

UNIVERSIDADE ESTADUAL DE MARINGÁ
CENTRO DE CIÊNCIAS DA SAÚDE
DEPARTAMENTO DE ANÁLISES CLÍNICAS E BIOMEDICINA
PROGRAMA DE PÓS-GRADUAÇÃO EM BIOCÊNCIAS E
FISIOPATOLOGIA

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Ação de sistemas de efluxo na resistência à isoniazida em *Mycobacterium
tuberculosis*

MARINGÁ

2018

JOÃO VÍTOR PEREZ DE SOUZA

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Dissertação apresentada ao Programa de Pós-Graduação em Biociências e Fisiopatologia do Departamento de Análises Clínicas e Biomedicina, Centro de Ciências da Saúde da Universidade Estadual de Maringá, como requisito parcial para obtenção do título de Mestre em Biociências e Fisiopatologia.

Área de concentração: Biociências e Fisiopatologia Aplicadas à Farmácia.

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Maringá

2018

Dados Internacionais de Catalogação na Publicação (CIP)
(Biblioteca Central - UEM, Maringá, PR, Brasil)

Souza, João Vítor Perez de
S729a Ação de sistemas de efluxo na resistência à isoniazida em
Mycobacterium tuberculosis / João Vítor Perez de Souza. -- Maringá, 2018.
62 f. : il. color.

Orientador: Prof^ª. Dr^ª. Rosilene Fressatti Cardoso.

Co-orientador: Prof^ª. Dr^ª. Katiany R. Caleffi Ferracioli.

Dissertação (mestrado) - Universidade Estadual de Maringá, Centro de Ciências da Saúde, Departamento de Ciências Básicas da Saúde, Programa de Pós-Graduação em Biociências e Fisiopatologia, 2018.

1. Tuberculose (*Mycobacterium tuberculosis*) -

Bombas de efluxo. 2. Tuberculose - Verapamil. I.

Cardoso, Rosilene Fressatti, orient. II. Ferracioli,

Katiany R. Caleffi, orient. III. Universidade Estadual de Maringá. Centro de Ciências da Saúde.

Departamento de Análises Clínicas e Biomedicina.

Programa de Pós-Graduação em Biociências e Fisiopatologia.

IV. Título.

CDD 23.ed. 616.995

FOLHA DE APROVAÇÃO

DEDICATÓRIA

Dedico esse trabalho para todas as pessoas que sofrem ou já sofreram de tuberculose. Espero que algum dia este trabalho possa contribuir de alguma forma para a vida dessas pessoas.

AGRADECIMENTOS

Em primeiro lugar aos meus pais, João e Renata, que forneceram a base para tudo que sou e serei na vida. Reconhecer em mim mesmo os traços, personalidade e caráter dos dois me faz muito feliz, uma vez que eles estarão sempre comigo, onde quer que eu esteja.

Aos familiares, que me mostraram exemplos a serem seguidos e valores a serem mantidos.

Às mentoras e mentores que me guiaram nessa jornada:

Todas as professoras do Laboratório de Bacteriologia Médica, por todos esses anos de orientação, atenção, carinho e paciência. Considero todas como amigas próximas e exemplos a serem seguidos pessoal e profissionalmente. Em especial às professoras Rosilene e Katiany, minhas mães científicas, por todo o conhecimento compartilhado e por acreditarem no meu potencial;

Às professoras Sandra Mara, Thaís Gomes e Maria Valdrinez pela orientação nos primeiros projetos de iniciação científica que participei na universidade. E por terem sido as que primeiro me direcionaram para a pesquisa;

Ao Paulo, Carla, dona Ivone e Marina do laboratório de imunologia do LEPAC por se mostrarem pacientes e interessados em ensinar conteúdos avançados para um aluno de primeiro ano durante meu estágio;

Aos professores Jorge Teixeira, Izabel Galhardo e Dennis Armando por me apresentarem o método científico, bem como o valor impar do sistema único de saúde brasileiro;

À toda equipe do Laboratório de Bacteriologia Médica do LEPAC, que são exemplos na prática da microbiologia. Graças ao carinho e atenção ao ensinar de todas as meninas, pude

aprender muito mais do que poderia esperar no breve período de estágio que passei no laboratório;

À professora Débora por mostrar como a ciência pode e deve ser compartilhada com a população para instigar, inspirar e plantar a semente da nova geração de cientistas. E juntamente a Luciane e professora Gessilda, agradeço pela paciência e disponibilidade para tirar dúvidas sobre a pós-graduação e o empenho em ajudar todos os alunos do PBF.

À minha namorada Letícia, por todo o amor, amizade, carinho e suporte. Não poderia por uma companheira melhor. Sinto que o nosso caminho será longo e cheio de obstáculos, porém repleto de frutos. Superaremos as dificuldades e colheremos os frutos um a um, estando lado a lado.

Ao grande amigo Jean por infinitas horas de conversas fiadas, alegrias, tristezas, comilanças e psicanálise. Além da grande amizade, agradeço pela parceria na pesquisa. Considerando os aspectos pessoais e profissionais, se algum dia eu me tornar um homem equivalente à metade do que ele é, estarei mais do que realizado.

