,53
<i>Melanophryniscus vilavelhenis</i>	Not Evaluated	Extinct	13	0	-100,00
<i>Rhinella abei</i>	Not Evaluated	Endangered	480	190	-60,42
<i>Rhinella achavali</i>	Not Evaluated	Critically Endangered	329,5	7,5	-97,72

<i>Rhinella crucifer</i>	Not Evaluated	Least Concern	2093,75	2257	7,80
<i>Rhinella dorbignyi</i>	Not Evaluated	Extinct	72,25	0	-100,00
<i>Rhinella fernadezae</i>	Not Evaluated	Endangered	814,75	194	-76,19
<i>Rhinella granulosa</i>	Not Evaluated	Least Concern	8672,75	9151,25	5,52
<i>Rhinella henseli</i>	Not Evaluated	Critically Endangered	1001	189,5	-81,07
<i>Rhinella hoogmoedi</i>	Not Evaluated	Least Concern	2764,25	1987,75	-28,09
<i>Rhinella icterica</i>	Not Evaluated	Endangered	7554	3402,75	-54,95
<i>Rhinella jimi</i>	Not Evaluated	Least Concern	2805,25	2781,25	-0,86
<i>Rhinella ornata</i>	Not Evaluated	Endangered	2202,75	915,25	-58,45
<i>Rhinella pygmaea</i>	Not Evaluated	Endangered	392,25	166	-57,68
<i>Rhinella schneideri</i>	Not Evaluated	Least Concern	9288,75	7128,25	-23,26
<b>Ceratophryidae</b>					
<i>Ceratophrys aurita</i>	Not Evaluated	Vulnerable	4893,5	2584,25	-47,19
<b>Craugastoridae</b>					
<i>Eleutherodactylus bilineata</i>	Not Evaluated	Critically Endangered	468,75	91,5	-80,48
<i>Euparkerella brasiliensis</i>	Not Evaluated	Endangered	443,5	105,5	-76,21
<i>Euparkerella cochranae</i>	Not Evaluated	Endangered	251,25	69,5	-72,34

<i>Euparkerella robusta</i>	Not Evaluated	Extinct	2,25	0	-100,00
<i>Euparkerella tridactyla</i>	Not Evaluated	Endangered	5,75	1,25	-78,26
<i>Haddadus aramunha</i>	Not Evaluated	Least Concern	3,5	175,75	4921,43
<i>Haddadus binotatus</i>	Not Evaluated	Endangered	4876,5	2366	-51,48
<i>Holoaden bradei</i>	Critically Endangered	Critically Endangered	44,75	2,75	-93,85
<i>Holoaden luederwaldti</i>	Endangered	Endangered	490,75	105,25	-78,55
<i>Holoaden pholeter</i>	Not Evaluated	Least Concern	6	82,25	1270,83
<i>Pristimantis paulodutraii</i>	Not Evaluated	Critically Endangered	650,75	120	-81,56
<i>Pristimantis ramagii</i>	Not Evaluated	Least Concern	1295,25	1170	-9,67
<i>Pristimantis vinhai</i>	Not Evaluated	Endangered	967,25	322,75	-66,63

### **Cycloramphidae**

<i>Cycloramphus acangatan</i>	Not Evaluated	Critically Endangered	161	27,5	-82,92
<i>Cycloramphus bandeirensis</i>	Not Evaluated	Least Concern	4,5	26,5	488,89
<i>Cycloramphus bolitoglossus</i>	Not Evaluated	Endangered	289,5	98,75	-65,89
<i>Cycloramphus boraceiensis</i>	Not Evaluated	Endangered	649,25	229,75	-64,61
<i>Cycloramphus brasiliensis</i>	Not Evaluated	Critically Endangered	389	57,25	-85,28
<i>Cycloramphus carvalhoi</i>	Not Evaluated	Critically Endangered	11,25	0,5	-95,56

<i>Cycloramphus catarinensis</i>	Not Evaluated	Critically Endangered	2,5	0,5	-80,00
<i>Cycloramphus dubius</i>	Not Evaluated	Critically Endangered	67	2,5	-96,27
<i>Cycloramphus eleutherodactylus</i>	Not Evaluated	Endangered	1879,25	472,5	-74,86
<i>Cycloramphus izecksohni</i>	Not Evaluated	Endangered	240,5	53,25	-77,86
<i>Cycloramphus juimirim</i>	Not Evaluated	Extinct	5,75	0	-100,00
<i>Cycloramphus lithomimeticus</i>	Not Evaluated	Extinct	118,75	0	-100,00
<i>Cycloramphus lutzorum</i>	Not Evaluated	Critically Endangered	616,25	119,75	-80,57
<i>Cycloramphus migueli</i>	Not Evaluated	Extinct	30,25	0	-100,00
<i>Cycloramphus organensis</i>	Not Evaluated	Critically Endangered	28	3,25	-88,39
<i>Cycloramphus rhyakonastes</i>	Not Evaluated	Extinct	14	0	-100,00
<i>Cycloramphus valae</i>	Not Evaluated	Critically Endangered	97,5	1	-98,97
<i>Thoropa lutzi</i>	Not Evaluated	Endangered	695,25	278	-60,01
<i>Thoropa miliaris</i>	Not Evaluated	Vulnerable	3967,5	2586,75	-34,80
<i>Thoropa saxatilis</i>	Vulnerable	Endangered	123,75	30,75	-75,15
<i>Thoropa taophora</i>	Not Evaluated	Endangered	492,25	117	-76,23
<i>Zachaenus carvalhoi</i>	Not Evaluated	Least Concern	5,25	73,75	1304,76
<i>Zachaenus parvulus</i>	Not Evaluated	Endangered	343	103	-69,97

**Eleutherodactylidae**

<i>Adelophryne mucronatus</i>	Not Evaluated	Extinct	34,5	0	-100,00
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<i>Adelophryne pachydactyla</i>	Not Evaluated	Extinct	220,75	0	-100,00
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**Hemiphractidae**

<i>Flectonotus fissilis</i>	Not Evaluated	Endangered	1175,75	398,75	-66,09
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<i>Flectonotus goeldii</i>	Not Evaluated	Endangered	2162	787,5	-63,58
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<i>Flectonotus ohausi</i>	Not Evaluated	Endangered	1091,5	418,75	-61,64
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<i>Gastrotheca albolineata</i>	Not Evaluated	Endangered	790,25	269,25	-65,93
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<i>Gastrotheca ernestoi</i>	Not Evaluated	Critically Endangered	398	47,75	-88,00
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<i>Gastrotheca fissipes</i>	Not Evaluated	Vulnerable	1722,5	1005,5	-41,63
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<i>Gastrotheca fulvorufa</i>	Not Evaluated	Critically Endangered	475,25	38,5	-91,90
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<i>Gastrotheca megacephala</i>	Not Evaluated	Endangered	769	162,75	-78,84
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<i>Gastrotheca microdiscus</i>	Not Evaluated	Endangered	2005,75	862	-57,02
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<i>Gastrotheca prasina</i>	Not Evaluated	Extinct	9,25	0	-100,00
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<i>Gastrotheca pulchra</i>	Not Evaluated	Extinct	4,25	0	-100,00
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<i>Gastrotheca recava</i>	Not Evaluated	Extinct	4,5	0	-100,00
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**Hylidae**

<i>Aparasphenodon arapapa</i>	Not Evaluated	Endangered	1179,5	377,75	-67,97
<i>Aparasphenodon bokermani</i>	Not Evaluated	Least Concern	5,25	11	109,52
<i>Aparasphenodon brunoi</i>	Not Evaluated	Vulnerable	2804,5	1425,75	-49,16
<i>Aplastodiscus albofrenatus</i>	Not Evaluated	Endangered	518,75	136,5	-73,69
<i>Aplastodiscus albosignatus</i>	Not Evaluated	Endangered	1281	453,75	-64,58
<i>Aplastodiscus arildae</i>	Not Evaluated	Endangered	2218,5	868	-60,87
<i>Aplastodiscus callipygius</i>	Not Evaluated	Endangered	429	112	-73,89
<i>Aplastodiscus cavicola</i>	Not Evaluated	Endangered	738	169,25	-77,07
<i>Aplastodiscus cochranae</i>	Not Evaluated	Endangered	152,5	41,75	-72,62
<i>Aplastodiscus ehrhardti</i>	Not Evaluated	Endangered	357,25	142,5	-60,11
<i>Aplastodiscus eugenioi</i>	Not Evaluated	Critically Endangered	278,5	38,5	-86,18
<i>Aplastodiscus flumineus</i>	Not Evaluated	Extinct	8	0	-100,00
<i>Aplastodiscus ibirapitanga</i>	Not Evaluated	Critically Endangered	468	50,75	-89,16
<i>Aplastodiscus leucopygius</i>	Not Evaluated	Endangered	1692	539,75	-68,10
<i>Aplastodiscus perviridis</i>	Not Evaluated	Vulnerable	5880,75	3162,25	-46,23
<i>Aplastodiscus sibilatus</i>	Not Evaluated	Extinct	180,75	0	-100,00
<i>Aplastodiscus weygoldti</i>	Not Evaluated	Critically Endangered	29,5	0,75	-97,46



<i>Bokermanohyla ahenea</i>	Not Evaluated	Extinct	3,75	0	-100,00
<i>Bokermanohyla astartea</i>	Not Evaluated	Endangered	1333,75	439,75	-67,03
<i>Bokermanohyla capra</i>	Not Evaluated	Extinct	33,5	0	-100,00
<i>Bokermanohyla caramaschii</i>	Not Evaluated	Endangered	488,5	230,75	-52,76
<i>Bokermanohyla carvalhoi</i>	Not Evaluated	Endangered	295,25	85,25	-71,13
<i>Bokermanohyla circumdata</i>	Not Evaluated	Critically Endangered	1085,75	156,25	-85,61
<i>Bokermanohyla diamantina</i>	Not Evaluated	Least Concern	4,5	304,75	6672,22
<i>Bokermanohyla gouveiai</i>	Not Evaluated	Least Concern	32,75	57,5	75,57
<i>Bokermanohyla hylax</i>	Not Evaluated	Endangered	923,25	381,75	-58,65
<i>Bokermanohyla ibitipoca</i>	Not Evaluated	Extinct	5,5	0	-100,00
<i>Bokermanohyla itapoty</i>	Not Evaluated	Extinct	26,25	0	-100,00
<i>Bokermanohyla izecksohni</i>	Not Evaluated	Extinct	2	0	-100,00
<i>Bokermanohyla lucianae</i>	Not Evaluated	Extinct	1	0	-100,00
<i>Bokermanohyla luctuosa</i>	Not Evaluated	Critically Endangered	345,5	36	-89,58
<i>Bokermanohyla martinsi</i>	Not Evaluated	Critically Endangered	155,75	12,75	-91,81
<i>Bokermanohyla nanuzae</i>	Not Evaluated	Extinct	73,25	0	-100,00
<i>Bokermanohyla oxente</i>	Not Evaluated	Extinct	1,25	0	-100,00

<i>Dendropsophus anceps</i>	Not Evaluated	Endangered	2420,5	1161,5	-52,01
<i>Dendropsophus berthaltutzae</i>	Not Evaluated	Endangered	1429	541	-62,14
<i>Dendropsophus bipunctatus</i>	Not Evaluated	Least Concern	1739,75	1571,25	-9,69
<i>Dendropsophus branneri</i>	Not Evaluated	Least Concern	3339,25	3774	13,02
<i>Dendropsophus decipiens</i>	Not Evaluated	Least Concern	4244,25	3150,75	-25,76
<i>Dendropsophus elegans</i>	Not Evaluated	Least Concern	4965	4342,75	-12,53
<i>Dendropsophus giesleri</i>	Not Evaluated	Endangered	949,25	391	-58,81
<i>Dendropsophus haddadi</i>	Not Evaluated	Endangered	838,75	253,75	-69,75
<i>Dendropsophus meridianus</i>	Not Evaluated	Endangered	281,5	96,5	-65,72
<i>Dendropsophus microps</i>	Not Evaluated	Endangered	3272,75	1526,25	-53,36
<i>Dendropsophus minutus</i>	Not Evaluated	Least Concern	9520,5	7160,5	-24,79
<i>Dendropsophus nahdereri</i>	Not Evaluated	Endangered	321,5	113,25	-64,77
<i>Dendropsophus nanus</i>	Not Evaluated	Least Concern	7355,5	6269,25	-14,77
<i>Dendropsophus novaisi</i>	Not Evaluated	Least Concern	51	60,25	18,14
<i>Dendropsophus oliveirai</i>	Not Evaluated	Endangered	758	230,5	-69,59
<i>Dendropsophus pseudomeridianus</i>	Not Evaluated	Critically Endangered	329,75	47,25	-85,67
<i>Dendropsophus ruschii</i>	Not Evaluated	Vulnerable	39,25	22	-43,95

<i>Dendropsophus samborni</i>	Not Evaluated	Endangered	3494,75	1726,5	-50,60
<i>Dendropsophus seniculus</i>	Not Evaluated	Endangered	2934,75	1435	-51,10
<i>Dendropsophus soaresi</i>	Not Evaluated	Endangered	475,75	227,5	-52,18
<i>Dendropsophus studarae</i>	Not Evaluated	Extinct	2	0	-100,00
<i>Dendropsophus weneri</i>	Not Evaluated	Endangered	754,75	288,75	-61,74
<i>Hypsiboas albomarginatus</i>	Not Evaluated	Least Concern	4933	3602,75	-26,97
<i>Hypsiboas albopunctatus</i>	Not Evaluated	Vulnerable	7665,5	4659,5	-39,21
<i>Hypsiboas atlanticus</i>	Not Evaluated	Endangered	893,75	306,75	-65,68
<i>Hypsiboas bichoffi</i>	Not Evaluated	Endangered	2497,5	790	-68,37
<i>Hypsiboas caingua</i>	Not Evaluated	Endangered	1273,25	280,5	-77,97
<i>Hypsiboas caipora</i>	Not Evaluated	Extinct	22,75	0	-100,00
<i>Hypsiboas crepitans</i>	Not Evaluated	Least Concern	7944,75	7303,75	-8,07
<i>Hypsiboas curupi</i>	Vulnerable	Critically Endangered	125,5	14,5	-88,45
<i>Hypsiboas exastis</i>	Not Evaluated	Extinct	49,25	0	-100,00
<i>Hypsiboas faber</i>	Not Evaluated	Vulnerable	9042	5090,25	-43,70
<i>Hypsiboas freicanecae</i>	Not Evaluated	Extinct	1	0	-100,00
<i>Hypsiboas guenteri</i>	Not Evaluated	Vulnerable	383,75	195	-49,19

<i>Hypsiboas joaquinii</i>	Not Evaluated	Endangered	177,5	52,5	-70,42
<i>Hypsiboas latistriatus</i>	Not Evaluated	Critically Endangered	16	2,5	-84,38
<i>Hypsiboas leptolineatus</i>	Not Evaluated	Critically Endangered	1557,75	128,25	-91,77
<i>Hypsiboas lundii</i>	Not Evaluated	Vulnerable	3080,25	1593,25	-48,28
<i>Hypsiboas marginatus</i>	Not Evaluated	Endangered	187,5	39,5	-78,93
<i>Hypsiboas pardalis</i>	Not Evaluated	Endangered	2630	1298,25	-50,64
<i>Hypsiboas poajii</i>	Not Evaluated	Extinct	7,5	0	-100,00
<i>Hypsiboas polytaenius</i>	Not Evaluated	Endangered	1764,75	643,25	-63,55
<i>Hypsiboas pombali</i>	Not Evaluated	Endangered	1060,25	481,25	-54,61
<i>Hypsiboas prasinus</i>	Not Evaluated	Endangered	2845,75	825,25	-71,00
<i>Hypsiboas pulchellus</i>	Not Evaluated	Endangered	2232,5	588,75	-73,63
<i>Hypsiboas punctatus</i>	Not Evaluated	Least Concern	6546	6312,25	-3,57
<i>Hypsiboas raniceps</i>	Not Evaluated	Least Concern	3436	3689	7,36
<i>Hypsiboas secedens</i>	Not Evaluated	Critically Endangered	732,75	90,5	-87,65
<i>Hypsiboas semiguttatus</i>	Endangered	Endangered	215,5	102,25	-52,55
<i>Hypsiboas semilineatus</i>	Not Evaluated	Least Concern	4980	3730,25	-25,10
<i>Hypsiboas stellae</i>	Not Evaluated	Extinct	193,75	0	-100,00

<i>Hypsiboas stenocephalus</i>	Not Evaluated	Extinct	7	0	-100,00
<i>Itapotihyla langsdorffii</i>	Not Evaluated	Least Concern	4504,5	3469,75	-22,97
<i>Ololygon agilis</i>	Not Evaluated	Critically Endangered	631,5	50	-92,08
<i>Ololygon albicans</i>	Not Evaluated	Endangered	583,5	148,25	-74,59
<i>Ololygon angrensis</i>	Not Evaluated	Critically Endangered	100,75	20	-80,15
<i>Ololygon arduous</i>	Not Evaluated	Least Concern	6,75	8,25	22,22
<i>Ololygon argyreornata</i>	Not Evaluated	Vulnerable	2665,25	1373	-48,49
<i>Ololygon ariadne</i>	Not Evaluated	Endangered	92,5	24	-74,05
<i>Ololygon aromothyella</i>	Not Evaluated	Extinct	10,5	0	-100,00
<i>Ololygon belloni</i>	Not Evaluated	Extinct	5,5	0	-100,00
<i>Ololygon brienti</i>	Not Evaluated	Critically Endangered	831,25	161,5	-80,57
<i>Ololygon carnevalli</i>	Not Evaluated	Critically Endangered	584	81,75	-86,00
<i>Ololygon catharinae</i>	Not Evaluated	Vulnerable	815	447	-45,15
<i>Ololygon cosenzai</i>	Not Evaluated	Extinct	13	0	-100,00
<i>Ololygon flavoguttata</i>	Not Evaluated	Endangered	2204	957,75	-56,54
<i>Ololygon heyeri</i>	Not Evaluated	Vulnerable	45,25	24,5	-45,86
<i>Ololygon hiemalis</i>	Not Evaluated	Endangered	855,75	227,25	-73,44

<i>Ololygon humilis</i>	Not Evaluated	Endangered	918	347,25	-62,17
<i>Ololygon insperata</i>	Not Evaluated	Extinct	5,5	0	-100,00
<i>Ololygon kautskyi</i>	Not Evaluated	Endangered	1,25	0,5	-60,00
<i>Ololygon litoralis</i>	Not Evaluated	Critically Endangered	225,75	32	-85,83
<i>Ololygon littoreus</i>	Not Evaluated	Critically Endangered	187	23,5	-87,43
<i>Ololygon longilinea</i>	Not Evaluated	Critically Endangered	787	73,5	-90,66
<i>Ololygon luizotavioi</i>	Not Evaluated	Critically Endangered	1045	156,5	-85,02
<i>Ololygon machadoi</i>	Not Evaluated	Critically Endangered	668	38,25	-94,27
<i>Ololygon obtriangulata</i>	Not Evaluated	Critically Endangered	1589,75	300,75	-81,08
<i>Ololygon peixotoi</i>	Critically Endangered	Critically Endangered	64	0,5	-99,22
<i>Ololygon perpusilla</i>	Not Evaluated	Endangered	1109,5	459,25	-58,61
<i>Ololygon ranki</i>	Not Evaluated	Extinct	67,5	0	-100,00
<i>Ololygon rizibilis</i>	Not Evaluated	Endangered	1670,5	649,75	-61,10
<i>Ololygon strigilata</i>	Not Evaluated	Critically Endangered	234,5	0,75	-99,68
<i>Ololygon v-signata</i>	Not Evaluated	Endangered	741,5	284,75	-61,60
<i>Phasmahyla cochranæ</i>	Not Evaluated	Critically Endangered	1097	170,75	-84,43
<i>Phasmahyla exilis</i>	Not Evaluated	Critically Endangered	536	50	-90,67

<i>Phasmahyla guttata</i>	Not Evaluated	Endangered	1085	369,25	-65,97
<i>Phasmahyla jandaia</i>	Not Evaluated	Critically Endangered	211,25	9,25	-95,62
<i>Phasmahyla spectabilis</i>	Not Evaluated	Critically Endangered	191	3,5	-98,17
<i>Phasmahyla timbo</i>	Not Evaluated	Extinct	4,75	0	-100,00
<i>Phrynomedusa marginata</i>	Not Evaluated	Endangered	891	366,5	-58,87
<i>Phyllodytes acuminatus</i>	Not Evaluated	Critically Endangered	797	130	-83,69
<i>Phyllodytes edelmoi</i>	Not Evaluated	Critically Endangered	102,75	0,25	-99,76
<i>Phyllodytes gyrinaethes</i>	Critically Endangered	Extinct	61	0	-100,00
<i>Phyllodytes kautskyi</i>	Not Evaluated	Critically Endangered	616,75	110	-82,16
<i>Phyllodytes luteolus</i>	Not Evaluated	Least Concern	1780,5	1451	-18,51
<i>Phyllodytes maculosus</i>	Not Evaluated	Critically Endangered	350	7,5	-97,86
<i>Phyllodytes melanomystax</i>	Not Evaluated	Endangered	518,25	104	-79,93
<i>Phyllodytes punctatus</i>	Not Evaluated	Least Concern	2	7,75	287,50
<i>Phyllodytes tuberculosus</i>	Not Evaluated	Extinct	31,75	0	-100,00
<i>Phyllodytes wuchereri</i>	Not Evaluated	Critically Endangered	377,5	28	-92,58
<i>Pseudis bolbodactyla</i>	Not Evaluated	Least Concern	2409,5	1758	-27,04
<i>Pseudis cardosoi</i>	Not Evaluated	Critically Endangered	286	24,5	-91,43

<i>Pseudis fusca</i>	Not Evaluated	Critically Endangered	233	9,25	-96,03
<i>Pseudis minuta</i>	Not Evaluated	Endangered	735	268	-63,54
<i>Pseudis paradoxa</i>	Not Evaluated	Least Concern	1579	1423,25	-9,86
<i>Scinax alter</i>	Not Evaluated	Vulnerable	3630,5	2190	-39,68
<i>Scinax auratus</i>	Not Evaluated	Least Concern	844,75	720,25	-14,74
<i>Scinax caldarum</i>	Not Evaluated	Critically Endangered	1009,75	118,75	-88,24
<i>Scinax cretatus</i>	Not Evaluated	Extinct	46,5	0	-100,00
<i>Scinax crospedospilus</i>	Not Evaluated	Endangered	827	260,75	-68,47
<i>Scinax cuspidatus</i>	Not Evaluated	Endangered	2502,25	1152,25	-53,95
<i>Scinax duartei</i>	Vulnerable	Critically Endangered	781,75	77,5	-90,09
<i>Scinax eurydice</i>	Not Evaluated	Least Concern	3276,75	2694,75	-17,76
<i>Scinax fuscomarginatus</i>	Not Evaluated	Least Concern	6943	5660,5	-18,47
<i>Scinax fuscovarius</i>	Not Evaluated	Vulnerable	9273	6189,25	-33,26
<i>Scinax granulatus</i>	Not Evaluated	Endangered	3019,75	1048,25	-65,29
<i>Scinax hayii</i>	Not Evaluated	Endangered	2290	974,75	-57,43
<i>Scinax imbegue</i>	Not Evaluated	Extinct	15	0	-100,00
<i>Scinax juncae</i>	Not Evaluated	Extinct	18,75	0	-100,00



<i>Scinax pachychrus</i>	Not Evaluated	Least Concern	845,75	1225,25	44,87
<i>Scinax perereca</i>	Not Evaluated	Endangered	3227,75	1364	-57,74
<i>Scinax similis</i>	Not Evaluated	Endangered	541,25	175,75	-67,53
<i>Scinax squalirostris</i>	Not Evaluated	Endangered	8474,25	4132,75	-51,23
<i>Scinax trapicheiroi</i>	Not Evaluated	Endangered	269,5	58,5	-78,29
<i>Scinax tymbamirim</i>	Not Evaluated	Extinct	12	0	-100,00
<i>Scinax uruguayus</i>	Not Evaluated	Critically Endangered	1468,25	71,75	-95,11
<i>Scinax x-signatus</i>	Not Evaluated	Least Concern	7916,25	6995,25	-11,63
<i>Sphaenorhynchus botucudo</i>	Not Evaluated	Extinct	18	0	-100,00
<i>Sphaenorhynchus caramaschii</i>	Not Evaluated	Endangered	793,5	338,5	-57,34
<i>Sphaenorhynchus mirim</i>	Not Evaluated	Endangered	9	2,25	-75,00
<i>Sphaenorhynchus orophilus</i>	Not Evaluated	Endangered	943,25	284,25	-69,86
<i>Sphaenorhynchus palustris</i>	Not Evaluated	Vulnerable	1311,25	692	-47,23
<i>Sphaenorhynchus pauloalvini</i>	Not Evaluated	Critically Endangered	420,75	3,5	-99,17
<i>Sphaenorhynchus planicola</i>	Not Evaluated	Endangered	879,75	282	-67,95
<i>Sphaenorhynchus prasinus</i>	Not Evaluated	Vulnerable	1982,25	1300,75	-34,38
<i>Sphaenorhynchus surdus</i>	Not Evaluated	Endangered	983,25	317,25	-67,73

<i>Trachycephalus atlas</i>	Not Evaluated	Least Concern	225,75	251,5	11,41
<i>Trachycephalus dibernadoi</i>	Not Evaluated	Endangered	795,5	248	-68,82
<i>Trachycephalus imitatrix</i>	Not Evaluated	Endangered	2866,75	922,25	-67,83
<i>Trachycephalus lepidus</i>	Not Evaluated	Extinct	4	0	-100,00
<i>Trachycephalus mesophaeus</i>	Not Evaluated	Least Concern	5320,25	4459	-16,19
<i>Trachycephalus nigromaculatus</i>	Not Evaluated	Vulnerable	3510,75	1916	-45,42
<i>Trachycephalus typhoni</i>	Not Evaluated	Least Concern	4572,75	4346,5	-4,95
<i>Vitreorana eurygnathum</i>	Not Evaluated	Endangered	4170	1776	-57,41
<i>Vitreorana uranoscopa</i>	Not Evaluated	Endangered	4987,25	2008,75	-59,72
<i>Xenohyla truncata</i>	Endangered	Endangered	341,25	84,25	-75,31
<b>Hylodidae</b>					
<i>Crossodactylus aeneus</i>	Not Evaluated	Endangered	1346,75	559,75	-58,44
<i>Crossodactylus bokermani</i>	Not Evaluated	Least Concern	3	6,5	116,67
<i>Crossodactylus caramaschii</i>	Not Evaluated	Endangered	600	207	-65,50
<i>Crossodactylus dantei</i>	Endangered	Extinct	2	0	-100,00
<i>Crossodactylus dispar</i>	Not Evaluated	Endangered	1074	224,5	-79,10
<i>Crossodactylus gaudichaudii</i>	Not Evaluated	Endangered	1154,75	458,75	-60,27

<i>Crossodactylus grandis</i>	Not Evaluated	Critically Endangered	188,75	19,75	-89,54
<i>Crossodactylus schmidti</i>	Not Evaluated	Least Concern	235	297,75	26,70
<i>Hylodes amnicola</i>	Not Evaluated	Extinct	6,5	0	-100,00
<i>Hylodes asper</i>	Not Evaluated	Endangered	1263	424,75	-66,37
<i>Hylodes babax</i>	Not Evaluated	Least Concern	31,5	42,25	34,13
<i>Hylodes cardosoi</i>	Not Evaluated	Critically Endangered	80,5	3,5	-95,65
<i>Hylodes charadranaetes</i>	Not Evaluated	Critically Endangered	133	19	-85,71
<i>Hylodes dactylocinus</i>	Not Evaluated	Critically Endangered	28	1,5	-94,64
<i>Hylodes heyeri</i>	Not Evaluated	Endangered	287,25	94,5	-67,10
<i>Hylodes lateristrigatus</i>	Not Evaluated	Endangered	1229,75	462,5	-62,39
<i>Hylodes magalhaesi</i>	Not Evaluated	Critically Endangered	66,5	0,5	-99,25
<i>Hylodes meridionalis</i>	Not Evaluated	Critically Endangered	99	10,75	-89,14
<i>Hylodes nasus</i>	Not Evaluated	Endangered	1664,25	754,25	-54,68
<i>Hylodes ornatus</i>	Not Evaluated	Critically Endangered	255,5	13,75	-94,62
<i>Hylodes otavioi</i>	Not Evaluated	Extinct	3,5	0	-100,00
<i>Hylodes perere</i>	Not Evaluated	Extinct	20,75	0	-100,00
<i>Hylodes perplicatus</i>	Not Evaluated	Endangered	97	41,75	-56,96

<i>Hylodes phyllodes</i>	Not Evaluated	Critically Endangered	493,75	84	-82,99
<i>Hylodes pipilans</i>	Not Evaluated	Extinct	4,75	0	-100,00
<i>Hylodes regius</i>	Not Evaluated	Critically Endangered	53	0,5	-99,06
<i>Hylodes sazimai</i>	Not Evaluated	Critically Endangered	169,5	2,25	-98,67
<i>Megaelosia apuana</i>	Not Evaluated	Least Concern	4	26,75	568,75
<i>Megaelosia bocainensis</i>	Not Evaluated	Extinct	5	0	-100,00
<i>Megaelosia goeldii</i>	Not Evaluated	Endangered	520,75	148,25	-71,53
<i>Megaelosia massarti</i>	Not Evaluated	Extinct	42,5	0	-100,00
<b>Leptodactylidae</b>					
<i>Adenomera ajurauna</i>	Not Evaluated	Extinct	42	0	-100,00
<i>Adenomera araucaria</i>	Not Evaluated	Critically Endangered	72,25	0,5	-99,31
<i>Adenomera bokermanni</i>	Not Evaluated	Endangered	2855,75	1013	-64,53
<i>Adenomera engelsi</i>	Not Evaluated	Extinct	46	0	-100,00
<i>Adenomera marmorata</i>	Not Evaluated	Endangered	2715,5	856	-68,48
<i>Adenomera nana</i>	Not Evaluated	Endangered	234,5	70,5	-69,94
<i>Adenomera thomei</i>	Not Evaluated	Endangered	309,25	78,25	-74,70
<i>Crossodactylodes bokermanni</i>	Not Evaluated	Endangered	39,25	14,75	-62,42

<i>Crossodactylodes izecksohni</i>	Not Evaluated	Extinct	4,25	0	-100,00
<i>Leptodactylus cupreus</i>	Not Evaluated	Extinct	8,5	0	-100,00
<i>Leptodactylus flavopictus</i>	Not Evaluated	Endangered	1280,75	586	-54,25
<i>Leptodactylus furnarius</i>	Not Evaluated	Vulnerable	3487,25	2277,25	-34,70
<i>Leptodactylus fuscus</i>	Not Evaluated	Least Concern	9581	7854,5	-18,02
<i>Leptodactylus gracilis</i>	Not Evaluated	Endangered	3689	1525,5	-58,65
<i>Leptodactylus jolyi</i>	Not Evaluated	Extinct	6,75	0	-100,00
<i>Leptodactylus labyrinthicus</i>	Not Evaluated	Least Concern	4822,5	3809,75	-21,00
<i>Leptodactylus latrans</i>	Not Evaluated	Least Concern	9425,25	7312,25	-22,42
<i>Leptodactylus macrosternum</i>	Not Evaluated	Vulnerable	8249,75	5416,25	-34,35
<i>Leptodactylus mystaceus</i>	Not Evaluated	Endangered	556	253	-54,50
<i>Leptodactylus mystacinus</i>	Not Evaluated	Least Concern	9607	7031,75	-26,81
<i>Leptodactylus natalensis</i>	Not Evaluated	Least Concern	2429,5	2202	-9,36
<i>Leptodactylus noaktites</i>	Not Evaluated	Endangered	2266	552,5	-75,62
<i>Leptodactylus plaumanni</i>	Not Evaluated	Endangered	2050,5	946,25	-53,85
<i>Leptodactylus podicipinus</i>	Not Evaluated	Least Concern	2605,75	2322	-10,89
<i>Leptodactylus spixi</i>	Not Evaluated	Least Concern	3893,5	3768,5	-3,21

<i>Leptodactylus troglodytes</i>	Not Evaluated	Least Concern	1142,75	1274,75	11,55
<i>Leptodactylus vastus</i>	Not Evaluated	Vulnerable	585	378,75	-35,26
<i>Leptodactylus viridis</i>	Not Evaluated	Critically Endangered	242,5	13	-94,64
<i>Paratelmatoobius cardosoi</i>	Not Evaluated	Extinct	31,5	0	-100,00
<i>Paratelmatoobius gaigeae</i>	Not Evaluated	Extinct	6,25	0	-100,00
<i>Paratelmatoobius lutzii</i>	Critically Endangered	Critically Endangered	4,75	0,25	-94,74
<i>Paratelmatoobius poecilogaster</i>	Not Evaluated	Extinct	47	0	-100,00
<i>Paratelmatoobius yepiranga</i>	Not Evaluated	Extinct	22	0	-100,00
<i>Physalaemus aguirrei</i>	Not Evaluated	Endangered	997,5	246,5	-75,29
<i>Physalaemus atlanticus</i>	Not Evaluated	Extinct	1	0	-100,00
<i>Physalaemus barrioi</i>	Not Evaluated	Extinct	10	0	-100,00
<i>Physalaemus bokermanni</i>	Not Evaluated	Extinct	4,75	0	-100,00
<i>Physalaemus camacan</i>	Not Evaluated	Extinct	1	0	-100,00
<i>Physalaemus crombiei</i>	Not Evaluated	Endangered	1214,25	339	-72,08
<i>Physalaemus cuvieri</i>	Not Evaluated	Least Concern	9574	7430,75	-22,39
<i>Physalaemus erikae</i>	Not Evaluated	Critically Endangered	278,5	9	-96,77
<i>Physalaemus feioi</i>	Not Evaluated	Critically Endangered	336,75	3,5	-98,96

<i>Physalaemus jordanensis</i>	Not Evaluated	Critically Endangered	382	14,5	-96,20
<i>Physalaemus kroyeri</i>	Not Evaluated	Least Concern	944	1291	36,76
<i>Physalaemus lateristriga</i>	Not Evaluated	Critically Endangered	176,75	28,75	-83,73
<i>Physalaemus lisei</i>	Not Evaluated	Critically Endangered	690,5	119,75	-82,66
<i>Physalaemus maculiventris</i>	Not Evaluated	Endangered	1716,25	560,5	-67,34
<i>Physalaemus maximus</i>	Vulnerable	Extinct	118,25	0	-100,00
<i>Physalaemus moreirae</i>	Not Evaluated	Critically Endangered	99,25	7,75	-92,19
<i>Physalaemus nanus</i>	Not Evaluated	Endangered	521,25	126	-75,83
<i>Physalaemus obtectrus</i>	Not Evaluated	Endangered	753,75	225,25	-70,12
<i>Physalaemus olfersii</i>	Not Evaluated	Endangered	2669	1184,25	-55,63
<i>Physalaemus signifer</i>	Not Evaluated	Endangered	1911	747,75	-60,87
<i>Physalaemus soaresi</i>	Critically Endangered	Extinct	4,75	0	-100,00
<i>Physalaemus spiniger</i>	Not Evaluated	Endangered	250,5	59	-76,45
<i>Pseudopaludicola falcipes</i>	Not Evaluated	Endangered	5582	2564,5	-54,06
<i>Scythrophrys sawayae</i>	Not Evaluated	Endangered	226,25	96,75	-57,24
<b>Microhylidae</b>					
<i>Arcomover passareli</i>	Not Evaluated	Endangered	951,75	364,5	-61,70

<i>Chiasmocleis alagoanus</i>	Endangered	Extinct	5,25	0	-100,00
<i>Chiasmocleis atlantica</i>	Not Evaluated	Endangered	674,25	175,25	-74,01
<i>Chiasmocleis capixaba</i>	Not Evaluated	Critically Endangered	444,75	12	-97,30
<i>Chiasmocleis carvalhoi</i>	Not Evaluated	Critically Endangered	883,75	154	-82,57
<i>Chiasmocleis crucis</i>	Not Evaluated	Extinct	1	0	-100,00
<i>Chiasmocleis gnoma</i>	Not Evaluated	Extinct	1	0	-100,00
<i>Chiasmocleis leucosticta</i>	Not Evaluated	Endangered	1414,5	480	-66,07
<i>Chiasmocleis mantiqueira</i>	Not Evaluated	Extinct	6,5	0	-100,00
<i>Chiasmocleis schubarti</i>	Not Evaluated	Endangered	1390,75	677	-51,32
<i>Dasypops schirchi</i>	Not Evaluated	Critically Endangered	187,75	0,25	-99,87
<i>Elachistocleis bicolor</i>	Not Evaluated	Endangered	3509,25	1374,5	-60,83
<i>Elachistocleis cesarii</i>	Not Evaluated	Least Concern	6902	5254,25	-23,87
<i>Elachistocleis erythrogaster</i>	Not Evaluated	Extinct	49,5	0	-100,00
<i>Myersiella microps</i>	Not Evaluated	Endangered	1479,75	563	-61,95
<i>Stereocyclops histrio</i>	Not Evaluated	Critically Endangered	11	1	-90,91
<i>Stereocyclops incrassatus</i>	Not Evaluated	Endangered	2158,75	1133,25	-47,50
<i>Stereocyclops parkeri</i>	Not Evaluated	Critically Endangered	377,25	64,25	-82,97



**Odontophrynidae**

<i>Macrogenioglottus alipioi</i>	Not Evaluated	Vulnerable	2874	2012	-29,99
<i>Odontophrynus americanus</i>	Not Evaluated	Vulnerable	8798	4404	-49,94
<i>Odontophrynus carvalhoi</i>	Not Evaluated	Least Concern	707,25	1145,5	61,97
<i>Odontophrynus maisuma</i>	Not Evaluated	Vulnerable	8250	5381,25	-34,77
<i>Proceratophrys appendiculata</i>	Not Evaluated	Endangered	1587,25	675,75	-57,43
<i>Proceratophrys avelinoi</i>	Not Evaluated	Vulnerable	2365,5	1317	-44,32
<i>Proceratophrys bigibbosa</i>	Not Evaluated	Critically Endangered	674,5	122,25	-81,88
<i>Proceratophrys boiei</i>	Not Evaluated	Least Concern	5307,25	4325	-18,51
<i>Proceratophrys brauni</i>	Not Evaluated	Critically Endangered	384,25	28,75	-92,52
<i>Proceratophrys cristiceps</i>	Not Evaluated	Endangered	736,25	347,75	-52,77
<i>Proceratophrys cururu</i>	Not Evaluated	Extinct	1	0	-100,00
<i>Proceratophrys laticeps</i>	Not Evaluated	Endangered	1213,75	345,25	-71,56
<i>Proceratophrys melanopogon</i>	Not Evaluated	Endangered	600	135,25	-77,46
<i>Proceratophrys minuta</i>	Not Evaluated	Least Concern	3,5	73,75	2007,14
<i>Proceratophrys paviotti</i>	Not Evaluated	Extinct	3,25	0	-100,00
<i>Proceratophrys renalis</i>	Not Evaluated	Vulnerable	1530	1058,5	-30,82

<i>Proceratophrys sanctarictae</i>	Critically Endangered	Least Concern	13,25	77,75	486,79
<i>Proceratophrys schirchi</i>	Not Evaluated	Endangered	1633,25	490,75	-69,95
<i>Proceratophrys subguttata</i>	Not Evaluated	Endangered	312,5	133,5	-57,28
<b>Pipidae</b>					
<i>Pipa carvalhoi</i>	Not Evaluated	Least Concern	2311,75	1841,75	-20,33
<b>Phyllomedusidae</b>					
<i>Hylomantis aspera</i>	Not Evaluated	Endangered	174	52	-70,11
<i>Hylomantis granulosa</i>	Vulnerable	Critically Endangered	267	16,75	-93,73
<i>Phyllomedusa bahiana</i>	Not Evaluated	Critically Endangered	303,25	10,5	-96,54
<i>Phyllomedusa burmeisteri</i>	Not Evaluated	Least Concern	4791,75	4257,25	-11,15
<i>Phyllomedusa distincta</i>	Not Evaluated	Endangered	1487,75	625,5	-57,96
<i>Phyllomedusa iheringii</i>	Not Evaluated	Extinct	56,5	0	-100,00
<i>Phyllomedusa nordestina</i>	Not Evaluated	Least Concern	1889,75	2180	15,36
<i>Phyllomedusa rohdei</i>	Not Evaluated	Endangered	2174,75	916,5	-57,86
<i>Phyllomedusa tetraploidea</i>	Not Evaluated	Vulnerable	3144,25	2068,5	-34,21
<b>Ranidae</b>					
<i>Lithobates catesbeianus</i>	Not Evaluated	Least Concern	1315,25	1032,5	-21,50

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<i>Lithobates palmipes</i>	Not Evaluated	Critically Endangered	272,25	6	-97,80
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**Table S2.** List of studied amphibians species and their respective functional traits.

Species	Activity	Members	Body Size(cm)	Calling site	Poisonous	Habit	Habitat	Reproductive mode	References
<i>Adelophryne mucronatus</i>	Nocturnal	Tetrapod	1,4	Forest floor	Non toxic	Cryptic	Forested areas	Reproductive mode 23	Lourenço-de-Moraes et al 2012; Haddad et al. 2013; present study
<i>Adelophryne pachydactyla</i>	Nocturnal	Tetrapod	1,4	Forest floor	Non toxic	Cryptic	Forested areas	Reproductive mode 23	Lourenço-de-Moraes et al 2012; Haddad et al. 2013; present study
<i>Adenomera ajurauna</i>	Nocturnal	Tetrapod	1,9	Forest floor	Non toxic	Cryptic	Forested areas	Reproductive mode 32	Haddad et al 2013
<i>Adenomera araucaria</i>	Nocturnal	Tetrapod	1,9	Forest floor	Non toxic	Cryptic	Forested areas	Reproductive mode 32	Haddad et al 2013
<i>Adenomera bokermanni</i>	Nocturnal	Tetrapod	2,4	Forest floor	Non toxic	Cryptic	Forested areas	Reproductive mode 32	Haddad et al 2013
<i>Adenomera engelsi</i>	Nocturnal and Diurnal	Tetrapod	2,2	Forest floor	Non toxic	Cryptic	Forested areas	Reproductive mode 32	Haddad et al 2013
<i>Adenomera marmorata</i>	Nocturnal and Diurnal	Tetrapod	2,1	Forest floor; Low vegetation	Non toxic	Cryptic	Open or forested areas	Reproductive mode 32	Haddad et al 2013; present study
<i>Adenomera nana</i>	Nocturnal	Tetrapod	1,8	Forest floor	Non toxic	Cryptic	Forested areas	Reproductive mode 32	Haddad et al 2013; present study
<i>Adenomera thomei</i>	Nocturnal	Tetrapod	2,4	Forest floor	Non toxic	Cryptic	Forested	Reproductive	Haddad et al 2013

							areas	mode 32	
<i>Allobates alagoanus</i>	Diurnal	Tetrapod	1,5	Stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 20	Haddad et al 2013
<i>Allobates capixaba</i>	Diurnal	Tetrapod	1,6	Stream or rivulet	Non toxic	Rheophilic	Open or forested areas	Reproductive mode 20	Haddad et al 2013
<i>Allobates olfersioides</i>	Diurnal	Tetrapod	1,6	Stream or rivulet	Non toxic	Rheophilic	Open or forested areas	Reproductive mode 20	Haddad et al 2013; present study
<i>Aparasphenodon arapapa</i>	Nocturnal	Tetrapod	4,5	Bromeliad	Non toxic	Arboreal	Forested areas	Reproductive mode 6	Haddad et al 2013; Lourenço-de-Moraes et al 2013; present study
<i>Aparasphenodon bokermanni</i>	Nocturnal	Tetrapod	4,6	Swamp or pond	Non toxic	Arboreal	Forested areas	Reproductive mode 1	Haddad et al 2013
<i>Aparasphenodon brunoi</i>	Nocturnal	Tetrapod	8	Swamp or pond	Non toxic	Arboreal	Forested areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Aplastodiscus albofrenatus</i>	Nocturnal	Tetrapod	3,9	Swamp or pond; River or stream backwaters	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 5	Haddad et al 2013; present study
<i>Aplastodiscus albosignatus</i>	Nocturnal	Tetrapod	4,3	Swamp or pond; River or stream backwaters	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 5	Haddad et al 2013; present study
<i>Aplastodiscus arildae</i>	Nocturnal	Tetrapod	3,7	Swamp or pond; River or stream backwaters	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 5	Haddad et al 2013; present study

<i>Aplastodiscus callipygius</i>	Nocturnal	Tetrapod	4,6	Swamp or pond; River or stream backwaters	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 5	Haddad et al 2013
<i>Aplastodiscus cavicola</i>	Nocturnal	Tetrapod	3,7	Swamp or pond; River or stream backwaters	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 5	Haddad et al 2013
<i>Aplastodiscus cochranæ</i>	Nocturnal	Tetrapod	4,5	Swamp or pond; River or stream backwaters	Unpalatable or bad odor	Arboreal	Open or forested areas	Reproductive mode 5	Haddad et al 2013
<i>Aplastodiscus ehrhardti</i>	Nocturnal	Tetrapod	3,1	Swamp or pond; River or stream backwaters	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 5	Haddad et al 2013; present study
<i>Aplastodiscus eugenioi</i>	Nocturnal	Tetrapod	3,5	Swamp or pond; River or stream backwaters	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 5	Haddad et al 2013
<i>Aplastodiscus flumineus</i>	Nocturnal	Tetrapod	4,5	Swamp or pond; River or stream backwaters	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 5	Haddad et al 2013
<i>Aplastodiscus ibirabitanga</i>	Nocturnal	Tetrapod	4,1	Swamp or pond; River or stream backwaters	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 5	Haddad et al 2013; present study
<i>Aplastodiscus leucoeygius</i>	Nocturnal	Tetrapod	4,5	Swamp or pond; River or stream backwaters	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 5	Haddad et al 2013
<i>Aplastodiscus perviridis</i>	Nocturnal	Tetrapod	4,5	Swamp or pond; River or stream backwaters	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 5	Haddad et al 2013; present study

<i>Aplastodiscus sibilatus</i>	Nocturnal	Tetrapod	3,2	Swamp or pond; River or stream backwaters	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 5	Haddad et al 2013
<i>Aplastodiscus weygoldti</i>	Nocturnal	Tetrapod	3,8	Swamp or pond; River or stream backwaters	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 5	Haddad et al 2013; present study
<i>Arcovomer passarellii</i>	Nocturnal	Tetrapod	2,4	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 1	Haddad et al 2013
<i>Bokermannohyla ahenea</i>	Nocturnal	Tetrapod	5,2	Swamp or pond; River or stream backwaters	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 1 and 2	Haddad et al 2013
<i>Bokermannohyla astartea</i>	Nocturnal	Tetrapod	4,4	Bromeliad;	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 6	Haddad et al 2013; present study
<i>Bokermannohyla capra</i>	Nocturnal	Tetrapod	6,1	Bromeliad; Stream or rivulet	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 2	Haddad et al 2013
<i>Bokermannohyla caramaschii</i>	Nocturnal	Tetrapod	6,2	River, stream or rivulet	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 2	Haddad et al 2013; present study
<i>Bokermannohyla carvalhoi</i>	Nocturnal	Tetrapod	6,4	River, stream or rivulet	Unpalatable or bad odor	Terrestrial	Forested areas	Reproductive mode 2	Haddad et al 2013
<i>Bokermannohyla circumdata</i>	Nocturnal	Tetrapod	5,4	Swamp or pond	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 2 and 4	Haddad et al 2013; present study
<i>Bokermannohyla diamantina</i>	Nocturnal	Tetrapod	4,9	River, stream or rivulet	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 2	Haddad et al 2013
<i>Bokermannohyla gouveai</i>	Nocturnal	Tetrapod	6,9	Swamp or pond	Unpalatable or bad odor	Arboreal	Open or forested	0	Haddad et al 2013





<i>Brachycephalus didactylus</i>	Diurnal	Tetrapod	0,8	Forest floor	Non toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013
<i>Brachycephalus ephippium</i>	Diurnal	Tetrapod	1,9	Forest floor	Toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013; present study
<i>Brachycephalus ferruginus</i>	Diurnal	Tetrapod	1,4	Forest floor	Toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013
<i>Brachycephalus garbeanus</i>	Diurnal	Tetrapod	1,7	Forest floor	Toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013
<i>Brachycephalus guarani</i>	Diurnal	Tetrapod	1,3	Forest floor	Toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013
<i>Brachycephalus hermogenesi</i>	Diurnal	Tetrapod	1,1	Forest floor	Non toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013
<i>Brachycephalus izecksohni</i>	Diurnal	Tetrapod	1,3	Forest floor	Non toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013
<i>Brachycephalus margaritatus</i>	Diurnal	Tetrapod	1,7	Forest floor	Toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013
<i>Brachycephalus nodoterga</i>	Diurnal	Tetrapod	1,3	Forest floor	Toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013
<i>Brachycephalus pernix</i>	Diurnal	Tetrapod	1,5	Forest floor	Toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013
<i>Brachycephalus pitanga</i>	Diurnal	Tetrapod	1,3	Forest floor	Toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013; present study
<i>Brachycephalus pombali</i>	Diurnal	Tetrapod	1,5	Forest floor	Toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013

<i>Brachycephalus pulex</i>	-----	Tetrapod	0,8	Forest floor	Non toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013
<i>Brachycephalus toby</i>	Diurnal	Tetrapod	1,4	Forest floor	Toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013
<i>Brachycephalus tridactylus</i>	Diurnal	Tetrapod	1,4	Forest floor	Toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013
<i>Brachycephalus vertebralis</i>	Diurnal	Tetrapod	1,4	Forest floor	Toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013
<i>Ceratophrys aurita</i>	Nocturnal	Tetrapod	14,9	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 1	Haddad et al 2013
<i>Chiasmocleis alagoana</i>	-----	Tetrapod	2,7	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 1	Haddad et al 2013
<i>Chiasmocleis atlantica</i>	Nocturnal	Tetrapod	3,2	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 1	Haddad et al 2013
<i>Chiasmocleis capixaba</i>	Nocturnal	Tetrapod	1,5	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Chiasmocleis carvalhoi</i>	Nocturnal	Tetrapod	2,3	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 1	Haddad et al 2013
<i>Chiasmocleis crucis</i>	Nocturnal	Tetrapod	2	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 1	Haddad et al 2013
<i>Chiasmocleis gnomia</i>	Nocturnal	Tetrapod	1,6	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 1	Haddad et al 2013
<i>Chiasmocleis leucosticta</i>	Nocturnal and	Tetrapod	2,5	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 10	Haddad et al 2013; present study

Diurnal										
<i>Chiasmocleis mantiqueira</i>	Nocturnal	Tetrapod	2,2	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 1	Haddad et al 2013	
<i>Chiasmocleis schubarti</i>	Nocturnal	Tetrapod	3,3	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 1	Haddad et al 2013; present study	
<i>Chthonerpeton indistinctum</i>	-----	Apod	43,5	Do not Call	-----	Fossorial; Aquatic	Open or forested areas	Reproductive mode 39	Haddad et al 2013	
<i>Crossodactylodes bokermanni</i>	Nocturnal	Tetrapod	1,4	Bromeliad	Non toxic	Arboreal	Forested areas	Reproductive mode 6	Haddad et al 2013	
<i>Crossodactylodes izecksohni</i>	Nocturnal	Tetrapod	1,5	Bromeliad	Non toxic	Arboreal	Forested areas	Reproductive mode 6	Haddad et al 2013	
<i>Crossodactylus aeneus</i>	-----	Tetrapod	2,3	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013	
<i>Crossodactylus bokermanni</i>	-----	Tetrapod	2,7	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013	
<i>Crossodactylus caramaschii</i>	Nocturnal and Diurnal	Tetrapod	2,3	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013; present study	
<i>Crossodactylus dantei</i>	-----	Tetrapod	2,2	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013	
<i>Crossodactylus dispar</i>	Diurnal	Tetrapod	3,1	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013	
<i>Crossodactylus</i>	Nocturnal	Tetrapod	3,1	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013	

<i>gaudichaudii</i>	and Diurnal			rivulet			areas	mode 3	
<i>Crossodactylus grandis</i>	Diurnal	Tetrapod	4,2	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013
<i>Crossodactylus schmidtii</i>	Nocturnal	Tetrapod	2,8	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013; present study
<i>Cycloramphus acangatan</i>	Nocturnal	Tetrapod	4,3	Forest floor	-----	Cryptic	Forested areas	Reproductive mode 21	Haddad et al 2013; present study
<i>Cycloramphus bandeirensis</i>	-----	Tetrapod	4,5	-----	-----	Rheophilic	Open areas	Reproductive mode 19	Haddad et al 2013
<i>Cycloramphus bolitoglossus</i>	Nocturnal	Tetrapod	3,8	Forest floor	-----	Cryptic	Forested areas	Reproductive mode 21	Haddad et al 2013
<i>Cycloramphus boraceiensis</i>	Nocturnal	Tetrapod	4,7	River, stream or rivulet	Unpalatable or bad odor	Rheophilic	Forested areas	Reproductive mode 19	Haddad et al 2013; present study
<i>Cycloramphus brasiliensis</i>	Nocturnal	Tetrapod	6,2	River, stream or rivulet	-----	Rheophilic	Forested areas	Reproductive mode 19	Haddad et al 2013; present study
<i>Cycloramphus carvalhoi</i>	-----	Tetrapod	5,6	Forest floor	-----	Cryptic	Forested areas	Reproductive mode 21	Haddad et al 2013
<i>Cycloramphus catarinensis</i>	Nocturnal	Tetrapod	3,5	River, stream or rivulet	-----	Rheophilic	Forested areas	Reproductive mode 19	Haddad et al 2013
<i>Cycloramphus dubius</i>	Nocturnal	Tetrapod	4,9	River, stream or rivulet	-----	Rheophilic	Forested areas	Reproductive mode 19	Haddad et al 2013
<i>Cycloramphus eleutherodactylus</i>	Nocturnal	Tetrapod	4,8	Caves, burrows	-----	Cryptic	Forested areas	Reproductive mode 21	Haddad et al 2013; present study

<i>Cycloramphus izecksohni</i>	Nocturnal	Tetrapod	4	River, stream or rivulet	-----	Rheophilic	Forested areas	Reproductive mode 19	Haddad et al 2013
<i>Cycloramphus juimirim</i>	Nocturnal	Tetrapod	3	River, stream or rivulet	-----	Rheophilic	Forested areas	Reproductive mode 19	Haddad et al 2013
<i>Cycloramphus lithomimeticus</i>	Nocturnal	Tetrapod	3,1	River, stream or rivulet	-----	Rheophilic	Forested areas	Reproductive mode 19	Haddad et al 2013
<i>Cycloramphus lutzorum</i>	Nocturnal	Tetrapod	4,1	River, stream or rivulet	-----	Rheophilic	Forested areas	Reproductive mode 19	Haddad et al 2013
<i>Cycloramphus migueli</i>	-----	Tetrapod	4,2	-----	-----	Cryptic	Forested areas	Reproductive mode 21	Haddad et al 2013
<i>Cycloramphus organensis</i>	Nocturnal	Tetrapod	3,2	-----	-----	-----	Open areas	-----	Haddad et al 2013
<i>Cycloramphus rhyakonastes</i>	Nocturnal	Tetrapod	4,2	River, stream or rivulet	-----	Rheophilic	Forested areas	Reproductive mode 19	Haddad et al 2013
<i>Cycloramphus valae</i>	-----	Tetrapod	3,6	River, stream or rivulet	-----	Rheophilic	Forested areas	Reproductive mode 19	Haddad et al 2013
<i>Dasypops schirchi</i>	Nocturnal	Tetrapod	5,2	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 1	Haddad et al 2013
<i>Dendrophryniscus berthallutzae</i>	Nocturnal	Tetrapod	2,4	Bromeliad	Non toxic	Cryptic	Forested areas	Reproductive mode 8	Haddad et al 2013
<i>Dendrophryniscus brevipollicatus</i>	Nocturnal	Tetrapod	2	Bromeliad	Non toxic	Arboreal	Forested areas	Reproductive mode 8	Haddad et al 2013; present study
<i>Dendrophryniscus carvalhoi</i>	-----	Tetrapod	1,9	Bromeliad	Non toxic	Cryptic	Forested areas	Reproductive mode 8	Haddad et al 2013

<i>Dendrophryniscus krausae</i>	-----	Tetrapod	2,3	-----	Non toxic	Arboreal; Cryptic	Forested areas	Reproductive mode 8	Haddad et al 2013
<i>Dendrophryniscus leucomystax</i>	Nocturnal	Tetrapod	2,5	Forest floor	Non toxic	Cryptic	Forested areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Dendrophryniscus oreites</i>	-----	Tetrapod	2,5	-----	Non toxic	Arboreal	Forested areas	-----	Haddad et al 2013
<i>Dendrophryniscus proboscideus</i>	Nocturnal	Tetrapod	4,1	-----	Toxic	Cryptic	Forested areas	-----	Haddad et al 2013; present study
<i>Dendropsophus anceps</i>	Nocturnal	Tetrapod	4,2	Swamp or pond	Non toxic	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013
<i>Dendropsophus berthallutzae</i>	Nocturnal	Tetrapod	2,4	Swamp or pond	Non toxic	Arboreal	Forested areas	Reproductive mode 24	Haddad et al 2013
<i>Dendropsophus bipunctatus</i>	Nocturnal	Tetrapod	2,6	Swamp or pond	Non toxic	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Dendropsophus branneri</i>	Nocturnal	Tetrapod	2,3	Swamp or pond	Unpalatable or bad odor	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Dendropsophus decipiens</i>	Nocturnal	Tetrapod	1,9	Swamp or pond	Non toxic	Arboreal	Open or forested areas	Reproductive mode 24	Haddad et al 2013; present study
<i>Dendropsophus elegans</i>	Nocturnal	Tetrapod	3,1	Swamp or pond	Non toxic	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Dendropsophus giesleri</i>	Nocturnal	Tetrapod	2,7	Swamp or pond	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 1	Haddad et al 2013; present study

<i>Dendropsophus haddadi</i>	Nocturnal	Tetrapod	2,3	Swamp or pond	Non toxic	Arboreal	Open or forested areas	Reproductive mode 24	Lourenço-de-Moraes et al 2012; Haddad et al 2013; present study
<i>Dendropsophus meridianus</i>	Nocturnal	Tetrapod	1,8	Swamp or pond	Non toxic	Arboreal	Open or forested areas	Reproductive mode 1	Haddad et al 2013
<i>Dendropsophus microps</i>	Nocturnal	Tetrapod	2,9	Swamp or pond	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Dendropsophus minutus</i>	Nocturnal	Tetrapod	2,5	Swamp or pond	Unpalatable or bad odor	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Dendropsophus nahderere</i>	Nocturnal	Tetrapod	4,6	Swamp or pond	Non toxic	Arboreal	Open or forested areas	Reproductive mode 1	Haddad et al 2013
<i>Dendropsophus nanus</i>	Nocturnal	Tetrapod	2,3	Swamp or pond	Unpalatable or bad odor	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Dendropsophus novaisi</i>	Nocturnal	Tetrapod	3,4	Swamp or pond	Non toxic	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013
<i>Dendropsophus oliveirai</i>	Nocturnal	Tetrapod	1,9	Swamp or pond	Non toxic	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013
<i>Dendropsophus pseudomeridianus</i>	Nocturnal	Tetrapod	1,7	Swamp or pond	Non toxic	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013
<i>Dendropsophus ruschii</i>	Nocturnal	Tetrapod	2,8	River, stream or rivulet	Non toxic	Arboreal	Forested areas	Reproductive mode 25	Haddad et al 2013
<i>Dendropsophus sanborni</i>	Nocturnal	Tetrapod	2	Swamp or pond	Unpalatable or bad odor	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013; present study

<i>Dendropsophus seniculus</i>	Nocturnal	Tetrapod	3,8	Swamp or pond	Non toxic	Arboreal	Open or forested areas	Reproductive mode 1	Haddad et al 2013
<i>Dendropsophus soaresi</i>	Nocturnal	Tetrapod	3,5	Swamp or pond	Non toxic	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013
<i>Dendropsophus studeræ</i>	Nocturnal	Tetrapod	2,8	Swamp or pond	Non toxic	Arboreal	Forested areas	Reproductive mode 1	Haddad et al 2013
<i>Dendropsophus wernerii</i>	Nocturnal	Tetrapod	1,9	Swamp or pond	Non toxic	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013
<i>Elaschitocleis bicolor</i>	Nocturnal	Tetrapod	3,8	Swamp or pond	Non toxic	Fossorial	Open areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Elaschitocleis cesarii</i>	Nocturnal	Tetrapod	3,2	Swamp or pond	Non toxic	Fossorial	Open areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Elaschitocleis erythrogaster</i>	Nocturnal	Tetrapod	3,5	Swamp or pond	Non toxic	Fossorial	Open areas	Reproductive mode 1	Haddad et al 2013
<i>Eleutherodactylus bilineatus</i>	Nocturnal	Tetrapod	2,6	-----	Non toxic	Arboreal; Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013; present study
<i>Euparkerella brasiliensis</i>	Nocturnal and Diurnal	Tetrapod	1,7	Forest floor	Non toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013
<i>Euparkerella cochranæ</i>	Nocturnal and Diurnal	Tetrapod	1,6	Forest floor	Non toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013; present study
<i>Euparkerella robusta</i>	Nocturnal and	Tetrapod	2,3	Forest floor	Non toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013



	Diurnal									
<i>Euparkerella tridactyla</i>	Nocturnal and Diurnal	Tetrapod	1,5	Forest floor	Non toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013; present study	
<i>Fritziana fissilis</i>	Nocturnal	Tetrapod	2,8	Bromeliad	Non toxic	Arboreal	Forested areas	Reproductive mode 36	Haddad et al 2013; present study	
<i>Fritziana goeldii</i>	Nocturnal	Tetrapod	3,7	Bromeliad	Non toxic	Arboreal	Forested areas	Reproductive mode 36	Haddad et al 2013	
<i>Fritziana ohausi</i>	Nocturnal	Tetrapod	3,1	Bamboo Grove	Non toxic	Arboreal	Forested areas	Reproductive mode 36	Haddad et al 2013; present study	
<i>Frostius erythrophthalmus</i>	Nocturnal	Tetrapod	2,3	Vegetation	Toxic	Arboreal; Cryptic	Forested areas	-----	Haddad et al 2013; present study	
<i>Frostius pernambucensis</i>	Nocturnal	Tetrapod	2,1	Bromeliad	Toxic	Arboreal; Cryptic	Forested areas	Reproductive mode 8	Haddad et al 2013; present study	
<i>Gastrotheca albolineata</i>	Nocturnal	Tetrapod	6	Tree crown	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 37	Haddad et al 2013; present study	
<i>Gastrotheca ernestoi</i>	Nocturnal	Tetrapod	6,9	Tree crown	Non toxic	Arboreal	Forested areas	Reproductive mode 37	Haddad et al 2013	
<i>Gastrotheca fissipes</i>	Nocturnal	Tetrapod	7,2	Tree crown	Non toxic	Arboreal	Forested areas	Reproductive mode 37	Haddad et al 2013	
<i>Gastrotheca fulvorufa</i>	Nocturnal	Tetrapod	6,9	Tree crown	Non toxic	Arboreal	Forested areas	Reproductive mode 37	Haddad et al 2013	
<i>Gastrotheca megacephala</i>	Nocturnal	Tetrapod	10,1	Tree crown	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 37	Haddad et al 2013; present study	

<i>Gastrotheca microdiscus</i>	Nocturnal	Tetrapod	5,9	Tree crown	Non toxic	Arboreal	Forested areas	Reproductive mode 37	Haddad et al 2013
<i>Gastrotheca prasina</i>	Nocturnal	Tetrapod	9,7	Tree crown	Non toxic	Arboreal	Forested areas	Reproductive mode 37	Haddad et al 2013
<i>Gastrotheca pulchra</i>	Nocturnal	Tetrapod	3,3	Tree crown	Non toxic	Arboreal	Forested areas	Reproductive mode 37	Haddad et al 2013
<i>Gastrotheca recava</i>	Nocturnal	Tetrapod	7,6	Tree crown	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 37	Haddad et al 2013; present study
<i>Haddadus aramunha</i>	Nocturnal	Tetrapod	6,2	Forest floor	Non toxic	Cryptic	Open areas	Reproductive mode 23	Haddad et al 2013
<i>Haddadus binotatus</i>	Nocturnal	Tetrapod	6,3	Forest floor	Unpalatable or bad odor	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013; present study
<i>Holoaden bradei</i>	-----	Tetrapod	2,5	-----	Toxic	Cryptic	Open or forested areas	Reproductive mode 23	Haddad et al 2013
<i>Holoaden luederwaldti</i>	Nocturnal	Tetrapod	4,6	Forest floor	Toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013
<i>Holoaden pholeter</i>	Nocturnal	Tetrapod	4,6	-----	Toxic	Cryptic	Open or forested areas	Reproductive mode 23	Haddad et al 2013
<i>Hylodes amnicola</i>	Diurnal	Tetrapod	2,7	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013
<i>Hylodes asper</i>	Diurnal	Tetrapod	4,7	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013; present study

<i>Hylodes babax</i>	Diurnal	Tetrapod	3,2	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013
<i>Hylodes cardosoi</i>	Diurnal	Tetrapod	4	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013
<i>Hylodes charadranaetes</i>	Diurnal	Tetrapod	4,3	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013
<i>Hylodes dactylocinus</i>	Diurnal	Tetrapod	2,7	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013; present study
<i>Hylodes heyeri</i>	Diurnal	Tetrapod	4,3	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013
<i>Hylodes lateristrigatus</i>	Diurnal	Tetrapod	3,5	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013
<i>Hylodes magalhaesi</i>	Diurnal	Tetrapod	3,4	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013
<i>Hylodes meridionalis</i>	Diurnal	Tetrapod	4,8	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013; present study
<i>Hylodes nasus</i>	Diurnal	Tetrapod	3,8	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013
<i>Hylodes ornatus</i>	Diurnal	Tetrapod	2,7	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013
<i>Hylodes otavioi</i>	Diurnal	Tetrapod	3,4	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013
<i>Hylodes perere</i>	Diurnal	Tetrapod	2,7	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013

<i>Hylodes perpicatus</i>	Diurnal	Tetrapod	4,4	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013
<i>Hylodes phyllodes</i>	Diurnal	Tetrapod	3	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013; present study
<i>Hylodes pipilans</i>	Diurnal	Tetrapod	2,8	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013; present study
<i>Hylodes regius</i>	Diurnal	Tetrapod	3,5	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013
<i>Hylodes sazimai</i>	Diurnal	Tetrapod	2,8	River, stream or rivulet	Non toxic	Rheophilic	Forested areas	Reproductive mode 3	Haddad et al 2013
<i>Hylomantis aspera</i>	Nocturnal	Tetrapod	4,4	Swamp or pond	Toxic	Arboreal	Forested areas	Reproductive mode 18	Haddad et al 2013; present study
<i>Hylomantis granulosa</i>	Nocturnal	Tetrapod	4,8	River, stream or rivulet	Toxic	Arboreal	Forested areas	Reproductive mode 25	Haddad et al 2013
<i>Hyophryne histrio</i>	Nocturnal	Tetrapod	4,8	Swamp or pond	-----	Cryptic	Forested areas	Reproductive mode 1	Haddad et al 2013
<i>Hypsiboas albomarginatus</i>	Nocturnal	Tetrapod	5,8	Swamp or pond	Unpalatable or bad odor	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Hypsiboas albopunctatus</i>	Nocturnal	Tetrapod	6,5	Swamp or pond	Unpalatable or bad odor	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Hypsiboas atlanticus</i>	Nocturnal	Tetrapod	4	Swamp or pond; River or stream backwaters	Non toxic	Arboreal	Open areas	Reproductive mode 1 and 2	Haddad et al 2013; present study
<i>Hypsiboas</i>	Nocturnal	Tetrapod	5,9	Swamp or pond	Unpalatable	Arboreal	Open areas	Reproductive	Haddad et al 2013; present

<i>bischoffi</i>					or bad odor			mode 1	study
<i>Hypsiboas caingua</i>	Nocturnal	Tetrapod	3,9	Swamp or pond	Unpalatable or bad odor	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Hypsiboas caipora</i>	Nocturnal	Tetrapod	4	River, stream or rivulet	Non toxic	Arboreal	Forested areas	Reproductive mode 2	Haddad et al 2013
<i>Hypsiboas crepitans</i>	Nocturnal	Tetrapod	6,2	Swamp or pond	Unpalatable or bad odor	Arboreal	Open areas	Reproductive mode 4	Haddad et al 2013; present study
<i>Hypsiboas curupi</i>	Nocturnal	Tetrapod	4,5	River, stream or rivulet	Non toxic	Arboreal	Forested areas	Reproductive mode 2	Haddad et al 2013
<i>Hypsiboas exastis</i>	Nocturnal	Tetrapod	9,2	Swamp or pond; River or stream backwaters	Non toxic	Arboreal	Forested areas	Reproductive mode 4	Haddad et al 2013; present study
<i>Hypsiboas faber</i>	Nocturnal	Tetrapod	8,9	Swamp or pond	Unpalatable or bad odor	Arboreal	Open or forested areas	Reproductive mode 1 and 4	Haddad et al 2013; present study
<i>Hypsiboas freicanecae</i>	Nocturnal	Tetrapod	4	River, stream or rivulet	Non toxic	Arboreal	Forested areas	Reproductive mode 2	Haddad et al 2013
<i>Hypsiboas guentheri</i>	Nocturnal	Tetrapod	4,6	Swamp or pond	Non toxic	Arboreal	Open or forested areas	Reproductive mode 1	Haddad et al 2013
<i>Hypsiboas joaquina</i>	Nocturnal	Tetrapod	5,4	River, stream or rivulet	Non toxic	Arboreal	Open or forested areas	Reproductive mode 2	Haddad et al 2013
<i>Hypsiboas latistriatus</i>	Nocturnal	Tetrapod	4,7	River, stream or rivulet	Non toxic	Arboreal	Forested areas	Reproductive mode 2	Haddad et al 2013

<i>Hypsiboas leptolineatus</i>	Nocturnal	Tetrapod	3,3	Swamp or pond; River or stream backwaters	Non toxic	Arboreal	Open areas	Reproductive mode 1 and 2	Haddad et al 2013
<i>Hypsiboas lundii</i>	Nocturnal	Tetrapod	6,9	Stream or rivulet	Non toxic	Arboreal	Open or forested areas	Reproductive mode 2	Haddad et al 2013
<i>Hypsiboas marginatus</i>	Nocturnal	Tetrapod	5,3	River, stream or rivulet	Non toxic	Arboreal	Forested areas	Reproductive mode 2	Haddad et al 2013
<i>Hypsiboas pardalis</i>	Nocturnal	Tetrapod	6,8	Swamp or pond	Unpalatable or bad odor	Arboreal	Open or forested areas	Reproductive mode 4	Haddad et al 2013; present study
<i>Hypsiboas poaju</i>	Nocturnal	Tetrapod	3,9	River, stream or rivulet	Non toxic	Arboreal	Forested areas	Reproductive mode 2	Haddad et al 2013
<i>Hypsiboas polytaenius</i>	Nocturnal	Tetrapod	3,6	-----	Non toxic	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013
<i>Hypsiboas pombali</i>	Nocturnal	Tetrapod	5,7	Swamp or pond; River or stream backwaters	Non toxic	Arboreal	Forested areas	Reproductive mode 1 and 2	Haddad et al 2013; present study
<i>Hypsiboas prasinus</i>	Nocturnal	Tetrapod	4,8	Swamp or pond; River or stream backwaters	Unpalatable or bad odor	Arboreal	Open or forested areas	Reproductive mode 1 and 2	Haddad et al 2013; present study
<i>Hypsiboas pulchellus</i>	Nocturnal	Tetrapod	4,5	Swamp or pond	Non toxic	Arboreal	Open or forested areas	Reproductive mode 1	Haddad et al 2013
<i>Hypsiboas punctatus</i>	Nocturnal	Tetrapod	3,9	Swamp or pond; River or stream	Unpalatable or bad odor	Arboreal	Open areas	Reproductive mode 1 and 2	Haddad et al 2013; present study

## backwaters

<i>Hypsiboas raniceps</i>	Nocturnal	Tetrapod	7,1	Swamp or pond	Unpalatable or bad odor	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Hypsiboas secedens</i>	Nocturnal	Tetrapod	4,9	-----	Non toxic	Arboreal	Forested areas	Reproductive mode 2	Haddad et al 2013
<i>Hypsiboas semiguttatus</i>	Nocturnal	Tetrapod	4,1	River, stream or rivulet	Non toxic	Arboreal	Forested areas	Reproductive mode 1 and 2	Haddad et al 2013
<i>Hypsiboas semilineatus</i>	Nocturnal	Tetrapod	5,6	Swamp or pond; River or stream backwaters	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 1 and 2	Haddad et al 2013; present study
<i>Hypsiboas stellae</i>	Nocturnal	Tetrapod	4,6	River, stream or rivulet	Non toxic	Arboreal	Forested areas	Reproductive mode 1 and 2	Haddad et al 2013
<i>Hypsiboas stenocephalus</i>	Nocturnal	Tetrapod	2,7	Swamp or pond; River or stream backwaters	Non toxic	Arboreal	Open areas	Reproductive mode 1 and 2	Haddad et al 2013
<i>Ischnocnema abdita</i>	-----	Tetrapod	1,6	Forest floor	Non toxic	Cryptic	Forested areas	Reproductive mode 1 and 2	Haddad et al 2013; present study
<i>Ischnocnema bolbodactyla</i>	-----	Tetrapod	1,9	Forest floor	Non toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013
<i>Ischnocnema concolor</i>	Nocturnal	Tetrapod	2	Forest floor; Low vegetation	Non toxic	Cryptic	Open or forested areas	Reproductive mode 23	Haddad et al 2013
<i>Ischnocnema erythromera</i>	Nocturnal	Tetrapod	3	Forest floor	Unpalatable or bad odor	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013; present study

<i>Ischnocnema guentheri</i>	Nocturnal	Tetrapod	4	Forest floor	Unpalatable or bad odor	Arboreal; Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013; present study
<i>Ischnocnema henselii</i>	Nocturnal	Tetrapod	3,6	Forest floor	Unpalatable or bad odor	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013; present study
<i>Ischnocnema hoehnei</i>	-----	Tetrapod	2,9	Forest floor	Non toxic	Arboreal; Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013
<i>Ischnocnema holti</i>	Nocturnal	Tetrapod	1,7	Forest floor; Low vegetation	Non toxic	Arboreal; Cryptic	Open or forested areas	Reproductive mode 23	Haddad et al 2013
<i>Ischnocnema izecksohni</i>	-----	Tetrapod	4,7	Forest floor	Non toxic	Cryptic	Forested areas	-----	Haddad et al 2013
<i>Ischnocnema juipoca</i>	-----	Tetrapod	2,3	Forest floor; Low vegetation	Non toxic	Cryptic	Open or forested areas	Reproductive mode 23	Haddad et al 2013
<i>Ischnocnema karst</i>	-----	Tetrapod	2,3	Forest floor	Non toxic	Cryptic	Forested areas	-----	Haddad et al 2013
<i>Ischnocnema manezinho</i>	Nocturnal	Tetrapod	3,3	River, stream or rivulet	Non toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013
<i>Ischnocnema melanopygia</i>	-----	Tetrapod	2	Forest floor	Non toxic	Cryptic	Forested areas	-----	Haddad et al 2013
<i>Ischnocnema nasuta</i>	-----	Tetrapod	3,9	Bromeliad	Non toxic	Arboreal; Cryptic	Forested areas	Reproductive mode 27	Haddad et al 2013
<i>Ischnocnema nigriventris</i>	-----	Tetrapod	1,9	Forest floor	Non toxic	Arboreal; Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013



<i>Ischnocnema octavioi</i>	-----	Tetrapod	3,2	Forest floor	Non toxic	Cryptic	Forested areas	-----	Haddad et al 2013
<i>Ischnocnema oea</i>	Nocturnal	Tetrapod	2,2	Forest floor	Non toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013
<i>Ischnocnema parva</i>	Nocturnal	Tetrapod	2	Forest floor	Non toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013; present study
<i>Ischnocnema randorum</i>	Nocturnal	Tetrapod	1,8	Forest floor; Low vegetation	Non toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013
<i>Ischnocnema sambaqui</i>	Nocturnal	Tetrapod	3,7	River, stream or rivulet	Non toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013
<i>Ischnocnema spanios</i>	-----	Tetrapod	2,1	Forest floor	Non toxic	Cryptic	Forested areas	-----	Haddad et al 2013; present study
<i>Ischnocnema verrucosa</i>	Nocturnal	Tetrapod	2,4	Forest floor	Non toxic	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013
<i>Ischnocnema vizottoi</i>	Nocturnal	Tetrapod	2	Forest floor; Low vegetation	Non toxic	Arboreal; Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013
<i>Itapotihyla langsdorffii</i>	Nocturnal	Tetrapod	10,3	Swamp or pond	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Leptodactylus cupreus</i>	Nocturnal	Tetrapod	5,7	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 30	Haddad et al 2013
<i>Leptodactylus flavopictus</i>	Nocturnal	Tetrapod	13,2	Swamp or pond	Toxic	Cryptic	Forested areas	Reproductive mode 13	Haddad et al 2013
<i>Leptodactylus furnarius</i>	Nocturnal	Tetrapod	4,2	Swamp or pond	Non toxic	Arboreal; Aquatic	Open areas	Reproductive mode 30	Haddad et al 2013

<i>Leptodactylus fuscus</i>	Nocturnal	Tetrapod	4,4	Swamp or pond	Non toxic	Terrestrial	Open areas	Reproductive mode 30	Haddad et al 2013; present study
<i>Leptodactylus gracilis</i>	Nocturnal	Tetrapod	4,4	Swamp or pond	Non toxic	Cryptic	Open areas	Reproductive mode 30	Haddad et al 2013
<i>Leptodactylus jolyi</i>	Nocturnal	Tetrapod	5,4	Swamp or pond	Non toxic	Arboreal; Aquatic	Open areas	Reproductive mode 30	Haddad et al 2013
<i>Leptodactylus labyrinthicus</i>	Nocturnal	Tetrapod	15,5	Swamp or pond	Toxic	Terrestrial	Open areas	Reproductive mode 11	Haddad et al 2013; present study
<i>Leptodactylus latrans</i>	Nocturnal	Tetrapod	9,6	Swamp or pond	Toxic	Terrestrial	Open or forested areas	Reproductive mode 11	Haddad et al 2013; present study
<i>Leptodactylus macrosternum</i>	Nocturnal	Tetrapod	9	Swamp or pond	Toxic	Terrestrial	Open or forested áreas	Reproductive mode 11	Haddad et al 2013
<i>Leptodactylus mystaceus</i>	Nocturnal	Tetrapod	5,2	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 30	Haddad et al 2013; present study
<i>Leptodactylus mystacinus</i>	Nocturnal	Tetrapod	5,5	Swamp or pond	Toxic	Terrestrial	Open areas	Reproductive mode 30	Haddad et al 2013; present study
<i>Leptodactylus natalensis</i>	Nocturnal	Tetrapod	5,2	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 13	Haddad et al 2013; present study
<i>Leptodactylus notoaktites</i>	Nocturnal	Tetrapod	5	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 30	Haddad et al 2013; present study
<i>Leptodactylus plaumanni</i>	Nocturnal	Tetrapod	3,8	Swamp or pond	Non toxic	Cryptic	Open areas	Reproductive mode 30	Haddad et al 2013

<i>Leptodactylus podicipinus</i>	Nocturnal	Tetrapod	4,2	Swamp or pond	Non toxic	Cryptic	Open areas	Reproductive mode 13	Haddad et al 2013; present study
<i>Leptodactylus spixii</i>	Nocturnal	Tetrapod	5,2	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 30	Haddad et al 2013
<i>Leptodactylus troglodytes</i>	Nocturnal	Tetrapod	5,1	Swamp or pond	Non toxic	Terrestrial	Open areas	Reproductive mode 30	Haddad et al 2013
<i>Leptodactylus vastus</i>	Nocturnal	Tetrapod	15	Swamp or pond	Toxic	Terrestrial	Open areas	Reproductive mode 11	Haddad et al 2013
<i>Leptodactylus viridis</i>	Nocturnal	Tetrapod	7,2	Swamp or pond	Toxic	Terrestrial	Open areas	Reproductive mode 11	Haddad et al 2013
<i>Limnomedusa macroglossa</i>	Nocturnal	Tetrapod	5,4	Stream or rivulet	-----	Terrestrial	Open or forested areas	Reproductive mode 1	Haddad et al 2013
<i>Lithobates catesbeianus</i>	Nocturnal	Tetrapod	8,9	Swamp or pond	Non toxic	Semi-aquatic	Open or forested areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Lithobates palmipes</i>	Nocturnal	Tetrapod	11,3	Swamp or pond	Non toxic	Semi-aquatic	Open or forested areas	Reproductive mode 1	Haddad et al 2013
<i>Luetkenotyphlus brasiliensis</i>	-----	Apod	27	Do not Call	-----	Fossorial	Open or forested areas	Reproductive mode 38	Haddad et al 2013
<i>Macrogenioglottus alipioi</i>	Nocturnal	Tetrapod	9,1	Swamp or pond	Toxic	Cryptic	Forested areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Megaelosia</i>	Diurnal	Tetrapod	8,1	-----	Non toxic	Rheophilic	Forested	-----	Haddad et al 2013

<i>apuana</i>							areas		
<i>Megaelosia bocainensis</i>	Diurnal	Tetrapod	10,1	-----	Non toxic	Rheophilic	Forested areas	-----	Haddad et al 2013
<i>Megaelosia goeldii</i>	Diurnal	Tetrapod	10	-----	Non toxic	Rheophilic	Forested areas	-----	Haddad et al 2013; present study
<i>Megaelosia massarti</i>	Diurnal	Tetrapod	11,5	-----	Non toxic	Rheophilic	Forested areas	-----	Haddad et al 2013
<i>Melanophryniscus admirabilis</i>	Diurnal	Tetrapod	3,8	Swamp or pond	Toxic	Terrestrial	Forested areas	Reproductive mode 1	Haddad et al 2013
<i>Melanophryniscus alipioi</i>	Diurnal	Tetrapod	2,5	Bromeliad	Toxic	Phytotelmata	Open areas	Reproductive mode 6	Haddad et al 2013; present study
<i>Melanophryniscus cambaraensis</i>	Nocturnal and Diurnal	Tetrapod	3,5	Swamp or pond	Toxic	Terrestrial	Open or forested areas	Reproductive mode 1	Haddad et al 2013
<i>Melanophryniscus dorsalis</i>	Diurnal	Tetrapod	2,5	Swamp or pond	Toxic	Terrestrial	Open areas	Reproductive mode 1	Haddad et al 2013
<i>Melanophryniscus macrogranulosus</i>	-----	Tetrapod	3,5	River, stream or rivulet	Toxic	Terrestrial	Forested areas	Reproductive mode 2	Haddad et al 2013
<i>Melanophryniscus moreirae</i>	Diurnal	Tetrapod	2,9	Swamp or pond	Toxic	Terrestrial	Open areas	Reproductive mode 1	Haddad et al 2013
<i>Melanophryniscus setiba</i>	Diurnal	Tetrapod	1,5	-----	Toxic	Cryptic	Forested areas	-----	Haddad et al 2013
<i>Melanophryniscus simplex</i>	Nocturnal	Tetrapod	2,8	River, stream or rivulet	Toxic	Terrestrial	Open areas	Reproductive mode 2	Haddad et al 2013

<i>Melanophryniscus spectabilis</i>	Diurnal	Tetrapod	3,1	River, stream or rivulet	Toxic	Terrestrial	Forested areas	Reproductive mode 2	Haddad et al 2013
<i>Melanophryniscus tumifrons</i>	-----	Tetrapod	3,1	Swamp or pond	Toxic	Terrestrial	Open or forested areas	Reproductive mode 1	Haddad et al 2013
<i>Melanophryniscus vilavelhensis</i>	Nocturnal	Tetrapod	1,7	Vegetation	Toxic	Phytotelmata	Open areas	Reproductive mode 6	Haddad et al 2013
<i>Myersiella microps</i>	Nocturnal	Tetrapod	3,1	Forest floor	Non toxic	Fossorial	Forested areas	Reproductive mode 23	Haddad et al 2013; present study
<i>Odontophrynus americanus</i>	Nocturnal	Tetrapod	4,9	Swamp or pond	Toxic	Terrestrial	Open areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Odontophrynus carvalhoi</i>	Nocturnal	Tetrapod	6,7	River, stream or rivulet	Toxic	Terrestrial	Open or forested areas	Reproductive mode 2	Haddad et al 2013
<i>Odontophrynus maisuma</i>	Nocturnal	Tetrapod	4,1	-----	Toxic	Terrestrial	Open areas	Reproductive mode 1	Haddad et al 2013
<i>Ololygon agilis</i>	Nocturnal and Diurnal	Tetrapod	1,8	Swamp or pond	Non toxic	Arboreal	Open or forested areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Ololygon albicans</i>	Nocturnal	Tetrapod	3	River, stream or rivulet	Non toxic	Arboreal	Forested areas	Reproductive mode 2	Haddad et al 2013; present study
<i>Ololygon angrensis</i>	Nocturnal and Diurnal	Tetrapod	3,4	Swamp or pond	Non toxic	Arboreal	Forested areas	Reproductive mode 1	Haddad et al 2013
<i>Ololygon arduous</i>	Nocturnal	Tetrapod	2,1	Bromeliad	Unpalatable	Arboreal	Forested	Reproductive	Haddad et al 2013; present

	and Diurnal				or bad odor		areas	mode 6	study
<i>Ololygon argyrorzata</i>	Nocturnal	Tetrapod	2,1	Swamp or pond	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Ololygon ariadne</i>	Nocturnal	Tetrapod	4,1	River, stream or rivulet	Non toxic	Arboreal	Forested areas	Reproductive mode 2	Haddad et al 2013
<i>Ololygon aromothyella</i>	Nocturnal	Tetrapod	2,6	Swamp or pond	Non toxic	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013
<i>Ololygon belloni</i>	Nocturnal	Tetrapod	3,9	Bromeliad	Non toxic	Arboreal	Open or forested areas	Reproductive mode 6	Haddad et al 2013
<i>Ololygon brieni</i>	Nocturnal	Tetrapod	3,5	River, stream or rivulet	Non toxic	Arboreal	Forested areas	Reproductive mode 2	Haddad et al 2013
<i>Ololygon carnevallii</i>	Nocturnal	Tetrapod	3,2	Swamp or pond	Non toxic	Arboreal	Forested areas	Reproductive mode 1	Haddad et al 2013
<i>Ololygon catharinae</i>	Nocturnal	Tetrapod	4,5	Swamp or pond	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Ololygon cosenzai</i>	Nocturnal	Tetrapod	2,3	Bromeliad	Non toxic	Arboreal	Open areas	Reproductive mode 6	Haddad et al 2013
<i>Ololygon flavoguttata</i>	Nocturnal	Tetrapod	4,5	River, stream or rivulet	Non toxic	Arboreal	Forested areas	Reproductive mode 2	Haddad et al 2013; present study
<i>Ololygon heyeri</i>	Nocturnal	Tetrapod	3,9	River, stream or rivulet	Non toxic	Arboreal	Forested areas	Reproductive mode 2	Haddad et al 2013
<i>Ololygon hiemalis</i>	Nocturnal	Tetrapod	3,5	Swamp or pond	Non toxic	Arboreal	Forested areas	Reproductive mode 1	Haddad et al 2013

<i>Ololygon humilis</i>	Nocturnal	Tetrapod	2,6	Swamp or pond	Non toxic	Arboreal	Forested areas	Reproductive mode 1	Haddad et al 2013
<i>Ololygon insperata</i>	Nocturnal	Tetrapod	2,3	Bromeliad	Non toxic	Arboreal	Forested areas	Reproductive mode 6	Haddad et al 2013
<i>Ololygon kautskyi</i>	-----	Tetrapod	2,8	River, stream or rivulet	Non toxic	Arboreal	Forested areas	Reproductive mode 2	Haddad et al 2013
<i>Ololygon littoralis</i>	Nocturnal	Tetrapod	3,8	Swamp or pond	Non toxic	Arboreal	Forested areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Ololygon littoreus</i>	Nocturnal	Tetrapod	2,2	Bromeliad	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 6	Haddad et al 2013
<i>Ololygon longilinea</i>	Nocturnal	Tetrapod	4,2	River, stream or rivulet	Non toxic	Arboreal	Forested areas	Reproductive mode 2	Haddad et al 2013
<i>Ololygon luizotavioi</i>	Nocturnal	Tetrapod	2,5	Swamp or pond; River or stream backwaters	Non toxic	Arboreal	Forested areas	Reproductive mode 1 and 2	Haddad et al 2013
<i>Ololygon machadoi</i>	Nocturnal	Tetrapod	2,4	River, stream or rivulet	Non toxic	Arboreal	Open or forested areas	Reproductive mode 2	Haddad et al 2013
<i>Ololygon obtriangulata</i>	Nocturnal	Tetrapod	3,5	River, stream or rivulet	Non toxic	Arboreal	Open or forested areas	Reproductive mode 2	Haddad et al 2013
<i>Ololygon peixotoi</i>	Nocturnal	Tetrapod	2,3	Bromeliad	Non toxic	Arboreal	Open or forested areas	Reproductive mode 6	Haddad et al 2013
<i>Ololygon</i>	Nocturnal	Tetrapod	2	Bromeliad	Unpalatable	Arboreal	Forested	Reproductive	Haddad et al 2013; present

<i>perpusilla</i>					or bad odor		areas	mode 6	study
<i>Ololygon ranki</i>	Nocturnal	Tetrapod	2,3	Swamp or pond	Non toxic	Arboreal	Forested areas	Reproductive mode 1	Haddad et al 2013
<i>Ololygon rizibilis</i>	Nocturnal and Diurnal	Tetrapod	3	Swamp or pond	Non toxic	Arboreal	Open or forested areas	Reproductive mode 11	Haddad et al 2013
<i>Ololygon strigilata</i>	Nocturnal	Tetrapod	3,8	Swamp or pond; River or stream backwaters	Non toxic	Arboreal	Forested areas	Reproductive mode 1 and 2	Haddad et al 2013
<i>Ololygon v-signata</i>	Nocturnal	Tetrapod	2,7	Bromeliad	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 6	Haddad et al 2013; present study
<i>Paratelmatoobius cardosoi</i>	Nocturnal	Tetrapod	1,7	Swamp or pond	-----	Cryptic	Forested areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Paratelmatoobius gaigeae</i>	Nocturnal	Tetrapod	1,8	Swamp or pond	-----	Cryptic	Forested areas	Reproductive mode 1	Haddad et al 2013
<i>Paratelmatoobius lutzii</i>	-----	Tetrapod	2,4	-----	-----	Cryptic	Open areas	-----	Haddad et al 2013
<i>Paratelmatoobius poecilogaster</i>	Nocturnal	Tetrapod	2,7	Swamp or pond	-----	Cryptic	Forested areas	Reproductive mode 18	Haddad et al 2013
<i>Paratelmatoobius yepiranga</i>	Nocturnal	Tetrapod	2,1	Swamp or pond	-----	Cryptic	Forested areas	Reproductive mode 1	Haddad et al 2013
<i>Phasmahyla cochranæ</i>	Nocturnal	Tetrapod	3,2	River, stream or rivulet	Toxic	Arboreal	Forested areas	Reproductive mode 25	Haddad et al 2013
<i>Phasmahyla exilis</i>	Nocturnal	Tetrapod	2,9	River, stream or	Toxic	Arboreal	Forested	Reproductive	Haddad et al 2013



				rivulet			areas	mode 25	
<i>Phasmahyla guttata</i>	Nocturnal	Tetrapod	3,8	River, stream or rivulet	Toxic	Arboreal	Forested areas	Reproductive mode 25	Haddad et al 2013; present study
<i>Phasmahyla jandaia</i>	Nocturnal	Tetrapod	3,6	River, stream or rivulet	Toxic	Arboreal	Forested areas	Reproductive mode 25	Haddad et al 2013
<i>Phasmahyla spectabilis</i>	Nocturnal	Tetrapod	4,6	River, stream or rivulet	Toxic	Arboreal	Forested areas	Reproductive mode 25	Haddad et al 2013
<i>Phasmahyla timbo</i>	Nocturnal	Tetrapod	3,5	River, stream or rivulet	Toxic	Arboreal	Forested areas	Reproductive mode 25	Haddad et al 2013
<i>Phrynomedusa marginata</i>	Nocturnal	Tetrapod	3,1	River, stream or rivulet	Toxic	Arboreal	Forested areas	Reproductive mode 18	Haddad et al 2013
<i>Phyllodytes acuminatus</i>	----	Tetrapod	2,4	Bromeliad	Non toxic	Arboreal	Forested areas	Reproductive mode 6	Haddad et al 2013
<i>Phyllodytes edelmoi</i>	----	Tetrapod	2,6	Bromeliad	Non toxic	Arboreal	Forested areas	Reproductive mode 6	Haddad et al 2013
<i>Phyllodytes gyrinaethes</i>	----	Tetrapod	2,6	Bromeliad	Non toxic	Arboreal	Forested areas	Reproductive mode 6	Haddad et al 2013
<i>Phyllodytes kautskyi</i>	Nocturnal and Diurnal	Tetrapod	4	Bromeliad	Non toxic	Arboreal	Forested areas	Reproductive mode 6	Haddad et al 2013
<i>Phyllodytes luteolus</i>	Nocturnal	Tetrapod	2,5	Bromeliad	Non toxic	Arboreal	Forested areas	Reproductive mode 6	Haddad et al 2013; present study
<i>Phyllodytes maculosus</i>	----	Tetrapod	4,9	----	Non toxic	Arboreal	Forested areas	Reproductive mode 6	Haddad et al 2013; Ferreira et al 2016; present study

<i>Phyllodytes melanomystax</i>	Nocturnal	Tetrapod	2,3	Bromeliad	Non toxic	Arboreal	Forested areas	Reproductive mode 6	Haddad et al 2013; present study
<i>Phyllodytes punctatus</i>	-----	Tetrapod	2,1	Bromeliad	Non toxic	Arboreal	Forested areas	Reproductive mode 6	Haddad et al 2013
<i>Phyllodytes tuberculosus</i>	Nocturnal	Tetrapod	2,4	Bromeliad	Non toxic	Arboreal	Open areas	Reproductive mode 6	Haddad et al 2013; present study
<i>Phyllodytes wuchereri</i>	Nocturnal	Tetrapod	2,7	Bromeliad	Non toxic	Arboreal	Forested areas	Reproductive mode 6	Haddad et al 2013
<i>Phyllomedusa bahiana</i>	Nocturnal	Tetrapod	7,5	Swamp or pond	Toxic	Arboreal	Forested areas	Reproductive mode 24	Haddad et al 2013; present study
<i>Phyllomedusa burmeisteri</i>	Nocturnal	Tetrapod	6	Swamp or pond	Toxic	Arboreal	Open or forested areas	Reproductive mode 24	Haddad et al 2013; present study
<i>Phyllomedusa distincta</i>	Nocturnal	Tetrapod	6	Swamp or pond	Toxic	Arboreal	Forested areas	Reproductive mode 24	Haddad et al 2013; present study
<i>Phyllomedusa iheringii</i>	Nocturnal	Tetrapod	6,8	Swamp or pond	Toxic	Arboreal	Open areas	Reproductive mode 24	Haddad et al 2013
<i>Phyllomedusa nordestina</i>	Nocturnal	Tetrapod	3,7	Swamp or pond	Toxic	Arboreal	Forested areas	Reproductive mode 24	Haddad et al 2013; present study
<i>Phyllomedusa rohdei</i>	Nocturnal	Tetrapod	4,6	Swamp or pond	Toxic	Arboreal	Forested areas	Reproductive mode 24	Haddad et al 2013; present study
<i>Phyllomedusa tetraploidea</i>	Nocturnal	Tetrapod	6,6	Swamp or pond	Toxic	Arboreal	Open or forested areas	Reproductive mode 24	Haddad et al 2013; present study

<i>Physalaemus aguirrei</i>	Nocturnal	Tetrapod	2,7	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 11	Haddad et al 2013
<i>Physalaemus atlanticus</i>	Nocturnal	Tetrapod	2,1	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 11 and 28	Haddad et al 2013
<i>Physalaemus barrioi</i>	Nocturnal	Tetrapod	2,9	Swamp or pond	Non toxic	Terrestrial	Open areas	Reproductive mode 11	Haddad et al 2013
<i>Physalaemus bokermanni</i>	Nocturnal	Tetrapod	1,7	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 11 and 28	Haddad et al 2013
<i>Physalaemus camacan</i>	Nocturnal	Tetrapod	2,4	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 11	Haddad et al 2013; present study
<i>Physalaemus crombiei</i>	Nocturnal	Tetrapod	2,3	Swamp or pond	Non toxic	Terrestrial	Forested areas	Reproductive mode 11 and 28	Haddad et al 2013
<i>Physalaemus cuvieri</i>	Nocturnal	Tetrapod	3,1	Swamp or pond	Non toxic	Terrestrial	Open areas	Reproductive mode 11	Haddad et al 2013; present study
<i>Physalaemus erikae</i>	Nocturnal	Tetrapod	2,6	Swamp or pond	Non toxic	Terrestrial	Open or forested areas	Reproductive mode 11	Haddad et al 2013
<i>Physalaemus feioi</i>	Nocturnal	Tetrapod	2,8	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 11	Haddad et al 2013
<i>Physalaemus jordanensis</i>	Nocturnal	Tetrapod	2,8	Swamp or pond	Non toxic	Cryptic	Open areas	Reproductive mode 11	Haddad et al 2013
<i>Physalaemus</i>	Nocturnal	Tetrapod	2,9	Swamp or pond	Non toxic	Cryptic	Open areas	Reproductive	Haddad et al 2013

<i>kroyeri</i>								mode 11	
<i>Physalaemus lateristriga</i>	Nocturnal	Tetrapod	3,5	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 11	Haddad et al 2013
<i>Physalaemus lisei</i>	Nocturnal	Tetrapod	3	Swamp or pond	Non toxic	Cryptic	Open or forested areas	Reproductive mode 11	Haddad et al 2013
<i>Physalaemus maculiventris</i>	Nocturnal	Tetrapod	2,3	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 11	Haddad et al 2013
<i>Physalaemus maximus</i>	Nocturnal	Tetrapod	4,8	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 11	Haddad et al 2013
<i>Physalaemus moreirae</i>	Nocturnal	Tetrapod	2,6	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 11	Haddad et al 2013
<i>Physalaemus nanus</i>	Nocturnal	Tetrapod	2,1	Swamp or pond	Non toxic	Cryptic	Open or forested areas	Reproductive mode 11 and 28	Haddad et al 2013
<i>Physalaemus obtectus</i>	Nocturnal	Tetrapod	2,8	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 11	Haddad et al 2013
<i>Physalaemus olfersii</i>	Nocturnal	Tetrapod	3,8	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 11	Haddad et al 2013
<i>Physalaemus signifer</i>	Nocturnal	Tetrapod	2,2	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 11 and 28	Haddad et al 2013; present study
<i>Physalaemus soaresi</i>	Nocturnal	Tetrapod	2,1	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 11	Haddad et al 2013
<i>Physalaemus</i>	Nocturnal	Tetrapod	2,2	Swamp or pond	Non toxic	Cryptic	Forested	Reproductive	Haddad et al 2013

<i>spiniger</i>							areas	mode 11, 14 and 28	
<i>Pipa carvalhoi</i>	Nocturnal and Diurnal	Tetrapod	5,5	Swamp or pond	Non toxic	Aquatic	Open areas	Reproductive mode 15	Haddad et al 2013; present study
<i>Pristimantis paulodutraii</i>	Nocturnal	Tetrapod	3,6	Forest floor; Low vegetation	Unpalatable or bad odor	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013; present study
<i>Pristimantis ramagii</i>	Nocturnal	Tetrapod	2,4	Forest floor; Low vegetation	Unpalatable or bad odor	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013; present study
<i>Pristimantis vinhai</i>	-----	Tetrapod	2,5	Forest floor	Unpalatable or bad odor	Cryptic	Forested areas	Reproductive mode 23	Haddad et al 2013; present study
<i>Proceratophrys appendiculata</i>	Nocturnal	Tetrapod	6,3	Swamp or pond; River or stream backwaters	Non toxic	Cryptic	Forested areas	-----	Haddad et al 2013; present study
<i>Proceratophrys avelinoi</i>	Nocturnal	Tetrapod	2,6	River, stream or rivulet	Non toxic	Cryptic	Open or forested areas	Reproductive mode 2	Haddad et al 2013; present study
<i>Proceratophrys bigibbosa</i>	Nocturnal	Tetrapod	5,3	River, stream or rivulet	Non toxic	Cryptic	Forested areas	Reproductive mode 2	Haddad et al 2013
<i>Proceratophrys boiei</i>	Nocturnal	Tetrapod	5,7	Swamp or pond; River or stream backwaters	Non toxic	Cryptic	Forested areas	Reproductive mode 1 and 2	Haddad et al 2013; present study
<i>Proceratophrys brauni</i>	Nocturnal and Diurnal	Tetrapod	3,9	River, stream or rivulet	Non toxic	Cryptic	Forested areas	Reproductive mode 2	Haddad et al 2013
<i>Proceratophrys</i>	Nocturnal	Tetrapod	6,4	River, stream or	Non toxic	Cryptic	Forested	Reproductive	Haddad et al 2013

<i>cristiceps</i>				rivulet			areas	mode 2	
<i>Proceratophrys cururu</i>	Nocturnal	Tetrapod	5,4	River or stream backwaters	Non toxic	Cryptic	Forested areas	Reproductive mode 2	Haddad et al 2013
<i>Proceratophrys laticeps</i>	Nocturnal	Tetrapod	6,7	Swamp or pond; River or stream backwaters	Non toxic	Cryptic	Forested areas	Reproductive mode 1 and 2	Haddad et al 2013; present study
<i>Proceratophrys melanopogon</i>	Nocturnal	Tetrapod	5,1	Swamp or pond; River or stream backwaters	Non toxic	Cryptic	Forested areas	Reproductive mode 1 and 2	Haddad et al 2013
<i>Proceratophrys minuta</i>	Nocturnal	Tetrapod	3	River, stream or rivulet	Non toxic	Cryptic	Forested areas	Reproductive mode 2	Haddad et al 2013
<i>Proceratophrys paviotii</i>	Nocturnal	Tetrapod	4,7	River, stream or rivulet	Non toxic	Cryptic	Forested areas	Reproductive mode 2	Haddad et al 2013
<i>Proceratophrys renalis</i>	Nocturnal	Tetrapod	5,5	River, stream or rivulet	Non toxic	Cryptic	Forested areas	Reproductive mode 2	Haddad et al 2013; present study
<i>Proceratophrys sanctaritae</i>	Nocturnal	Tetrapod	4,1	River, stream or rivulet	Non toxic	Cryptic	Forested areas	Reproductive mode 2	Haddad et al 2013
<i>Proceratophrys schirchi</i>	Nocturnal	Tetrapod	5	River, stream or rivulet	Non toxic	Cryptic	Forested areas	Reproductive mode 2	Haddad et al 2013
<i>Proceratophrys subguttata</i>	Nocturnal	Tetrapod	4,8	River, stream or rivulet	Non toxic	Cryptic	Forested areas	Reproductive mode 2	Haddad et al 2013
<i>Pseudis bolbodactyla</i>	Nocturnal	Tetrapod	4,5	Swamp or pond	Non toxic	Aquatic	Open areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Pseudis cardosoi</i>	Nocturnal	Tetrapod	4,5	Swamp or pond	Non toxic	Aquatic	Open areas	Reproductive mode 1	Haddad et al 2013

<i>Pseudis fusca</i>	Nocturnal	Tetrapod	4,1	Swamp or pond	Non toxic	Aquatic	Open areas	Reproductive mode 1	Haddad et al 2013
<i>Pseudis minuta</i>	Nocturnal	Tetrapod	3,7	Swamp or pond	Non toxic	Aquatic	Open areas	Reproductive mode 1	Haddad et al 2013
<i>Pseudis paradoxa</i>	Nocturnal	Tetrapod	5,3	Swamp or pond	Non toxic	Aquatic	Open areas	Reproductive mode 1	Haddad et al 2013
<i>Pseudopaludicola falcipes</i>	Nocturnal and Diurnal	Tetrapod	1,8	Swamp or pond	Non toxic	Cryptic	Open areas	Reproductive mode 1	Haddad et al 2013
<i>Rhinella abei</i>	Nocturnal	Tetrapod	9,3	Swamp or pond; River or stream backwaters	Toxic	Terrestrial	Open or forested areas	Reproductive mode 1 and 2	Haddad et al 2013; present study
<i>Rhinella achavali</i>	Nocturnal	Tetrapod	12	Swamp or pond	Toxic	Terrestrial	Open areas	Reproductive mode 1	Haddad et al 2013
<i>Rhinella crucifer</i>	Nocturnal	Tetrapod	10,1	Swamp or pond; River or stream backwaters	Toxic	Terrestrial	Open or forested areas	Reproductive mode 1 and 2	Haddad et al 2013; present study
<i>Rhinella dorbignyi</i>	Nocturnal	Tetrapod	5,6	Swamp or pond	Toxic	Terrestrial	Open areas	Reproductive mode 1	Haddad et al 2013
<i>Rhinella fernandezae</i>	Nocturnal	Tetrapod	5,7	Swamp or pond	Toxic	Terrestrial	Open areas	Reproductive mode 1	Haddad et al 2013
<i>Rhinella granulosa</i>	Nocturnal	Tetrapod	5,3	Swamp or pond	Toxic	Terrestrial	Open or forested areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Rhinella henseli</i>	Nocturnal	Tetrapod	7	Swamp or pond	Toxic	Terrestrial	Open or	Reproductive	Haddad et al 2013; present

							forested areas	mode 1 and 2	study
<i>Rhinella hoogmoedi</i>	Nocturnal	Tetrapod	5,6	Swamp or pond	Toxic	Terrestrial	Forested areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Rhinella icterica</i>	Nocturnal	Tetrapod	13,7	Swamp or pond; River or stream backwaters	Toxic	Terrestrial	Open or forested areas	Reproductive mode 1 and 2	Haddad et al 2013; present study
<i>Rhinella jimi</i>	Nocturnal	Tetrapod	14,5	Swamp or pond	Toxic	Terrestrial	Open or forested areas	Reproductive mode 1	Haddad et al 2013
<i>Rhinella ornata</i>	Nocturnal	Tetrapod	7,9	Swamp or pond; River or stream backwaters	Toxic	Terrestrial	Open or forested areas	Reproductive mode 1 and 2	Haddad et al 2013; present study
<i>Rhinella pygmaea</i>	Nocturnal	Tetrapod	4,1	Swamp or pond	Toxic	Terrestrial	Open areas	Reproductive mode 1	Haddad et al 2013
<i>Rhinella schneideri</i>	Nocturnal	Tetrapod	13	Swamp or pond; River or stream backwaters	Toxic	Terrestrial	Open areas	Reproductive mode 1 and 2	Haddad et al 2013; present study
<i>Scinax alter</i>	Nocturnal	Tetrapod	2,8	Swamp or pond	Unpalatable or bad odor	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Scinax auratus</i>	-----	Tetrapod	2,6	Swamp or pond	Non toxic	Arboreal	Open or forested areas	Reproductive mode 1	Haddad et al 2013
<i>Scinax caldarum</i>	Nocturnal	Tetrapod	2,9	Swamp or pond	Non toxic	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013
<i>Scinax cretatus</i>	Nocturnal	Tetrapod	3,2	Swamp or pond	Non toxic	Arboreal	Open areas	Reproductive	Haddad et al 2013



										mode 1
<i>Scinax crospedospilus</i>	Nocturnal	Tetrapod	3,1	Swamp or pond	Unpalatable or bad odor	Arboreal	Forested areas	Reproductive mode 1	Haddad et al 2013; present study	
<i>Scinax cuspidatus</i>	Nocturnal	Tetrapod	3,1	Swamp or pond	Non toxic	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013	
<i>Scinax duartei</i>	Nocturnal	Tetrapod	3	Swamp or pond	Non toxic	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013	
<i>Scinax eurydice</i>	Nocturnal	Tetrapod	5,3	Swamp or pond	Unpalatable or bad odor	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013; present study	
<i>Scinax fuscomarginatus</i>	Nocturnal	Tetrapod	2,2	Swamp or pond	Unpalatable or bad odor	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013; present study	
<i>Scinax fuscovarios</i>	Nocturnal	Tetrapod	4,5	Swamp or pond	Unpalatable or bad odor	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013; present study	
<i>Scinax granulatus</i>	Nocturnal	Tetrapod	4,1	Swamp or pond	Unpalatable or bad odor	Arboreal; Terrestrial	Open areas	Reproductive mode 1	Haddad et al 2013; present study	
<i>Scinax hayii</i>	Nocturnal	Tetrapod	5,1	Swamp or pond	Unpalatable or bad odor	Arboreal; Terrestrial	Forested areas	Reproductive mode 1	Haddad et al 2013; present study	
<i>Scinax imbegue</i>	Nocturnal	Tetrapod	3,2	Swamp or pond	Non toxic	Arboreal; Terrestrial	Open areas	Reproductive mode 1	Haddad et al 2013	
<i>Scinax juncae</i>	Nocturnal	Tetrapod	2,4	Swamp or pond	Non toxic	Arboreal	Forested areas	Reproductive mode 1	Haddad et al 2013	
<i>Scinax pachycrus</i>	Nocturnal	Tetrapod	3,2	Swamp or pond	Non toxic	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013	

<i>Scinax perereca</i>	Nocturnal	Tetrapod	4,3	Swamp or pond	Unpalatable or bad odor	Arboreal	Open or forested areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Scinax similis</i>	Nocturnal	Tetrapod	3,8	Swamp or pond	Unpalatable or bad odor	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Scinax squalitostris</i>	Nocturnal	Tetrapod	2,7	Swamp or pond	Non toxic	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013
<i>Scinax trapicheiroi</i>	Nocturnal	Tetrapod	2,9	River, stream or rivulet	Non toxic	Arboreal	Forested areas	Reproductive mode 1 and 2	Haddad et al 2013
<i>Scinax tymbamirim</i>	Nocturnal	Tetrapod	2,6	Swamp or pond	Non toxic	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013
<i>Scinax uruguayus</i>	Nocturnal	Tetrapod	2,8	Swamp or pond	Non toxic	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013
<i>Scinax x-signatus</i>	Nocturnal	Tetrapod	4,1	Swamp or pond	Unpalatable or bad odor	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Scythroehrys sawayae</i>	Nocturnal	Tetrapod	2,1	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 1	Haddad et al 2013
<i>Siphonops annuatus</i>	-----	Apod	45,4	Do not Call	-----	Fossorial	Open or forested areas	Reproductive mode 38	Haddad et al 2013; present study
<i>Siphonops hardyi</i>	-----	Apod	17,8	Do not Call	-----	Fossorial	Open or forested areas	Reproductive mode 38	Haddad et al 2013
<i>Siphonops paulensis</i>	-----	Apod	30	Do not Call	-----	Fossorial	Open or forested	Reproductive mode 38	Haddad et al 2013

								areas		
<i>Sphaenorhynchus botocudo</i>	Nocturnal	Tetrapod	2,7	Swamp or pond	Non toxic	Arboreal; Aquatic	Open areas	Reproductive mode 1	Haddad et al 2013	
<i>Sphaenorhynchus caramaschii</i>	Nocturnal	Tetrapod	2,7	Swamp or pond	Non toxic	Arboreal; Aquatic	Open or forested areas	Reproductive mode 1	Haddad et al 2013	
<i>Sphaenorhynchus mirim</i>	Nocturnal	Tetrapod	1,7	Swamp or pond	Non toxic	Arboreal; Aquatic	Open areas	Reproductive mode 1	Haddad et al 2013	
<i>Sphaenorhynchus orophilus</i>	Nocturnal	Tetrapod	3,2	Swamp or pond	Non toxic	Arboreal	Open or forested areas	Reproductive mode 1	Haddad et al 2013	
<i>Sphaenorhynchus palustris</i>	Nocturnal	Tetrapod	3,1	Swamp or pond	Non toxic	Arboreal	Open or forested areas	Reproductive mode 1	Haddad et al 2013	
<i>Sphaenorhynchus pauloalvini</i>	Nocturnal	Tetrapod	1,9	Swamp or pond	Non toxic	Arboreal	Forested areas	Reproductive mode 24	Haddad et al 2013	
<i>Sphaenorhynchus planicola</i>	Nocturnal	Tetrapod	2	Swamp or pond	Non toxic	Arboreal	Open or forested areas	Reproductive mode 1	Haddad et al 2013	
<i>Sphaenorhynchus prasinus</i>	Nocturnal	Tetrapod	2,8	Swamp or pond	Non toxic	Arboreal	Open or forested areas	Reproductive mode 1	Haddad et al 2013	
<i>Sphaenorhynchus surdus</i>	Nocturnal	Tetrapod	2,8	Swamp or pond	Non toxic	Arboreal; Aquatic	Open or forested areas	Reproductive mode 1	Haddad et al 2013	
<i>Stereocyclops</i>	Nocturnal	Tetrapod	6,2	Swamp or pond	Non toxic	Cryptic	Forested	Reproductive	Haddad et al 2013	

<i>incrassatus</i>							areas	mode 1	
<i>Stereocyclops parkeri</i>	Nocturnal	Tetrapod	6	Swamp or pond	Non toxic	Cryptic	Forested areas	Reproductive mode 1	Haddad et al 2013
<i>Thoropa lutzi</i>	Nocturnal	Tetrapod	2,6	Rock wall	Unpalatable or bad odor	Terrestrial	Forested areas	Reproductive mode 19	Haddad et al 2013
<i>Thoropa miliaris</i>	Nocturnal	Tetrapod	6,3	Rock wall	Non toxic	Terrestrial	Forested areas	Reproductive mode 19	Haddad et al 2013; present study
<i>Thoropa saxatilis</i>	Nocturnal	Tetrapod	5,9	Rock wall	Non toxic	Terrestrial	Forested areas	Reproductive mode 19	Haddad et al 2013
<i>Thoropa taophora</i>	Nocturnal	Tetrapod	6,3	Rock wall	Non toxic	Terrestrial	Forested areas	Reproductive mode 19	Haddad et al 2013
<i>Trachycephalus atlas</i>	Nocturnal	Tetrapod	10,7	Swamp or pond	Toxic	Arboreal	Open areas	Reproductive mode 1	Haddad et al 2013
<i>Trachycephalus dibernardoii</i>	Nocturnal	Tetrapod	7,8	Swamp or pond	Toxic	Arboreal	Forested areas	Reproductive mode 1	Haddad et al 2013
<i>Trachycephalus imitatrix</i>	Nocturnal	Tetrapod	7,1	Swamp or pond	Toxic	Arboreal	Forested areas	Reproductive mode 1	Haddad et al 2013
<i>Trachycephalus lepidus</i>	Nocturnal	Tetrapod	5,9	Swamp or pond	Toxic	Arboreal	Forested areas	Reproductive mode 1	Haddad et al 2013
<i>Trachycephalus mesophaeus</i>	Nocturnal and Diurnal	Tetrapod	6,9	Swamp or pond	Toxic	Arboreal	Forested areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Trachycephalus nigromaculatus</i>	Nocturnal	Tetrapod	9,2	Swamp or pond; River or stream backwaters	Toxic	Arboreal	Forested areas	Reproductive mode 1 and 2	Haddad et al 2013

<i>Trachycephalus typhonius</i>	Nocturnal	Tetrapod	9,2	Swamp or pond	Toxic	Arboreal	Open or forested areas	Reproductive mode 1	Haddad et al 2013; present study
<i>Vitreorana eurygnatha</i>	Nocturnal	Tetrapod	2,4	River, stream or rivulet	Non toxic	Arboreal	Forested areas	Reproductive mode 25	Haddad et al 2013; present study
<i>Vitreorana uranoscopa</i>	Nocturnal	Tetrapod	2,7	River or stream backwaters	Non toxic	Arboreal	Forested areas	Reproductive mode 25	Haddad et al 2013; present study
<i>Xenohyla truncata</i>	Nocturnal	Tetrapod	5	Swamp or pond	-----	Arboreal	Forested areas	Reproductive mode 1	Haddad et al 2013
<i>Zachaenus carvalhoi</i>	Nocturnal	Tetrapod	2,8	Forest floor	Non toxic	Cryptic	Forested areas	Reproductive mode 21	Haddad et al 2013
<i>Zachaenus parvulus</i>	Nocturnal	Tetrapod	2,9	Forest floor	Non toxic	Cryptic	Forested areas	Reproductive mode 21	Haddad et al 2013; present study

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**Table S3.** GenBank accession numbers for 208 sampled amphibian taxa. (\*) indicates outgroup.

<b>Species</b>	<b>12S</b>	<b>16S</b>	<b>Cytb</b>	<b>CXCR4</b>	<b>H3A</b>	<b>NCX1</b>	<b>POMC</b>	<b>RAG1</b>	<b>RHOD</b>	<b>SIA</b>	<b>SLC8A3</b>	<b>TYR</b>
<i>Adelophryne mucronatus</i>	--	JX298291.1	JX298385.1	--	--	--	JX298112.1	JX298158.1	--	--	--	JX298213.1
<i>Adelophryne pachydactyla</i>	JX298259.1	JX298294.1	JX298387.1	--	--	--	JX298115.1	JX298161.1	--	--	--	JX298216.1
<i>Adenomera araucaria</i>	KC470091.1	KC477241.1	KC603969.1	--	--	--	KC604065.1	KC604019.1	KC604099.1	--	--	KC604089.1
<i>Adenomera bokermani</i>	KC470107.1	KC477243.1	KF548072.1	--	--	--	--	--	KF613580.1	--	--	--
<i>Adenomera engelsi</i>	KC603940.1	--	KC603970.1	--	--	--	KC604066.1	KC604020.1	KC604100.1	--	--	KC604090.1
<i>Adenomera thomei</i>	KC603945.1	KC603946.1	KC603971.1	--	--	--	KC604067.1	KC604021.1	KC604101.1	--	--	KC604091.1
<i>Allobates alagoanus</i>	DQ502126.1	--	DQ502557.1	--	DQ502342.1	--	--	--	DQ503232.1	DQ503093.1	--	--
<i>Aparasphenodon brunoi</i>	AY843567.1	--	AY843789.1	--	--	--	--	AY844364.1	AY844541.1	AY844769.1	--	AY844023.1
<i>Aplastodiscus albofrenatus</i>	AY819422.1	AY819539.1	--	--	--	--	--	--	--	--	--	--
<i>Aplastodiscus albosignatus</i>	AY843596.1	--	AY843817.1	--	--	--	--	AY844385.1	AY844570.1	AY844796.1	--	AY844042.1
<i>Aplastodiscus arildae</i>	AY843604.1	--	AY843825.1	--	--	--	--	AY844392.1	AY844578.1	AY844803.1	--	AY844049.1

<i>Aplastodiscus callipygius</i>	AY843614.1	AY843614.1	AY843840.1	--	--	--	--	AY844402.1	AY844592.1	AY844813.1	--	AY844058.1
<i>Aplastodiscus cavicola</i>	AY843617.1	AY843617.1	AY843843.1	--	--	--	--	AY844405.1	AY844594.1	AY844814.1	--	--
<i>Aplastodiscus cochranæ</i>	AY843568.1	AY843568.1	AY843790.1	--	--	--	--	AY844365.1	AY844542.1	AY844770.1	--	AY844024.1
<i>Aplastodiscus ehrhardti</i>	--	--	--	--	--	--	--	AY844456.1	--	--	--	--
<i>Aplastodiscus eugenioi</i>	AY843669.1	AY843669.1	AY843913.1	KF751465.1	--	--	--	AY844456.1	AY844660.1	AY844875.1	--	--
<i>Aplastodiscus leucopygius</i>	AY843638.1	AY843638.1	AY843873.1	KF751466.1	--	--	--	AY844425.1	AY844622.1	AY844840.1	--	AY844084.1
<i>Aplastodiscus perviridis</i>	AY843569.1	AY843569.1	AY843791.1	KF751467.1	DQ284044.1	--	--	AY844366.1	AY844543.1	AY844771.1	--	AY844025.1
<i>Aplastodiscus weygoldti</i>	AY843685.1	AY843685.1	AY843931.1	--	--	--	--	AY844467.1	AY844678.1	AY844887.1	--	--
<i>Bokermanohyla astartea</i>	AY549322.1	AY549322.1	AY843827.1	--	--	--	AY819113.1	--	AY844580.1	--	--	--
<i>Bokermanohyla circumdata</i>	AY549328.1	--	AY843847.1	KF751468.1	--	--	--	AY844409.1	AY844598.1	AY844817.1	--	AY844064.1
<i>Bokermanohyla hylax</i>	AY549338.1	AY549338.1	AY843865.1	--	--	--	--	AY844419.1	AY844614.1	AY844832.1	--	AY844077.1
<i>Bokermanohyla martinsi</i>	AY843641.1	AY843641.1	AY843878.1	--	--	--	--	--	AY844626.1	AY844844.1	--	AY844086.1
<i>Brachycephalus</i>	HQ435676.1	HQ435690.1	HQ435703.1	--	--	--	--	HQ435718.1	--	--	--	HQ435732.1

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<i>Brachycephalus brunneus</i>	HQ435677.1	HQ435691.1	HQ435704.1	--	--	--	--	HQ435719.1	--	--	--	HQ435733.1
<i>Brachycephalus ephipium</i>	HM216368.1	HM216369.1	HM216367.1	GQ345180.1	GQ345212.1	GQ345228.1	GQ345256.1	HM216370.1	DQ283808.1	DQ282673.1	GQ345326.1	HQ435735.1
<i>Brachycephalus ferruginus</i>	HQ435681.1	HQ435695.1	HQ435708.1	--	--	--	--	HQ435723.1	--	--	--	HQ435737.1
<i>Brachycephalus garbeanus</i>	HQ435680.1	HQ435694.1	HQ435707.1	--	--	--	--	HQ435722.1	--	--	--	HQ435736.1
<i>Brachycephalus hermogenesi</i>	HQ435682.1	--	HQ435709.1	--	--	--	--	HQ435724.1	--	--	--	HQ435738.1
<i>Brachycephalus izecksohni</i>	HQ435683.1	HQ435696.1	HQ435710.1	--	--	--	--	HQ435725.1	--	--	--	HQ435739.1
<i>Brachycephalus pernix</i>	HQ435685.1	HQ435698.1	HQ435712.1	--	--	--	--	HQ435727.1	--	--	--	HQ435741.1
<i>Brachycephalus pitanga</i>	HQ435686.1	HQ435699.1	HQ435713.1	--	--	--	--	HQ435728.1	--	--	--	HQ435742.1
<i>Brachycephalus pombali</i>	HQ435687.1	HQ435700.1	HQ435714.1	--	--	--	--	HQ435729.1	--	--	--	HQ435743.1
<i>Brachycephalus toby</i>	HQ435688.1	HQ435701.1	HQ435715.1	--	--	--	--	HQ435730.1	--	--	--	HQ435744.1
<i>Brachycephalus vertebralis</i>	HQ435689.1	HQ435702.1	HQ435716.1	--	--	--	--	HQ435731.1	--	--	--	HQ435745.1
<i>Brahycephalus didactylus</i>	JX267389.1	JX267467.1	HQ435705.1	--	--	--	--	JX267544.1	--	--	--	JX267681.1



<i>Chiasmocleis capixaba</i>	--	KC180044.1	--	--	--	--	--	--	--	--	--	KC180235.1
<i>Chiasmocleis leucosticta</i>	--	KC180039.1	--	--	--	--	--	--	--	--	--	KC180243.1
<i>Chiasmocleis schubarti</i>	--	KC180071.1	--	--	--	--	--	--	--	--	--	KC180247.1
<i>Chthonerpeton indistinctum</i>	--	EF107202.1	--	--	--	EF107266.1	--	EF107325.1	--	--	EF107428.1	--
<i>Crossodactylodes bokermanni</i>	KF534640.1	KF534650.1	KF534668.1	--	--	--	--	--	KF534677.1	--	--	KF534683.1
<i>Crossodactylodes izecksohni</i>	KF534633.1	KF534646.1	KF534664.1	--	--	--	--	--	KF534673.1	--	--	KF534680.2
<i>Crossodactylus caramaschii</i>	AY143346.1	AY263235.1	KC603961.1	--	--	--	KC604072.1	KC604005.1	--	--	--	--
<i>Cycloramphus acangatan</i>	HQ634162.1	FJ685683.1	FJ685663.1	--	--	--	--	HQ634170.1	KF214198.1	--	--	--
<i>Cycloramphus bandeirensis</i>	HQ634161.1	HQ634166.1	--	--	--	--	--	HQ634169.1	--	--	--	--
<i>Cycloramphus eleutherodactylus</i>	HQ634160.1	HQ634165.1	--	--	--	--	--	HQ634168.1	--	--	--	--
<i>Cycloramphus organensis</i>	HQ634159.1	HQ634164.1	--	--	--	--	--	HQ634167.1	--	--	--	--
<i>Dendrophryniscus berthaltzae</i>	JN867524.1	JN867551.1	--	--	--	--	--	JN867497.1	--	--	--	--
<i>Dendrophryniscus</i>	AF375490.1	AF375515.1	--	--	--	--	--	JN867499.1	--	--	--	--

*brevipollicatus**Dendrophryniscus  
carvalhoi*

JN867539.1	JN867564.1	--	--	--	--	--	--	JN867512.1	--	--	--	--
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*Dendrophryniscus  
krause*

JN867541.1	JN867569.1	--	--	--	--	--	--	JN867515.1	--	--	--	--
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*Dendrophryniscus  
leucomystax*

JN867525.1	JN867552.1	--	--	--	--	--	--	JN867498.1	--	--	--	--
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*Dendrophryniscus  
oreites*

JN867531.1	JN867558.1	--	--	--	--	--	--	JN867504.1	--	--	--	--
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*Dendrophryniscus  
proboscideus*

JN867540.1	JN867567.1	--	--	--	--	--	--	JN867513.1	--	--	--	--
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*Dendropsophus  
anceps*

AY843597.1	AY843597.1	AY843818.1	--	--	--	--	--	AY844386.1	AY844571.1	AY844797.1	--	AY844043.1
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*Dendropsophus  
berthalutzae*

AY843607.1	AY843607.1	AY843831.1	--	--	--	--	--	AY844397.1	AY844584.1	AY844807.1	--	AY844052.1
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*Dendropsophus  
bipunctatus*

AY843608.1	AY843608.1	AY843832.1	--	--	--	--	--	--	AY844585.1	AY844808.1	--	AY844053.1
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*Dendropsophus  
branneri*

--	--	AF549336.1	--	--	--	--	--	--	--	--	--	--
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*Dendropsophus  
elegans*

DQ380355.1	AF308102.1	AF308124.1	--	--	--	--	--	--	--	--	--	--
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*Dendropsophus  
giesleri*

AY843629.1	AY843629.1	AY843860.1	--	--	--	--	--	AY844417.1	--	AY844827.1	--	AY844075.1
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*Dendropsophus  
minutus*

AY549345.1	AY549345.1	AY843883.1	--	DQ284046.1	--	--	--	AY844432.1	DQ283758.1	--	--	AY844089.1
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<i>Dendropsophus nanus</i>	AY549346.1	AY549346.1	AY843888.1	GQ365985.1	DQ284051.1	--	AY819123.1	AY844437.1	AY844634.1	AY844852.1	--	EF376132.1
<i>Dendropsophus samborni</i>	AY843663.1	AY843663.1	AY843906.1	--	--	--	--	AY844450.1	AY844653.1	AY844868.1	--	AY844106.1
<i>Dendropsophus seniculus</i>	AY843666.1	AY843666.1	AY843910.1	--	--	--	--	AY844454.1	AY844657.1	AY844872.1	--	AY844109.1
<i>Elachistocleis bicolor</i>	JF836935.1	KC180005.1	--	--	--	--	--	--	JF837037.1	--	--	KC180318.1
<i>Eleutherodactylus bilineata</i>	JX267323.1	JX267323.1	--	--	--	--	--	JX267556.1	--	--	--	JX267691.1
<i>Euparkerella brasiliensis</i>	JX267390.1	JX298316.1	JX298402.1	--	--	--	JX298137.1	KF625104.1	--	--	--	KF625126.1
<i>Euparkerella cochranae</i>	--	--	--	--	--	--	--	KF625094.1	--	--	--	KF625116.1
<i>Euparkerella robusta</i>	--	--	--	--	--	--	--	KF625089.1	--	--	--	KF625111.1
<i>Euparkerella tridactyla</i>	--	--	--	--	--	--	--	KF625088.1	--	--	--	KF625110.1
<i>Fritziana fissilis</i>	--	JN157630.1	--	--	--	--	JN157628.1	--	--	--	--	--
<i>Fritziana goeldii</i>	--	JN157631.1	--	--	--	--	--	--	--	--	--	--
<i>Fritziana ohausi</i>	--	JN157635.1	--	--	--	--	JN157629.1	KC844991.1	--	--	--	--
<i>Gastrotheca albolineata</i>	--	KC844924.1	--	--	--	--	KC844971.1	KC844992.1	--	--	--	--
<i>Gastrotheca</i>	--	KC844927.1	--	--	--	--	KC844975.1	KC844995.1	--	--	--	--

*ernestoi*

<i>Gastrotheca fulvorufa</i>	--	KC844929.1	--	--	--	--	KC844977.1	KC844997.1	--	--	--	--
<i>Gastrotheca microdiscus</i>	--	KC844933.1	--	--	--	--	KC844981.1	KC845000.1	--	--	--	--
<i>Haddadus binotatus</i>	KF740846.1	KF740846.1	JX298405.1	GQ345183.1	DQ284142.1	GQ345231.1	GQ345259.1	JX267548.1	DQ283807.1	GQ345309.1	GQ345329.1	JX267685.1
<i>Holoaden bradei</i>	--	--	JX298403.1	--	--	--	JX298138.1	EF493449.1	--	--	--	EU186779.1
<i>Holoaden luederwaldti</i>	EU186728.1	EU186710.1	--	--	--	--	--	JX267549.1	--	--	--	EU186768.1
<i>Hylodes nasus</i>	--	--	--	GQ345194.1	--	GQ345242.1	GQ345272.1	GQ345289.1	--	--	GQ345340.1	--
<i>Hylodes phyllodes</i>	DQ283096.1	DQ283096.1	DQ502587.1	--	DQ284146.1	--	--	KC604006.1	DQ283812.1	DQ282674.1	--	DQ282923.1
<i>Hylomantis aspera</i>	GQ366222.1	--	--	GQ365978.1	--	--	--	--	GQ366098.1	--	--	--
<i>Hylomantis granulosa</i>	AY843687.1	GQ366292.1	AY843933.1	GQ365979.1	--	--	GQ366032.1	AY844469.1	AY844680.1	--	--	AY844127.1
<i>Hyophryne histrio</i>	--	KC180064.1	--	--	--	--	--	--	--	--	--	--
<i>Hypsiboas albomarginatus</i>	AY549316.1	AY549316.1	AY843815.1	--	--	--	--	AY844384.1	AY844568.1	AY844794.1	--	--
<i>Hypsiboas albopunctatus</i>	AY549317.1	AY549317.1	AY843816.1	--	--	--	--	--	AY844569.1	AY844795.1	--	AY844041.1
<i>Hypsiboas bichoffi</i>	AY549324.1	AY549324.1	AY843833.1	--	--	--	--	AY844398.1	AY844586.1	--	--	--
<i>Hypsiboas caingua</i>	AY549326.1	AY549326.1	AY843838.1	KF751479.1	--	--	--	--	AY844591.1	AY844812.1	--	AY844057.1

<i>Hypsiboas crepitans</i>	AY843621.1	AY843621.1	AY843850.1	KF751482.1	--	--	--	AY844412.1	AY844601.1	--	--	AY844067.1
<i>Hypsiboas curupi</i>	--	--	--	KF751483.1	--	--	--	--	--	--	--	--
<i>Hypsiboas faber</i>	AY549334.1	AY549334.1	AY843857.1	--	--	--	--	--	AY844607.1	AY844825.1	--	--
<i>Hypsiboas guenteri</i>	AY843631.1	AY843631.1	AY843863.1	--	--	--	--	--	AY844612.1	AY844830.1	--	--
<i>Hypsiboas joaquina</i>	AY549340.1	AY549340.1	AY843867.1	KF751484.1	--	--	--	AY844421.1	AY844616.1	AY844834.1	--	--
<i>Hypsiboas latistriatus</i>	AY549360.1	AY549360.1	AY843921.1	--	--	--	--	--	AY844668.1	--	--	--
<i>Hypsiboas leptolineatus</i>	AY549341.1	AY549341.1	AY843872.1	--	--	--	--	AY844424.1	AY844621.1	AY844839.1	--	AY844083.1
<i>Hypsiboas lundii</i>	AY843639.1	AY843639.1	AY843874.1	--	--	--	--	--	AY844623.1	AY844841.1	--	AY844085.1
<i>Hypsiboas marginatus</i>	AY549342.1	AY549342.1	AY843875.1	KF751486.1	--	--	--	AY844426.1	AY844624.1	AY844842.1	--	--
<i>Hypsiboas pardalis</i>	AY843651.1	AY843651.1	AY843891.1	--	--	--	--	--	AY844637.1	AY844855.1	--	AY844096.1
<i>Hypsiboas polytaenius</i>	AY843655.1	AY843655.1	AY843895.1	--	--	--	AY819124.1	AY844443.1	AY844641.1	AY844859.1	--	--
<i>Hypsiboas prasinus</i>	AY549347.1	AY549347.1	AY843896.1	--	--	--	--	--	AY844642.1	AY844860.1	--	AY844100.1
<i>Hypsiboas pulchellus</i>	AY549352.1	AY549352.1	AY843898.1	--	--	--	--	AY844445.1	AY844644.1	AY844862.1	--	AY844102.1
<i>Hypsiboas</i>	AY549353.1	AY549353.1	AY843899.1	--	--	--	--	--	AY844645.1	--	--	--

*punctatus*

<i>Hypsiboas raniceps</i>	AY843657.1	AY843657.1	AY843900.1	KF751489.1	--	--	AY819125.1	--	JQ023459.1	AY844863.1	--	AY844103.1
<i>Hypsiboas semiguttatus</i>	AY549358.1	AY549358.1	AY843908.1	--	--	--	--	AY844452.1	AY844655.1	AY844870.1	--	--
<i>Hypsiboas semilineatus</i>	AY843779.1	AY843779.1	AY843909.1	KF751491.1	--	--	--	AY844453.1	AY844656.1	AY844871.1	--	AY844108.1
<i>Ischnocnema abdita</i>	JX267325.1	JX267471.1	--	--	--	--	--	JX267551.1	--	--	--	JX267687.1
<i>Ischnocnema bolbodactyla</i>	JX267327.1	JX267476.1	--	--	--	--	--	JX267557.1	--	--	--	JX267692.1
<i>Ischnocnema concolor</i>	JX267366.1	JX267366.1	--	--	--	--	--	JX267595.1	--	--	--	JX267728.1
<i>Ischnocnema erythromera</i>	JX267340.1	JX267340.1	--	--	--	--	--	JX267596.1	--	--	--	JX267730.1
<i>Ischnocnema hoehnei</i>	JX267347.1	--	--	--	--	--	--	JX267616.1	--	--	--	JX267752.1
<i>Ischnocnema holti</i>	JX267306.1	JX267306.1	--	--	--	--	--	JX267617.1	--	--	--	JX267754.1
<i>Ischnocnema izecksoni</i>	JX267307.1	JX267307.1	--	--	--	--	--	JX267618.1	--	--	--	JX267756.1
<i>Ischnocnema juipoca</i>	DQ283093.1	DQ283093.1	--	--	DQ284143.1	--	--	JX267624.1	DQ283809.1	--	--	JX267762.1
<i>Ischnocnema melonopygia</i>	JX267430.1	JX267292.1	--	--	--	--	--	JX267634.1	--	--	--	JX267771.1



*gracilis**Leptodactylus jolyi*

KC470093.1	KC477250.1	KF548066.1	--	--	--	--	--	--	KF613581.1	--	--	--
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*Leptodactylus latrans*

AY143353.1	AY162395.1	KC603976.1	DQ306492.1	DQ284104.1	--	--	KC604055.1	KC604026.1	AY844681.1	--	--	KC604082.1
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*Leptodactylus macrosternum*

KC470106.1	KC477255.1	KF548084.1	--	--	--	--	--	--	KF613594.1	--	--	--
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*Leptodactylus mystaceus*

KC470101.1	KC477252.1	KF548080.1	--	--	--	--	KC604058.1	KC604036.1	KF613589.1	--	--	JN706737.1
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*Leptodactylus mystacinus*

KC470105.1	KC477256.1	KF548067.1	--	--	--	--	--	AY323771.1	AY323747.1	--	--	--
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*Leptodactylus notoaktites*

--	KC477254.1	KF548083.1	--	--	--	--	--	--	KF613592.1	--	--	--
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*Leptodactylus plaumanni*

--	--	AY332231.1	--	--	--	--	--	--	--	--	--	--
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*Leptodactylus podicipinus*

KC470094.1	KC477245.1	KF548073.1	--	--	--	--	--	--	KF613582.1	--	--	--
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*Lithobates palmipes*

DQ347037.1	DQ283384.1	--	--	DQ284369.1	--	--	--	DQ347263.1	DQ347382.1	DQ282847.1	--	DQ347170.1
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*Luetkenotyphlus brasiliensis*

--	EF107198.1	--	EF107483.1	--	EF107262.1	--	--	EF107321.1	--	--	EF107424.1	--
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*Macrogenioglottus alipioi*

KF214098.1	FJ685685.1	FJ685665.1	--	--	--	--	--	FJ685705.1	KF214200.1	--	--	--
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*Melanophryniscus setiba*

JX961679.1	JX961679.1	--	JX961675.1	--	--	--	--	--	JX961677.1	--	--	--
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<i>Myersiella microps</i>	--	KC179973.1	--	--	--	--	--	--	--	--	--	--	KC180279.1
<i>Odontophrynus carvalhoi</i>	KF214100.1	FJ685687.1	FJ685667.1	--	--	--	--	FJ685707.1	KF214202.1	--	--	--	--
<i>Ololygon catharinae</i>	AY843756.1	AY843756.1	AY844001.1	--	--	--	AY819140.1	AY844517.1	AY844742.1	AY844941.1	--	--	--
<i>Ololygon obtriangulata</i>	GQ896259.1	--	--	--	--	--	--	--	--	--	--	--	--
<i>Paratelmatobius cardosoi</i>	EU224404.1	EU224404.1	--	--	--	--	--	--	--	--	--	--	--
<i>Paratelmatobius gaigeae</i>	EU224397.1	EU224397.1	--	--	--	--	--	--	--	--	--	--	--
<i>Paratelmatobius poecilogaster</i>	EU224400.1	EU224400.1	--	--	--	--	--	--	--	--	--	--	--
<i>Phasmahyla cochranae</i>	--	GQ366309.1	--	GQ365996.1	--	--	GQ366038.1	GQ366076.1	GQ366105.1	GQ366164.1	--	--	--
<i>Phasmahyla exilis</i>	GQ366231.1	GQ366310.1	GQ365920.1	GQ365997.1	--	--	GQ366039.1	GQ366077.1	GQ366106.1	GQ366165.1	--	--	--
<i>Phasmahyla guttata</i>	GQ366232.1	GQ366232.1	GQ365921.1	--	--	--	GQ366040.1	--	GQ366107.1	--	--	--	--
<i>Phasmahyla jandata</i>	GQ366233.1	GQ366312.1	GQ365922.1	GQ365999.1	--	--	GQ366042.1	--	GQ366108.1	GQ366166.1	--	--	--
<i>Phrynomedusa marginata</i>	GQ366234.1	GQ366313.1	GQ365923.1	--	--	--	--	GQ366078.1	GQ366109.1	GQ366167.1	--	--	GQ366199.1
<i>Phyllodytes luteolus</i>	AY843721.1	AY843721.1	AY843966.1	--	--	--	GQ366043.1	AY844494.1	AY844708.1	AY844913.1	--	--	AY844150.1



*signifer*

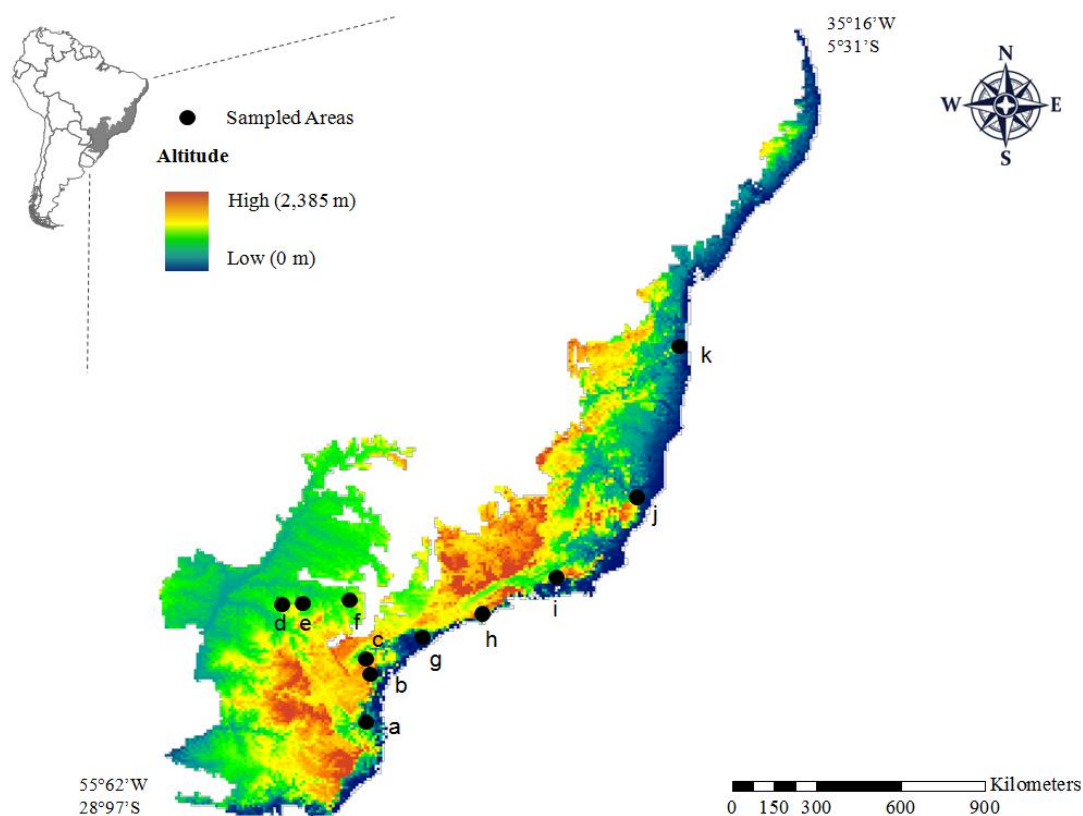
<i>Physalaemus spiniger</i>	KC692086.1	KC692086.1	--	--	--	--	--	--	--	--	--	--
<i>Pipa carvalhoi</i>	DQ283251	DQ283251	--	--	DQ284277	--	--	--	DQ283922	DQ282774	--	--
<i>Pristimantis paulodutraii</i>	JX267399.1	JX267360.1	--	--	--	--	--	JX267573.1	--	--	--	JX267707.1
<i>Pristimantis ramagii</i>	JX267319.1	JX267319.1	--	--	--	--	--	JX267659.1	--	--	--	JX267798.1
<i>Pristimantis vinhai</i>	X267411.1	--	--	--	--	--	--	JX267592.1	--	--	--	JX267725.1
<i>Proceratophrys appendiculata</i>	KF214131.1	--	KF214173.1	--	--	--	--	KF214190.1	KF214232.1	--	--	--
<i>Proceratophrys avelinoi</i>	KF214102.1	FJ685691.1	FJ685671.1	--	--	--	--	FJ685711.1	KF214204.1	--	--	--
<i>Proceratophrys bigibbosa</i>	KF214103.1	FJ685692.1	FJ685672.1	--	--	--	--	FJ685712.1	KF214205.1	--	--	--
<i>Proceratophrys cristiceps</i>	KF214106.1	FJ685695.1	FJ685675.1	--	--	--	--	FJ685715.1	KF214208.1	--	--	--
<i>Proceratophrys cururu</i>	KF214107.1	FJ685696.1	FJ685676.1	--	--	--	--	FJ685716.1	KF214209.1	--	--	--
<i>Proceratophrys laticeps</i>	KF214109.1	FJ685698.1	FJ685678.1	--	--	--	--	FJ685718.1	KF214211.1	--	--	--
<i>Proceratophrys melanopogon</i>	KF214120.1	KF214142.1	KF214164.1	--	--	--	GQ345270.1	KF214181.1	KF214217.1	--	--	--





*Zachaenus*  
*parvulus* KC593362.1 KC593362.1 -- -- -- -- -- -- -- -- -- --

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**Figure S1.** Map showed the localtion of Brazilian Atlantic Forest hotspot (in gray) in South American territory. The black circle indicates the Protected areas visited as complementary field works. a. Parque Ecológico Spitzkopf, SC; b. Parque Estadual Pico do Marumbi, PR; c. Parque estadual de Campinhos, PR; d. Parque municipal das Perobas, PR; e. Parque Estadual Mata dos Godoy, PR; f. Refugio municipal da vida Silvestre, Horto Florestal de Jacarezinho, PR; g. Estação Ecológica de Juréia-Itatins, SP; h. Parque Estadual da Serra do Mar Núcleo Caraguatatuba, SP; i. Parque Nacional da Serra dos Órgãos, RJ; j. Reserva Biológica Augusto Ruschi, ES; k. Reserva Biológica de Una, BA. From January of 2014 (PA h five days), July 2014 (PA f three days), November 2014 (PAs d, f three days in each), December 2014 (PA d two days), January to March 2015 (PAs a, b, g-k seven days in each), May 2015 (PA f three days), November 2015 (PA d two days), December 2015 (PA c five days), February 2016 (PAs e two days, f three days), March 2016 (PA d one day, e two days). We sampled amphibians using acoustic and visual nocturnal and diurnal surveys (Crump & Scott Jr, 1994; Zimmerman, 1994). We searched at the margins of water bodies, in the streams, and along 2000 m forest transects. The species collected were euthanized with hydrochloride benzocaine > or = 250 mg / L (ASH, 2004). Number of authorization / license SISBIO: 30344.

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APÊNDICE 3 - **Table S1.** Lista das espécies estudadas e seus mecanismos antipredação.

Espécies	Produzir Secreções	Camuflagem	Imobilidade	Contrair	Inflar o corpo	Elevar o corpo	Esticar as pernas	Escapar	Aposematismo	Descarga Cloacal	Abrir a boca	Vocalizações defensivas	Luta contra o predador	Referências
<i>Adelophryne mucronatus</i>	___	Presença	Encolhido&Alerta& Tanatose	___	Ausência	Ausência	Ausência	Esconder	Ausência	___	Presença	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Adelophryne pachydactyla</i>	___	Presença	Encolhido&Alerta& Tanatose	___	Ausência	Ausência	Ausência	Esconder	Ausência	___	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Aparasphenodon brunoi</i>	Venenosa	Ausência	Encolhido&Alerta& Tanatose	Fragmose	Presença	Ausência	Ausência	Único pulo	Ausência	___	Ausência	___	Lutar& Cabeçada	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Aplastodiscus albosignatus</i>	Odorifera& Desagradavel	Presença	Encolhido	___	Ausência	Pernas esticadas verticalmente	Ausência	Único pulo	Ausência	___	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Aplastodiscus arildae</i>	Odorifera& Desagradavel	Presença	Encolhido&Alerta	Contração	Presença	Pernas esticadas verticalmente & Pernas esticadas lateralmente	Ausência	Único pulo	Ausência	Liquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação);

<i>Aplastodiscus callipygius</i>	Odorifera& Desagradavel	Presença	Encolhido	___	Ausência	Pernas esticadas verticalmente	Ausência	Único pulo	Ausência	Liquida	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Aplastodiscus cochranæ</i>	Odorifera& Desagradavel	Presença	Encolhido	Proteger o olho	Ausência	Pernas esticadas verticalmente	Ausência	Único pulo	Ausência	Liquida	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Aplastodiscus ehrhardti</i>	Odorifera& Desagradavel	Presença	Encolhido& Tanatose	Contração	Ausência	Ausência	Ausência	Varios pulos& Esconder	Ausência	Liquida	Presença	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Aplastodiscus leucopygius</i>	Odorifera& Desagradavel	Presença	Encolhido	Agachar	Presença	Pernas esticadas verticalmente	Ausência	Único pulo	Ausência	Liquida	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Aplastodiscus perviridis</i>	Odorifera& Desagradavel	Presença	Encolhido& Alerta	___	Ausência	Pernas esticadas verticalmente	Ausência	Único pulo	Ausência	Liquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Aplastodiscus weygoldti</i>	Odorifera& Desagradavel	Presença	Encolhido	___	Ausência	Ausência	Ausência	Único pulo	Ausência	Liquida	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação)
<i>Bokermannohyla astartea</i>	Odorifera& Desagradavel	Presença	Encolhido& Alerta	Contração	Presença	Ausência	Ausência	Varios pulos&Esconder	Ausência	Liquida	Ausência	___	Agredir com espinho	Haddad et al 2013; Ferreira et al (em preparação);

													Presente estudo	
<i>Bokermannohyla circumdata</i>	Odorifera&		Encolhido&										Lutar&	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
	Desagradavel	Presença	Tanatose	Agachar	Ausência	Ausência	Ausência	Único pulo	Ausência	Liquida	Ausência	Grito de agônia	Agredir com espinho	
<i>Bokermannohyla hylax</i>	Odorifera&		Encolhido&										Agredir com espinho	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
	Desagradavel	Presença	Alerta	Agachar	Ausência	Ausência	Ausência	Único pulo	Ausência	Liquida	Ausência	---		
<i>Brachycephalus ephippium</i>	Venenosa	Ausência	Encolhido&	---	Presença	Ausência	Ausência	Varios pulos& Esconder	Total	---	Presença	---	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Brachycephalus hermogenesi</i>	---	Presença	Encolhido	---	Ausência	Ausência	Ausência	---	Ausência	---	Presença	---	---	Haddad et al 2013; Ferreira et al (em preparação)
<i>Brachycephalus pitanga</i>	Venenosa	Ausência	Encolhido	---	Ausência	Ausência	Ausência	Varios pulos& Esconder	Total	---	Presença	---	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Chiasmocleis capixaba</i>	Odorifera	Presença	Encolhido&	---	Ausência	Ausência	Ausência	Varios pulos	Ausência	Liquida	Ausência	---	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo

<i>Chiasmocleis leucosticta</i>	Odorifera	Presença	Alerta& Tanatose	___	Ausência	Ausência	Ausência	Único pulo& Esconder	Ausência	Liquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Chiasmocleis schubarti</i>	Odorifera	Presença	Encolhido& Alerta& Tanatose	___	Presença	Ausência	Ausência	Único pulo	Ausência	Liquida	Presença	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Crossodactylodes bokermanni</i>	___	Presença	Encolhido	___	Ausência	Ausência	Ausência	Único pulo	Ausência	Liquida	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação)
<i>Crossodactylodes izecksohni</i>	___	Presença	Encolhido	Agachar	Ausência	Ausência	Ausência	Único pulo	Ausência	Liquida	Ausência	___	Agredir com espinho	Haddad et al 2013; Ferreira et al (em preparação)
<i>Crossodactylus caramaschii</i>	Escorregadia	Presença	Alerta& Tanatose	___	Presença	Ausência	Ausência	Único pulo& Esconder	Ausência	Liquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Cycloramphus acangatan</i>	___	Presença	Encolhido& Tanatose	Contração	Presença	Ausência	Ausência	Único pulo	Ausência	Liquida	Presença	Grito de agônia	Lutar& Morder	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Cycloramphus eleutherodactylus</i>	___	Presença	Alerta	___	Ausência	Ausência	Ausência	___	Ausência	Liquida	Ausência	___	Morder	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo

<i>Dendrophryniscus brevipollicatus</i>	___	Presença	Encolhido&Alerta&Tanatose	Contração&Agachar	Ausência	Ausência	Presença	Varios pulos&Esconder	Ausência	Liquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Dendrophryniscus carvalhoi</i>	___	Presença	Encolhido&Alerta&Tanatose	Contração	Presença	Ausência	Presença	Único pulo	Ausência	___	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Dendrophryniscus leucomystax</i>	___	Presença	Encolhido&Alerta	Contração	Ausência	Ausência	Presença	Único pulo	Ausência	Liquida	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Dendrophryniscus proboscideus</i>	Venenosa	Presença	Encolhido&Alerta	Contração	Ausência	Ausência	Ausência	Único pulo	Ausência	___	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Dendropsophus anceps</i>	___	Presença	Encolhido&Alerta	Contração	Ausência	Ausência	Ausência	Único pulo	Ausência	___	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Dendropsophus bipunctatus</i>	___	Presença	Encolhido&Alerta&Tanatose	___	Ausência	Ausência	Ausência	Único pulo	Parcial	Liquida	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Dendropsophus branneri</i>	Odorifera	Presença	Encolhido&Alerta&	___	Presença	Ausência	Ausência	Varios pulos&	Ausência	Liquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et

			Tanatose					Esconder						al (em preparação); Presente estudo
<i>Dendropsophus elegans</i>	---	Presença	Encolhido& Alerta& Tanatose	---	Ausência	Ausência	Ausência	Único pulo	Ausência	Líquida	Ausência	---	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Dendropsophus giesleri</i>	Odorifera	Presença	Encolhido& Alerta& Tanatose	---	Ausência	Ausência	Ausência	Único pulo	Parcial	Líquida	Ausência	---	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Dendropsophus minutus</i>	Odorifera	Presença	Encolhido& Alerta& Tanatose	---	Presença	Ausência	Ausência	Varios pulos& Esconder	Ausência	Líquida	Ausência	---	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Dendropsophus nanus</i>	Odorifera& Adesiva	Presença	Encolhido& Alerta& Tanatose	---	Presença	Ausência	Ausência	Varios pulos& Esconder	Ausência	Líquida	Ausência	---	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Dendropsophus sanborni</i>	Odorifera	Presença	Encolhido& Alerta& Tanatose	---	Presença	Ausência	Ausência	Varios pulos& Esconder	Ausência	Líquida	Ausência	---	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Elachistocleis bicolor</i>	Odorifera	Presença	Encolhido& Tanatose	---	Presença	Pernas esticadas verticalmente	Ausência	Único pulo& Esconder	Parcial	Líquida	Ausência	---	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo

<i>Eleutherodactylus bilineatus</i>	___	Presença	Encolhido&Alerta&Tanatose	___	Ausência	Ausência	Ausência	Único pulo	Ausência	___	Presença	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Euparkerella tridactyla</i>	___	Presença	Encolhido	___	Ausência	Pernas esticadas verticalmente	Ausência	Único pulo	Ausência	___	Presença	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Fritziana ohausi</i>	___	Presença	Encolhido&Alerta&Tanatose	Contração	Ausência	Ausência	Ausência	Único pulo	Ausência	Liquida	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Gastrotheca albolineata</i>	Odorifera	Presença	Encolhido&Tanatose	Contração&Agachar	Presença	Ausência	Ausência	Único pulo	Ausência	Liquida	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Gastrotheca microdiscus</i>	___	Presença	Encolhido	Contração	Ausência	Ausência	Ausência	Único pulo	Ausência	___	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação)
<i>Haddadus binotatus</i>	Odorifera	Presença	Encolhido&Alerta&Tanatose	Agachar	Presença	Ausência	Ausência	Único pulo&Esconder	Ausência	Liquida	Presença	Grito de agônia	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Hylodes phyllodes</i>	Escorregadia	Presença	Alerta&Tanatose	___	Ausência	Ausência	Ausência	Único pulo&Esconder	Ausência	Liquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação);

														Presente estudo
<i>Hylomantis aspera</i>	Venosa	Presença	Encolhido	Contração	Ausência	Ausência	Ausência	Único pulo	Parcial	Líquida	Ausência	—	—	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
	Odorifera&		Encolhido& Alerta&											
<i>Hypsiboas albomarginatus</i>	Adesiva	Presença	Tanatose	Contração	Presença	Ausência	Ausência	Único pulo	Parcial	Líquida	Ausência	Grito de agonia	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
			Encolhido& Alerta&	Contração& Proteger o olho	Presença	Ausência	Ausência	Único pulo	Parcial	Líquida	Ausência	—	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Hypsiboas albopunctatus</i>	Odorifera	Presença	Tanatose		Presença	Ausência	Ausência	Único pulo	Parcial	Líquida	Ausência	—	Lutar	
			Encolhido& Alerta&	Abaixar o queixo	Ausência	Ausência	Ausência	Único pulo	Parcial	Líquida	Ausência	Grito de alarme	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Hypsiboas bischoffi</i>	Odorifera	Presença	Tanatose		Ausência	Ausência	Ausência	Único pulo	Parcial	Líquida	Ausência	—	Lutar	
			Encolhido& Alerta&		Ausência	Ausência	Ausência	Varios pulos&						Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Hypsiboas caingua</i>	Odorifera	Presença	Tanatose	—	Ausência	Ausência	Ausência	Esconder	Ausência	Líquida	Ausência	—	Lutar	
	Odorifera&		Encolhido& Alerta&											
<i>Hypsiboas crepitans</i>	Adesiva	Presença	Tanatose	—	Presença	Ausência	Ausência	Único pulo	Ausência	Líquida	Ausência	Grito de agonia	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo



<i>Hypsiboas faber</i>	Odorifera& Escorregadia	Presença	Encolhido& Alerta& Tanatose	Contração& Agachar&A baixar o queixo	Presença	Ausência	Ausência	Varios pulos& Esconder	Ausência	Liquida& Sólida	Ausência	Grito de agônia	Lutar& Agredir com espinho	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Hypsiboas pardalis</i>	Odorifera	Presença	Encolhido& Alerta	___	Presença	Ausência	Ausência	Único pulo	Ausência	Liquida	Ausência	Grito de agônia	Agredir com espinho& Cabeçada	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Hypsiboas prasinus</i>	Odorifera	Presença	Encolhido& Alerta	___	Ausência	Ausência	Ausência	Único pulo	Ausência	Liquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Hypsiboas punctatus</i>	Odorifera	Presença	Encolhido& Alerta	Contração	Ausência	Ausência	Ausência	Varios pulos& Esconder	Parcial	Liquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Hypsiboas raniceps</i>	Odorifera	Presença	Encolhido& Alerta& Tanatose	___	Presença	Ausência	Ausência	Único pulo	Ausência	Liquida	Ausência	Grito de agônia	Lutar& Agredir com espinho	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Hypsiboas semilineatus</i>	Odorifera& Adesiva	Presença	Encolhido& Alerta& Tanatose	Contração	Presença	Ausência	Ausência	Único pulo	Parcial	Liquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Ischnocnema</i>	___	Presença	Alerta&	___	Presença	Ausência	Ausência	Varios	Ausência	Liquida	Ausência	___	___	Haddad et al 2013; Ferreira et

<i>abdita</i>			Tanatose					pulos						al (em preparação); Presente estudo	
<i>Ischnocnema erythromera</i>	Odorifera	Presença	Tanatose	Alerta&	___	Presença	Ausência	Ausência	Esconder	Parcial	Liquida	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Ischnocnema nigriventris</i>	___	Presença	Tanatose	Alerta&	___	Ausência	Ausência	Ausência	Esconder	Ausência	Liquida	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Ischnocnema parva</i>	Odorifera	Presença	Tanatose	Alerta&	___	Presença	Ausência	Ausência	Varios pulos	Ausência	___	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Ischnocnema spanios</i>	___	Presença	Tanatose	Alerta&	___	Ausência	Ausência	Ausência	Varios pulos	Ausência	___	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Ischnocnema verrucosa</i>	___	Presença	Tanatose	Alerta&	___	Ausência	Ausência	Ausência	Varios pulos	Ausência	Liquida	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Itapotihyla langsdorffii</i>	Odorifera& Desagradavel	Presença	Tanatose	Encolhido& Alerta&	Contração& Agachar&P roteger o olho	Ausência	Ausência	Ausência	Único pulo	Ausência	Liquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo

<i>Leptodactylus fuscus</i>	Escorregadia	Presença	Encolhido&Alerta& Tanatose	___	Ausência	Ausência	Ausência	Varios pulos	Ausência	Liquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
													Lutar&	
													Agredir com espinho&	
<i>Leptodactylus latrans</i>	Escorregadia	Presença	Encolhido&Alerta& Tanatose	___	Presença	Pernas esticadas lateralmente	Ausência	Varios pulos	Ausência	Liquida	Ausência	___	Avançar&Cabeçada	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Leptodactylus mystaceus</i>	___	Presença	Encolhido&Alerta& Tanatose	___	Ausência	Ausência	Ausência	Varios pulos	Parcial	Liquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Leptodactylus mystacinus</i>	Venosa	Presença	Encolhido&Alerta& Tanatose	___	Presença	Ausência	Ausência	Varios pulos	Ausência	Liquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Leptodactylus notoaktites</i>	Odorifera	Presença	Encolhido&Alerta& Tanatose	___	Presença	Ausência	Ausência	Varios pulos	Ausência	Liquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Leptodactylus podicipinus</i>	Odorifera	Presença	Encolhido&Alerta& Tanatose	___	Presença	Ausência	Ausência	Único pulo&Esconder	Ausência	Liquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação);

														Presente estudo
<i>Macrogenioglottus alipioi</i>	Venenosa	Presença	Encolhido	Contração	Presença	Pernas esticadas verticalmente & Pernas esticadas lateralmente	Ausência	Único pulo	Parcial	Liquida	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Myersiella microps</i>	Odorifera	Presença	Tanatose	___	Presença	Pernas esticadas verticalmente	Presença	Esconder	Ausência	___	Ausência	Grito de agônia	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Odontophrynus carvalhoi</i>	Venenosa	Presença	Encolhido	Contração	Presença	Ausência	Ausência	Único pulo	Ausência	___	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Oolygon catharinae</i>	Odorifera & Adesiva	Presença	Tanatose	___	Presença	Ausência	Ausência	Esconder	Ausência	Liquida	Presença	Grito de agônia	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Paratelmatobius cardosoi</i>	___	Presença	Tanatose	___	Ausência	Ausência	Ausência	Esconder	Parcial	___	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Phasmahyla guttata</i>	Venenosa	Presença	Encolhido	Contração	Ausência	Ausência	Ausência	Único pulo	Parcial	___	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo

<i>Phyllodytes luteolus</i>	___	Ausência	Encolhido&Alerta&Tanatose	___	Ausência	Ausência	Ausência	Esconder	Ausência	Líquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Phyllomedusa bahiana</i>	Venosa	Presença	Encolhido	Contração&Abaixar o queixo	Ausência	Ausência	Ausência	Único pulo	Parcial	___	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Phyllomedusa burmeisteri</i>	Venosa	Presença	Encolhido	Contração&Abaixar o queixo	Ausência	Ausência	Ausência	Único pulo	Parcial	___	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Phyllomedusa distincta</i>	Venosa	Presença	Encolhido	Contração	Ausência	Ausência	Ausência	Único pulo	Parcial	___	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Phyllomedusa nordestina</i>	Venosa	Presença	Encolhido	Contração	Presença	Ausência	Ausência	Único pulo	Parcial	___	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Phyllomedusa rohdei</i>	Venosa	Presença	Encolhido	Contração	Ausência	Ausência	Ausência	Único pulo	Parcial	___	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Phyllomedusa tetraploidea</i>	Venosa	Presença	Encolhido	Contração	Presença	Ausência	Ausência	Único pulo	Parcial	___	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo

														al (em preparação); Presente estudo
<i>Physalaemus crombiei</i>	___	Presença	Encolhido& Alerta& Tanatose	Agachar	Ausência	Elevação	Ausência	Varios pulos	Ausência	Liquida	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Physalaemus cuvieri</i>	Escorregadia	Presença	Encolhido& Alerta& Tanatose	___	Ausência	Ausência	Ausência	Único pulo& Esconder	Parcial	Liquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Physalaemus olfersii</i>	___	Presença	Encolhido& Alerta& Tanatose	___	Ausência	Ausência	Ausência	Único pulo& Esconder	Ausência	Liquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação)
<i>Physalaemus signifer</i>	___	Presença	Encolhido& Alerta& Tanatose	___	Presença	Ausência	Ausência	Único pulo& Esconder	Ausência	Liquida	Ausência	Grito de agônia	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Pipa carvalhoi</i>	Escorregadia	Presença	Encolhido& Alerta& Tanatose	___	Ausência	Ausência	Ausência	Único pulo& Esconder	Ausência	___	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Pristimantis paulodutraii</i>	Odorifera	Presença	Encolhido& Alerta& Tanatose	___	Ausência	Ausência	Ausência	Varios pulos	Ausência	Liquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo

<i>Pristimantis ramagii</i>	Odorifera	Presença	Encolhido& Alerta& Tanatose	___	Ausência	Ausência	Ausência	Varios pulos	Ausência	Liquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Pristimantis vinhai</i>	Odorifera	Presença	Encolhido& Alerta& Tanatose	___	Ausência	Ausência	Ausência	Varios pulos	Parcial	Liquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Proceratophrys appendiculata</i>	___	Presença	Encolhido& Alerta	___	Ausência	Ausência	Presença	Único pulo	Ausência	___	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Proceratophrys avelinoi</i>	___	Presença	Encolhido& Alerta	Contração	Ausência	Ausência	Ausência	Único pulo	Parcial	Liquida	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Proceratophrys cururu</i>	Odorifera	Presença	Encolhido& Alerta	Agachar	Presença	Ausência	Ausência	Único pulo	Ausência	___	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação)
<i>Proceratophrys laticeps</i>	___	Presença	Encolhido& Alerta	Contração	Ausência	Ausência	Presença	Único pulo	Ausência	___	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Proceratophrys melanopogon</i>	___	Presença	Encolhido& Alerta	___	Ausência	Ausência	Presença	Único pulo	Ausência	___	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação);

														Presente estudo
<i>Proceratophrys renalis</i>	___	Presença	Encolhido&Alerta&Tanatose	Contração	Ausência	Ausência	Presença	Único pulo	Ausência	___	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Proceratophrys schirchi</i>	___	Presença	Encolhido&Alerta	___	Ausência	Elevação lateral	Ausência	Único pulo	Ausência	Liquida	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação);
<i>Pseudis bolbodactyla</i>	Escorregadia	Presença	Encolhido&Alerta&Tanatose	___	Presença	Ausência	Ausência	Único pulo&Esconder	Ausência	Liquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Rhinella crucifer</i>	Venenosa	Presença	Encolhido&Alerta	Agachar	Presença	Ausência	Ausência	Único pulo	Ausência	Liquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Rhinella granulosa</i>	Venenosa	Presença	Encolhido&Alerta	Contração	Presença	Ausência	Presença	Único pulo	Ausência	Liquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Rhinella hoogmoedi</i>	Venenosa	Presença	Encolhido&Alerta	Contração	Presença	Ausência	Ausência	Único pulo	Ausência	Liquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo



<i>Rhinella ictericus</i>	Venenosa	Presença	Encolhido&Alerta& Tanatose	___	Presença	Elevação lateral	Ausência	Único pulo	Ausência	Líquida	Ausência	Grito de agônia	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Rhinella schneideri</i>	Venenosa	Presença	Encolhido&Alerta& Tanatose	Contração	Presença	Elevação lateral	Ausência	Único pulo	Ausência	Líquida	Ausência	Grito de agônia	Avançar&Cabeçada	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Scinax crospedospilus</i>	Odorifera	Presença	Encolhido&Alerta	___	Ausência	Ausência	Ausência	Vários pulos	Ausência	Líquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Scinax fuscovarius</i>	Odorifera	Presença	Encolhido&Alerta& Tanatose	___	Presença	Ausência	Ausência	Único pulo&Esconder	Parcial	Líquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Scinax hayii</i>	Odorifera	Presença	Encolhido&Alerta& Tanatose	___	Presença	Ausência	Ausência	Único pulo&Esconder	Parcial	Líquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Scinax similis</i>	___	Presença	Encolhido&Alerta& Tanatose	___	Ausência	Ausência	Ausência	Único pulo&Esconder	Parcial	Líquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Scythrophrys sawayae</i>	___	Presença	Encolhido&Alerta	___	Ausência	Ausência	Presença	___	Ausência	___	Ausência	___	___	Haddad et al 2013; Ferreira et

														al (em preparação)
<i>Siphonops annulatus</i>	Venenosa	Presença	Encolhido	___	Ausência	Ausência	Ausência	Esconder	Ausência	___	Ausência	___	Movimentos erraticos	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Stereocyclops incrassatus</i>	___	Presença	Encolhido	___	Ausência	Ausência	Presença	___	Ausência	___	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Trachycephalus mesophaeus</i>	Venenosa	Presença	Encolhido&Alerta	Contração	Presença	Ausência	Ausência	Único pulo	Ausência	Liquida	Ausência	___	Lutar	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Trachycephalus typhonius</i>	Venenosa	Presença	Encolhido&Alerta&Tanatose	___	Presença	Ausência	Ausência	Único pulo	Ausência	Liquida	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo
<i>Xenohyla truncata</i>	___	Presença	Encolhido&Alerta	___	Presença	Pernas esticadas lateralmente	Ausência	___	Ausência	___	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação)
<i>Zachaenus parvulus</i>	___	Presença	Encolhido&Tanatose	___	Presença	Ausência	Presença	Único pulo	Parcial	___	Ausência	___	___	Haddad et al 2013; Ferreira et al (em preparação); Presente estudo

**Table S2.** Lista das espécies estudadas e suas respectivas características funcionais.

<b>Espécies</b>	<b>Range</b>	<b>Atividade</b>	<b>Tamanho</b>	<b>Habito</b>	<b>Habitat</b>	<b>Modo de desenvolvimento reprodutivo</b>	<b>Referências</b>
<i>Adelophryne mucronatus</i>	38	Noturno	Pequeno	Criptozoico	Áreas florestais	Direto	Lourenço-de-Moraes et al 2012; Haddad et al. 2013; presente estudo
<i>Adelophryne pachydactyla</i>	301	Noturno	Pequeno	Criptozoico	Áreas florestais	Direto	Lourenço-de-Moraes et al 2012; Haddad et al. 2013; presente estudo
<i>Aparasphenodon brunoi</i>	2532	Noturno	Médio	Arborea	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Aplastodiscus albosignatus</i>	1077	Noturno	Médio	Arborea	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Aplastodiscus arildae</i>	1903	Noturno	Médio	Arborea	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Aplastodiscus callipygius</i>	404	Noturno	Médio	Arborea	Áreas florestais	indireto	Haddad et al 2013
<i>Aplastodiscus cochranæ</i>	173	Noturno	Médio	Arborea	Áreas florestal/área aberta	indireto	Haddad et al 2013
<i>Aplastodiscus ehrhardti</i>	395	Noturno	Médio	Arborea	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Aplastodiscus leucoeygius</i>	1588	Noturno	Médio	Arborea	Áreas florestais	indireto	Haddad et al 2013
<i>Aplastodiscus perviridis</i>	5781	Noturno	Médio	Arborea	Áreas florestais	indireto	Haddad et al 2013; presente estudo

<i>Aplastodiscus weygoldti</i>	34	Noturno	Médio	Arborea	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Bokermannohyla astartea</i>	1101	Noturno	Médio	Arborea	Áreas florestais	indireto	Haddad et al 2013
<i>Bokermannohyla circumdata</i>	1109	Noturno	Médio	Arborea	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Bokermannohyla hylax</i>	979	Noturno	Médio	Arborea	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Brachycephalus ephippium</i>	1863	Diurno	Pequeno	Criptoico	Áreas florestais	Direto	Haddad et al 2013; presente estudo
<i>Brachycephalus hermogenesi</i>	653	Diurno	Pequeno	Criptoico	Áreas florestais	Direto	Haddad et al 2013
<i>Brachycephalus pitanga</i>	8	Diurno	Pequeno	Criptoico	Áreas florestais	Direto	Haddad et al 2013; presente estudo
<i>Chiasmocleis capixaba</i>	495	Noturno	Pequeno	Criptoico	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Chiasmocleis leucosticta</i>	1533	Noturno e diurno	Pequeno	Criptoico	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Chiasmocleis schubarti</i>	1398	Noturno	Médio	Criptoico	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Crossodactylodes bokermanni</i>	36	Noturno	Pequeno	Arborea	Áreas florestais	indireto	Haddad et al 2013
<i>Crossodactylodes izecksohni</i>	4	Noturno	Pequeno	Arborea	Áreas florestais	indireto	Haddad et al 2013

<i>Crossodactylus caramaschii</i>	672	Noturno e diurno	Pequeno	Reofilico	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Cycloramphus acangatan</i>	125	Noturno	Médio	Criptozoico	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Cycloramphus eleutherodactylus</i>	1891	Noturno	Médio	Criptozoico	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Dendrophryniscus brevipollicatus</i>	758	Noturno	Pequeno	Arborea	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Dendrophryniscus carvalhoi</i>	31	—	Pequeno	Criptozoico	Áreas florestais	indireto	Haddad et al 2013
<i>Dendrophryniscus leucomystax</i>	905	Noturno	Pequeno	Criptozoico	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Dendrophryniscus proboscideus</i>	1353	Noturno	Médio	Criptozoico	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Dendropsophus anceps</i>	2294	Noturno	Médio	Arborea	Áreas aberta	indireto	Haddad et al 2013
<i>Dendropsophus bipunctatus</i>	1622	Noturno	Pequeno	Arborea	Áreas aberta	indireto	Haddad et al 2013; presente estudo
<i>Dendropsophus branneri</i>	3285	Noturno	Pequeno	Arborea	Áreas aberta	indireto	Haddad et al 2013; presente estudo
<i>Dendropsophus elegans</i>	5047	Noturno	Médio	Arborea	Áreas aberta	indireto	Haddad et al 2013; presente estudo
<i>Dendropsophus giesleri</i>	855	Noturno	Pequeno	Arborea	Áreas florestais	indireto	Haddad et al 2013; presente estudo

<i>Dendropsophus minutus</i>	9676	Noturno	Pequeno	Arborea	Áreas aberta	indireto	Haddad et al 2013; presente estudo
<i>Dendropsophus nanus</i>	7282	Noturno	Pequeno	Arborea	Áreas aberta	indireto	Haddad et al 2013; presente estudo
<i>Dendropsophus sanorni</i>	3410	Noturno	Pequeno	Arborea	Áreas aberta	indireto	Haddad et al 2013; presente estudo
<i>Elaschitocleis bicolor</i>	3457	Noturno	Médio	Fossorial	Áreas aberta	indireto	Haddad et al 2013; presente estudo
<i>Eleutherodactylus bilineatus</i>	550	Noturno	Pequeno	Fossorial	Áreas florestais	Direto	Haddad et al 2013; presente estudo
<i>Euparkerella tridactyla</i>	2	Noturno e diurno	Pequeno	Criptozoico	Áreas florestais	Direto	Haddad et al 2013; presente estudo
<i>Fritziana ohausi</i>	977	Noturno	Médio	Arborea	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Gastrotheca albolineata</i>	703	Noturno	Médio	Arborea	Áreas florestais	Direto	Haddad et al 2013; presente estudo
<i>Gastrotheca microdiscus</i>	1645	Noturno	Médio	Arborea	Áreas florestais	Direto	Haddad et al 2013
<i>Haddadus binotatus</i>	4965	Noturno	Médio	Criptozoico	Áreas florestais	Direto	Haddad et al 2013; presente estudo
<i>Hylodes phyllodes</i>	468	Diurno	Pequeno	Reofilico	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Hylomantis aspera</i>	234	Noturno	Médio	Arborea	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Hypsiboas albomarginatus</i>	4594	Noturno	Médio	Arborea	Áreas aberta	indireto	Haddad et al 2013; presente estudo
<i>Hypsiboas albopunctatus</i>	7663	Noturno	Médio	Arborea	Áreas aberta	indireto	Haddad et al 2013; presente estudo

<i>Hypsiboas bischoffi</i>	2422	Noturno	Médio	Arborea	Áreas aberta	indireto	Haddad et al 2013; presente estudo
<i>Hypsiboas caingua</i>	1241	Noturno	Médio	Arborea	Áreas aberta	indireto	Haddad et al 2013; presente estudo
<i>Hypsiboas crepitans</i>	7978	Noturno	Médio	Arborea	Áreas aberta	indireto	Haddad et al 2013
<i>Hypsiboas faber</i>	9183	Noturno	Médio	Arborea	Áreas florestal/área aberta	indireto	Haddad et al 2013; presente estudo
<i>Hypsiboas pardalis</i>	2279	Noturno	Médio	Arborea	Áreas florestal/área aberta	indireto	Haddad et al 2013; presente estudo
<i>Hypsiboas prasinus</i>	2974	Noturno	Médio	Arborea	Áreas florestal/área aberta	indireto	Haddad et al 2013; presente estudo
<i>Hypsiboas punctatus</i>	6355	Noturno	Médio	Arborea	Áreas aberta	indireto	Haddad et al 2013; presente estudo
<i>Hypsiboas raniceps</i>	3978	Noturno	Médio	Arborea	Áreas aberta	indireto	Haddad et al 2013; presente estudo
<i>Hypsiboas semilineatus</i>	5012	Noturno	Médio	Arborea	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Ischnocnema abdita</i>	23	___	Pequeno	Criptozoico	Áreas florestais	Direto	Haddad et al 2013; presente estudo
<i>Ischnocnema erythromera</i>	35	Noturno	Médio	Criptozoico	Áreas florestais	Direto	Haddad et al 2013; presente estudo
<i>Ischnocnema nigriventris</i>	4	___	Pequeno	Fossorial	Áreas florestais	Direto	Haddad et al 2013; presente estudo
<i>Ischnocnema parva</i>	1423	Noturno	Pequeno	Criptozoico	Áreas florestais	Direto	Haddad et al 2013; presente estudo
<i>Ischnocnema spanios</i>	190	___	Pequeno	Criptozoico	Áreas florestais	Direto	Haddad et al 2013; presente estudo
<i>Ischnocnema</i>	855	Noturno	Pequeno	Criptozoico	Áreas florestais	Direto	Haddad et al 2013; presente estudo

*verrucosa*

<i>Itapotihyla langsdorffii</i>	4454	Noturno	Grande	Arborea	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Leptodactylus fuscus</i>	9759	Noturno	Médio	Terrestre	Áreas aberta	indireto	Haddad et al 2013; presente estudo
<i>Leptodactylus latrans</i>	9670	Noturno	Médio	Terrestre	Áreas florestal/área aberta	indireto	Haddad et al 2013; presente estudo
<i>Leptodactylus mystaceus</i>	830	Noturno	Médio	Criptoico	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Leptodactylus mystacinus</i>	9605	Noturno	Médio	Terrestre	Áreas aberta	indireto	Haddad et al 2013; presente estudo
<i>Leptodactylus notoakites</i>	2253	Noturno	Médio	Criptoico	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Leptodactylus podicipinus</i>	2529	Noturno	Médio	Criptoico	Áreas aberta	indireto	Haddad et al 2013; presente estudo
<i>Macrogenioglottus alipioi</i>	2512	Noturno	Médio	Criptoico	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Myersiella microps</i>	1236	Noturno	Médio	Fossorial	Áreas florestais	Direto	Haddad et al 2013; presente estudo
<i>Odontophrynus carvalhoi</i>	657	Noturno	Médio	Terrestre	Áreas florestal/área aberta	indireto	Haddad et al 2013
<i>Ololygon catharinae</i>	716	Noturno	Médio	Arborea	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Paratelmatobius cardosoi</i>	28	Noturno	Pequeno	Criptoico	Áreas florestais	indireto	Haddad et al 2013; presente estudo



<i>Phasmahyla guttata</i>	917	Noturno	Médio	Arborea	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Phyllodytes luteolus</i>	1671	Noturno	Pequeno	Fitotelmata	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Phyllomedusa bahiana</i>	337	Noturno	Médio	Arborea	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Phyllomedusa burmeisteri</i>	4892	Noturno	Médio	Arborea	Áreas florestal/área aberta	indireto	Haddad et al 2013; presente estudo
<i>Phyllomedusa distincta</i>	1766	Noturno	Médio	Arborea	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Phyllomedusa nordestina</i>	1805	Noturno	Médio	Arborea	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Phyllomedusa rohdei</i>	2021	Noturno	Médio	Arborea	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Phyllomedusa tetraploidea</i>	3440	Noturno	Médio	Arborea	Áreas florestal/área aberta	indireto	Haddad et al 2013; presente estudo
<i>Physalaemus crombiei</i>	1267	Noturno	Pequeno	Terrestre	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Physalaemus cuvieri</i>	9704	Noturno	Médio	Terrestre	Áreas aberta	indireto	Haddad et al 2013; presente estudo
<i>Physalaemus olfersii</i>	2346	Noturno	Médio	Criptozoico	Áreas florestais	indireto	Haddad et al 2013
<i>Physalaemus signifer</i>	1688	Noturno	Pequeno	Criptozoico	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Pipa carvalhoi</i>	2152	Noturno e diurno	Médio	Aquático	Áreas aberta	indireto	Haddad et al 2013; presente estudo
<i>Pristimantis paulodutraí</i>	708	Noturno	Médio	Criptozoico	Áreas florestais	Direto	Haddad et al 2013; presente estudo
<i>Pristimantis ramagii</i>	1272	Noturno	Pequeno	Criptozoico	Áreas florestais	Direto	Haddad et al 2013; presente estudo

<i>Pristimantis vinhai</i>	763	Noturno	Pequeno	Criptozoico	Áreas florestais	Direto	Haddad et al 2013; presente estudo
<i>Proceratophrys appendiculata</i>	1226	Noturno	Médio	Criptozoico	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Proceratophrys avelinoi</i>	2412	Noturno	Pequeno	Criptozoico	Áreas florestal/área aberta	indireto	Haddad et al 2013; presente estudo
<i>Proceratophrys cururu</i>	1	Noturno	Médio	Criptozoico	Áreas florestais	indireto	Haddad et al 2013
<i>Proceratophrys laticeps</i>	1287	Noturno	Médio	Criptozoico	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Proceratophrys melanopogon</i>	532	Noturno	Médio	Criptozoico	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Proceratophrys renalis</i>	1366	Noturno	Médio	Criptozoico	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Proceratophrys schirchi</i>	1531	Noturno	Médio	Criptozoico	Áreas florestais	indireto	Haddad et al 2013
<i>Pseudis bolbodactyla</i>	1951	Noturno	Médio	Aquático	Áreas aberta	indireto	Haddad et al 2013; presente estudo
<i>Rhinella crucifer</i>	1943	Noturno	Grande	Terrestre	Áreas florestal/área aberta	indireto	Haddad et al 2013; presente estudo
<i>Rhinella granulosa</i>	8540	Noturno	Médio	Terrestre	Áreas florestal/área aberta	indireto	Haddad et al 2013; presente estudo
<i>Rhinella hoogmoedi</i>	2257	Noturno	Médio	Terrestre	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Rhinella icterica</i>	7322	Noturno	Grande	Terrestre	Áreas florestal/área aberta	indireto	Haddad et al 2013; presente estudo

<i>Rhinella schneideri</i>	9416	Noturno	Grande	Terrestre	Áreas aberta	indireto	Haddad et al 2013; presente estudo
<i>Scinax crospedospilus</i>	777	Noturno	Médio	Arborea	Áreas florestais	indireto	Haddad et al 2013
<i>Scinax fuscovarios</i>	9363	Noturno	Médio	Arborea	Áreas aberta	indireto	Haddad et al 2013; presente estudo
<i>Scinax hayii</i>	1965	Noturno	Médio	Arborea	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Scinax similis</i>	451	Noturno	Médio	Arborea	Áreas aberta	indireto	Haddad et al 2013; presente estudo
<i>Scythroehrys sawayae</i>	226	Noturno	Pequeno	Criptozoico	Áreas florestais	indireto	Haddad et al 2013
<i>Siphonops annulatus</i>	8685	___	Grande	Fossorial	Áreas florestal/área aberta	Direto	Haddad et al 2013; presente estudo
<i>Stereocyclops incrassatus</i>	2051	Noturno	Médio	Criptozoico	Áreas florestais	indireto	Haddad et al 2013
<i>Trachycephalus mesophaeus</i>	5477	Noturno e diurno	Médio	Arborea	Áreas florestais	indireto	Haddad et al 2013; presente estudo
<i>Trachycephalus typhonius</i>	4913	Noturno	Médio	Arborea	Áreas florestal/área aberta	indireto	Haddad et al 2013; presente estudo
<i>Xenohyla truncata</i>	218	Noturno	Médio	Arborea	Áreas florestais	indireto	Haddad et al 2013
<i>Zachaenus parvulus</i>	268	Noturno	Pequeno	Criptozoico	Áreas florestais	indireto	Haddad et al 2013; presente estudo

**Table S3.** Lista das 115 espécies estudadas e suas respectivas números de acesso ao GenBank. (\*) indica o grupo externo.

Especies	12S	16S	Cytb	CXCR4	H3A	NCX1	POMC	RAG1	RHOD	SIA	SLC8A3	TYR
<i>Adelophryne mucronatus</i>	--	JX298291.1	JX298385.1	--	--	--	JX298112.1	JX298158.1	--	--	--	JX298213.1
<i>Adelophryne pachydactyla</i>	JX298259.1	JX298294.1	JX298387.1	--	--	--	JX298115.1	JX298161.1	--	--	--	JX298216.1
<i>Aparasphenodon brunoii</i>	AY843567.1	--	AY843789.1	--	--	--	--	AY844364.1	AY844541.1	AY844769.1	--	AY844023.1
<i>Aplastodiscus albosignatus</i>	AY843596.1	--	AY843817.1	--	--	--	--	AY844385.1	AY844570.1	AY844796.1	--	AY844042.1
<i>Aplastodiscus arildae</i>	AY843604.1	--	AY843825.1	--	--	--	--	AY844392.1	AY844578.1	AY844803.1	--	AY844049.1
<i>Aplastodiscus callipygius</i>	AY843614.1	AY843614.1	AY843840.1	--	--	--	--	AY844402.1	AY844592.1	AY844813.1	--	AY844058.1
<i>Aplastodiscus cochranee</i>	AY843568.1	AY843568.1	AY843790.1	--	--	--	--	AY844365.1	AY844542.1	AY844770.1	--	AY844024.1
<i>Aplastodiscus ehrhardti</i>	--	--	--	--	--	--	--	AY844456.1	--	--	--	--
<i>Aplastodiscus leucopygius</i>	AY843638.1	AY843638.1	AY843873.1	KF751466.1	--	--	--	AY844425.1	AY844622.1	AY844840.1	--	AY844084.1
<i>Aplastodiscus perviridis</i>	AY843569.1	AY843569.1	AY843791.1	KF751467.1	DQ284044.1	--	--	AY844366.1	AY844543.1	AY844771.1	--	AY844025.1
<i>Aplastodiscus weygoldti</i>	AY843685.1	AY843685.1	AY843931.1	--	--	--	--	AY844467.1	AY844678.1	AY844887.1	--	--
<i>Bokermanohyla astartea</i>	AY549322.1	AY549322.1	AY843827.1	--	--	--	AY819113.1	--	AY844580.1	--	--	--
<i>Bokermanohyla circumdata</i>	AY549328.1	--	AY843847.1	KF751468.1	--	--	--	AY844409.1	AY844598.1	AY844817.1	--	AY844064.1
<i>Bokermanohyla hylax</i>	AY549338.1	AY549338.1	AY843865.1	--	--	--	--	AY844419.1	AY844614.1	AY844832.1	--	AY844077.1
<i>Brachycephalus ephippium</i>	HM216368.1	HM216369.1	HM216367.1	GQ345180.1	GQ345212.1	GQ345228.1	GQ345256.1	HM216370.1	DQ283808.1	DQ282673.1	GQ345326.1	HQ435735.1
<i>Brachycephalus hermogenesi</i>	HQ435682.1	--	HQ435709.1	--	--	--	--	HQ435724.1	--	--	--	HQ435738.1



<i>Dendropsophus giesleri</i>	AY843629.1	AY843629.1	AY843860.1	--	--	--	--	AY844417.1	--	AY844827.1	--	AY844075.1
<i>Dendropsophus minutus</i>	AY549345.1	AY549345.1	AY843883.1	--	DQ284046.1	--	--	AY844432.1	DQ283758.1	--	--	AY844089.1
<i>Dendropsophus nanus</i>	AY549346.1	AY549346.1	AY843888.1	GQ365985.1	DQ284051.1	--	AY819123.1	AY844437.1	AY844634.1	AY844852.1	--	EF376132.1
<i>Dendropsophus samborni</i>	AY843663.1	AY843663.1	AY843906.1	--	--	--	--	AY844450.1	AY844653.1	AY844868.1	--	AY844106.1
<i>Elachistocleis bicolor</i>	JF836935.1	KC180005.1	--	--	--	--	--	--	JF837037.1	--	--	KC180318.1
<i>Eleutherodactylus bilineata</i>	JX267323.1	JX267323.1	--	--	--	--	--	JX267556.1	--	--	--	JX267691.1
<i>Euparkerella tridactyla</i>	--	--	--	--	--	--	--	KF625088.1	--	--	--	KF625110.1
<i>Fritziana ohausi</i>	--	JN157635.1	--	--	--	--	JN157629.1	KC844991.1	--	--	--	--
<i>Gastrotheca albolineata</i>	--	KC844924.1	--	--	--	--	KC844971.1	KC844992.1	--	--	--	--
<i>Gastrotheca microdiscus</i>	--	KC844933.1	--	--	--	--	KC844981.1	KC845000.1	--	--	--	--
<i>Haddadus binotatus</i>	KF740846.1	KF740846.1	JX298405.1	GQ345183.1	DQ284142.1	GQ345231.1	GQ345259.1	JX267548.1	DQ283807.1	GQ345309.1	GQ345329.1	JX267685.1
<i>Hylodes phyllodes</i>	DQ283096.1	DQ283096.1	DQ502587.1	--	DQ284146.1	--	--	KC604006.1	DQ283812.1	DQ282674.1	--	DQ282923.1
<i>Hylomantis aspera</i>	GQ366222.1	--	--	GQ365978.1	--	--	--	--	GQ366098.1	--	--	--
<i>Hypsiboas albomarginatus</i>	AY549316.1	AY549316.1	AY843815.1	--	--	--	--	AY844384.1	AY844568.1	AY844794.1	--	--
<i>Hypsiboas albopunctatus</i>	AY549317.1	AY549317.1	AY843816.1	--	--	--	--	--	AY844569.1	AY844795.1	--	AY844041.1
<i>Hypsiboas bichoffi</i>	AY549324.1	AY549324.1	AY843833.1	--	--	--	--	AY844398.1	AY844586.1	--	--	--
<i>Hypsiboas caingua</i>	AY549326.1	AY549326.1	AY843838.1	KF751479.1	--	--	--	--	AY844591.1	AY844812.1	--	AY844057.1
<i>Hypsiboas crepitans</i>	AY843621.1	AY843621.1	AY843850.1	KF751482.1	--	--	--	AY844412.1	AY844601.1	--	--	AY844067.1
<i>Hypsiboas faber</i>	AY549334.1	AY549334.1	AY843857.1	--	--	--	--	--	AY844607.1	AY844825.1	--	--

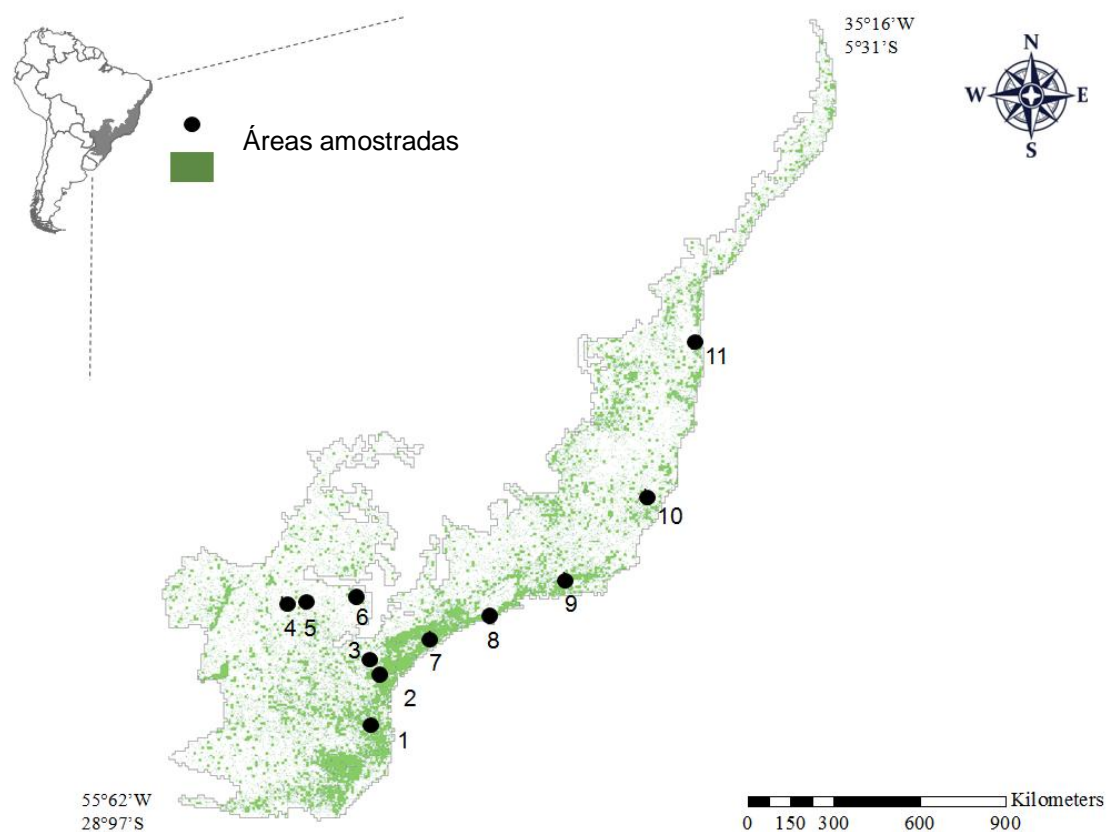
<i>Hypsiboas pardalis</i>	AY843651.1	AY843651.1	AY843891.1	--	--	--	--	--	AY844637.1	AY844855.1	--	AY844096.1
<i>Hypsiboas prasinus</i>	AY549347.1	AY549347.1	AY843896.1	--	--	--	--	--	AY844642.1	AY844860.1	--	AY844100.1
<i>Hypsiboas punctatus</i>	AY549353.1	AY549353.1	AY843899.1	--	--	--	--	--	AY844645.1	--	--	--
<i>Hypsiboas raniceps</i>	AY843657.1	AY843657.1	AY843900.1	KF751489.1	--	--	AY819125.1	--	JQ023459.1	AY844863.1	--	AY844103.1
<i>Hypsiboas semilineatus</i>	AY843779.1	AY843779.1	AY843909.1	KF751491.1	--	--	--	AY844453.1	AY844656.1	AY844871.1	--	AY844108.1
<i>Ischnocnema abdita</i>	JX267325.1	JX267471.1	--	--	--	--	--	JX267551.1	--	--	--	JX267687.1
<i>Ischnocnema erythromera</i>	JX267340.1	JX267340.1	--	--	--	--	--	JX267596.1	--	--	--	JX267730.1
<i>Ischnocnema nigriventris</i>	JX267398.1	JX267483.1	--	--	--	--	--	JX267568.1	--	--	--	JX267704.1
<i>Ischnocnema parva</i>	JX267446.1	KC569989.1	HQ435717.1	--	--	--	--	JX267657.1	--	--	--	JX267796.1
<i>Ischnocnema spanios</i>	JX267407.1	JX267490.1	--	--	--	--	--	JX267584.1	--	--	--	JX267717.1
<i>Ischnocnema verrucosa</i>	JX267457.1	JX267538.1	--	--	--	--	--	JX267670.1	--	--	--	JX267810.1
<i>Itapotihyla langsdorffii</i>	JQ868500.1	AY819511.1	AY843951.1	--	--	--	KF002003.1	--	AY844697.1	--	--	AY844137.1
<i>Leptodactylus fuscus</i>	AY905715.1	AY911284.1	KF548074.1	--	--	--	--	AY323770.1	KF613583.1	--	--	AY341760.1
<i>Leptodactylus latrans</i>	AY143353.1	AY162395.1	KC603976.1	DQ306492.1	DQ284104.1	--	KC604055.1	KC604026.1	AY844681.1	--	--	KC604082.1
<i>Leptodactylus mystaceus</i>	KC470101.1	KC477252.1	KF548080.1	--	--	--	KC604058.1	KC604036.1	KF613589.1	--	--	JN706737.1
<i>Leptodactylus mystacinus</i>	KC470105.1	KC477256.1	KF548067.1	--	--	--	--	AY323771.1	AY323747.1	--	--	--
<i>Leptodactylus notoakites</i>	--	KC477254.1	KF548083.1	--	--	--	--	--	KF613592.1	--	--	--
<i>Leptodactylus podicipinus</i>	KC470094.1	KC477245.1	KF548073.1	--	--	--	--	--	KF613582.1	--	--	--
<i>Macrogenioglottus alipioi</i>	KF214098.1	FJ685685.1	FJ685665.1	--	--	--	--	FJ685705.1	KF214200.1	--	--	--

<i>Myersiella microps</i>	--	KC179973.1	--	--	--	--	--	--	--	--	--	KC180279.1
<i>Odontophrynus carvalhoi</i>	KF214100.1	FJ685687.1	FJ685667.1	--	--	--	--	FJ685707.1	KF214202.1	--	--	--
<i>Oloolygon catharinae</i>	AY843756.1	AY843756.1	AY844001.1	--	--	--	AY819140.1	AY844517.1	AY844742.1	AY844941.1	--	--
<i>Paratelmatoebius cardosoi</i>	EU224404.1	EU224404.1	--	--	--	--	--	--	--	--	--	--
<i>Phasmahyla guttata</i>	GQ366232.1	GQ366232.1	GQ365921.1	--	--	--	GQ366040.1	--	GQ366107.1	--	--	--
<i>Phyllodytes luteolus</i>	AY843721.1	AY843721.1	AY843966.1	--	--	--	GQ366043.1	AY844494.1	AY844708.1	AY844913.1	--	AY844150.1
<i>Phyllomedusa bahiana</i>	GQ366251.1	GQ366251.1	GQ365940.1	--	--	--	--	--	GQ366126.1	GQ366177.1	--	GQ366205.1
<i>Phyllomedusa burmeisteri</i>	GQ366255.1	GQ366324.1	GQ365944.1	GQ366011.1	--	--	GQ366052.1	GQ366087.1	GQ366130.1	GQ366180.1	--	GQ366207.1
<i>Phyllomedusa distincta</i>	GQ366263.1	GQ366326.1	GQ365951.1	GQ366013.1	--	--	GQ366056.1	--	GQ366135.1	--	--	GQ366210.1
<i>Phyllomedusa nordestina</i>	GQ366271.1	GQ366330.1	GQ365959.1	GQ366016.1	--	--	--	GQ366091.1	GQ366143.1	--	--	GQ366215.1
<i>Phyllomedusa rohdei</i>	GQ366238.1	GQ366316.1	GQ365927.1	GQ366001.1	--	--	--	GQ366081.1	GQ366113.1	--	--	--
<i>Phyllomedusa tetraploidea</i>	GQ366284.1	GQ366334.1	GQ365972.1	GQ366021.1	--	--	GQ366066.1	GQ366096.1	GQ366155.1	GQ366194.1	--	GQ366218.1
<i>Physalaemus crombiei</i>	KC692082.1	KC692082.1	--	--	--	--	--	--	--	--	--	--
<i>Physalaemus cuvieri</i>	AY843729.1	AY162399	AY843975	--	--	--	AY819096	AY844499	--	AY844922	--	--
<i>Physalaemus olfersii</i>	KC692085.1	KC692085.1	--	--	--	--	--	--	--	--	--	--
<i>Physalaemus signifer</i>	DQ337209.1	DQ337209.1	--	--	--	--	--	--	--	--	--	--
<i>Pipa carvalhoi</i>	DQ283251	DQ283251	--	--	DQ284277	--	--	--	DQ283922	DQ282774	--	--
<i>Pristimantis paulodutra</i>	JX267399.1	JX267360.1	--	--	--	--	--	JX267573.1	--	--	--	JX267707.1
<i>Pristimantis ramagii</i>	JX267319.1	JX267319.1	--	--	--	--	--	JX267659.1	--	--	--	JX267798.1



<i>Pristimantis vinhai</i>	X267411.1	--	--	--	--	--	--	JX267592.1	--	--	--	JX267725.1
<i>Proceratophrys appendiculata</i>	KF214131.1	--	KF214173.1	--	--	--	--	KF214190.1	KF214232.1	--	--	--
<i>Proceratophrys avelinoi</i>	KF214102.1	FJ685691.1	FJ685671.1	--	--	--	--	FJ685711.1	KF214204.1	--	--	--
<i>Proceratophrys cururu</i>	KF214107.1	FJ685696.1	FJ685676.1	--	--	--	--	FJ685716.1	KF214209.1	--	--	--
<i>Proceratophrys laticeps</i>	KF214109.1	FJ685698.1	FJ685678.1	--	--	--	--	FJ685718.1	KF214211.1	--	--	--
<i>Proceratophrys melanopogon</i>	KF214120.1	KF214142.1	KF214164.1	--	--	--	GQ345270.1	KF214181.1	KF214217.1	--	--	--
<i>Proceratophrys renalis</i>	KF214111.1	JN814584.1	JN814523.1	--	--	--	--	JN814664.1	KF214213.1	--	--	--
<i>Proceratophrys schirchi</i>	KF214112.1	FJ685701.1	FJ685681.1	--	--	--	--	FJ685721.1	KF214214.1	--	--	--
<i>Pseudis bolbodactyla</i>	EF153007.1	EF153007.1	--	--	--	--	--	--	--	--	--	--
<i>Rhinella crucifer</i>	DQ158447	DQ158447	DQ415596	--	--	--	DQ158288	--	--	--	--	--
<i>Rhinella granulosa</i>	DQ158458.1	DQ158458.1	HM159225.1	FJ882728.1	--	FJ882673.1	DQ158299.1	DQ158380.1	--	--	--	--
<i>Rhinella hoogmoedi</i>	JN867545.1	JN867571.1	--	--	--	--	--	--	--	--	--	--
<i>Rhinella icterica</i>	DQ158462.1	DQ158462.1	JN594575.1	--	--	--	DQ158303.1	DQ158384.1	HM159240.1	--	--	--
<i>Rhinella schneideri</i>	DQ158480.1	DQ158480.1	HM159235.1	--	--	FJ882674.1	--	DQ158399.1	--	--	--	--
<i>Scinax crospedospilus</i>	AY819391.1	AY819523.1	--	--	--	--	AY819141.1	--	--	--	--	--
<i>Scinax fuscovarius</i>	AY843758.1	AY843758.1	AY844003.1	--	--	--	--	AY844519.1	AY844744.1	AY844943.1	--	AY844179.1
<i>Scinax hayii</i>	GQ896257.1	--	--	--	--	--	--	--	--	--	--	--
<i>Scinax similis</i>	GQ896263.1	--	--	--	--	--	--	--	--	--	--	--
<i>Scythrophrys sawayae</i>	DQ283099.1	DQ283099.1	--	--	DQ284149.1	--	--	--	DQ283815.1	--	--	DQ282926.1





**Figura S1.** Mapa mostrando a área de estudo no hotspot Floresta Atlântica brasileira (em cinza) no território sulamericano. Os pontos negros indicam as Áreas protegidas que foram amostradas como dados complementares. 1. Parque Ecológico Spitzkopf, SC; 2. Parque Estadual Pico do Marumbi, PR; 3. Parque estadual de Campinhos, PR; 4. Parque municipal das Perobas, PR; 5. Parque Estadual Mata dos Godoy, PR; 6. Refugio municipal da vida Silvestre, Horto Florestal de Jacarezinho, PR; 7. Estação Ecológica de Juréia-Itatins, SP; 8. Parque Estadual da Serra do Mar Núcleo Caraguatatuba, SP; 9. Parque Nacional da Serra dos Órgãos, RJ; 10. Reserva Biológica Augusto Ruschi, ES; 11. Reserva Biológica de Una, BA. De janeiro de 2014 (PA 8 de cinco dias), em julho de 2014 (PA 6 três dias), em novembro de 2014 (PAs 4, 6 três dias em cada), dezembro de 2014 (PA 4 dois dias), de janeiro-março de 2015 (PAs 1, 2, 7-11 sete dias em cada), Maio de 2015 (PA 6 três dias), em novembro de 2015 (PA 4 dois dias), dezembro de 2015 (PA 3 cinco dias), em fevereiro de 2016 (PAs 5, dois dias, três dias 6) , março de 2016 (PA 4 um dia, dois dias 5). Foram amostrados anfíbios utilizando inquéritos noturnos e diurnos acústicos e visuais (Crump & Scott Jr, 1994; Zimmerman, 1994). Foram pesquisados nas margens de corpos d'água, nos rios, e ao longo de 2000 transectos m florestais. As espécies encontradas foram testados seus mecanismos de defesa utilizado o método I proposto por

Lourenço-de-Moraes et al (2016). Todas as suas características funcionais foram registradas. As espécies coletadas foram eutanasiadas com hidrócloro de benzocaína > ou = 250mg/L(ASH, 2004). Nº da autorização/licença SISBIO: 30344.

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