Agradeço a todos os colegas da pesquisa do Laboratório de Bacteriologia Médica pela convivência e companheirismo do dia-a-dia.

Ter a oportunidade de realizar uma pós-graduação em uma universidade pública renomada é com certeza algo que serei grato para a vida toda. Assim, serei eternamente grato à Universidade Estadual de Maringá, ao Programa de Pós-graduação em Biociências e Fisiopatologia, ao Governo do Estado do Paraná, ao Governo Federal e aos órgãos de financiamento como o Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq), Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES) e Fundação Araucária (FA).

Por fim, agradeço imensamente ao carinho e confiança que todas as pessoas e entidades depositaram em mim nessa jornada até aqui, e espero algum dia retribuir tudo isso de alguma forma.

Ação de sistemas de efluxo na resistência à isoniazida em *Mycobacterium tuberculosis*

RESUMO

A tuberculose (TB) pulmonar se mantém como uma ameaça global devido à alta transmissão e prevalência de humanos infectados por bacilos do complexo *Mycobacterium tuberculosis*. Características específicas da patogenia da doença e da estrutura do bacilo fazem com que o tratamento efetivo seja dificultado. Dentre elas, podemos citar: i) a presença de uma parede celular espessa e rica em lipídeos e ácidos micólicos; ii) a produção de enzimas hidrolíticas; iii) mutações aleatórias nos genes que codificam para os alvos dos fármacos; iv) e sistemas de efluxo capazes de reduzir a concentração intracelular de antimicrobianos. As bombas de efluxo (BEs) micobacterianas já são aceitas como mecanismos secundários de resistência a fármacos, mas ainda não há correlação entre efluxo e resistência à isoniazida (INH). Com o objetivo de estudar as BEs, pesquisadores vêm empregando moléculas com propriedades de inibidoras de bomba de efluxo (IBE). Verapamil (VP) é um dos fármacos mais utilizados com essa finalidade, uma vez que a redução da atividade de efluxo por ele exercida aumenta a suscetibilidade de *M. tuberculosis* a alguns medicamentos anti-TB. Assim, o objetivo deste estudo foi avaliar o papel das BEs na resistência à INH, bem como o efeito combinatório de VP e INH em *M. tuberculosis*. Os resultados obtidos neste estudo estão apresentados no manuscrito “Efflux pump gene expression and isoniazid resistance in *Mycobacterium tuberculosis*”. Primeiramente foi determinada a concentração inibitória mínima (CIM) e o efeito combinatório dos fármacos INH e VP, através dos métodos *resazurin microtiter assay plate* (REMA) e *resazurin drugs combination microtiter assay* (REDCA). Em posse desses resultados, quatro bacilos com diferentes perfis de resistência foram selecionados para ensaios de expressão gênica de BEs. Cada *M. tuberculosis* foi exposto separadamente a 0,5 x CIM de INH e INH+VP por 48 h. Por meio da RT-qPCR, foi determinada a expressão relativa de 10 genes que codificam para BEs em *M. tuberculosis*. Foi observada redução significativa da CIM de INH após combinação com VP para isolados suscetíveis aos fármacos anti-TB. Resumidamente, os ensaios de expressão gênica revelaram padrões de resposta contra a pressão de INH que se correlacionaram com cada perfil de resistência, presença ou ausência de mutações em genes de resistência e efeito combinatório com VP.

Palavras-chave: Tuberculose, bombas de efluxo, isoniazida, verapamil.

Efflux pump gene expression and isoniazid resistance in *Mycobacterium tuberculosis*

ABSTRACT

Pulmonary tuberculosis (TB) remains a global threat due to the high transmission and prevalence of humans infected with bacilli of the *Mycobacterium tuberculosis* complex. Specific characteristics of the pathogenesis of the disease and the structure of the bacillus make effective treatment difficult. Among them, we can mention: i) the presence of a thick cell wall rich in lipids and mycolic acids; ii) the production of hydrolytic enzymes; iii) random mutations in the genes that code for drug targets; iv) and efflux systems capable of reducing the intracellular concentration of antimicrobials. Mycobacterial efflux pumps (EPs) are already accepted as secondary mechanisms of drug resistance, but we still lack strong correlation between efflux and resistance to isoniazid (INH). In order to study the EPs, researchers have been using molecules with properties of efflux pump inhibitors (EPI). Verapamil (VP) is one of the drugs most used for this purpose, since the reduction of the efflux activity exerted by it increases the susceptibility of *M. tuberculosis* to some anti-TB drugs. Thus, the objective of this study was to evaluate the role of EPs in resistance to INH, as well as the combinatorial effect of VP and INH in *M. tuberculosis*. The results obtained in this study are presented in the manuscript "Efflux pump gene expression and isoniazid resistance in *Mycobacterium tuberculosis*". First, the minimum inhibitory concentration (MIC) and the combinatorial effect of INH and VP were determined by resazurin microtiter assay plate (REMA) and resazurin drugs combination microtiter assay (REDCA) methods. With these results, four bacilli with different resistance profiles were selected for EPs gene expression assays. Each *M. tuberculosis* was exposed separately to 0.5 x MIC of INH and INH + VP for 48 h. Through RT-qPCR, the relative expression of 10 genes coding for EPs in *M. tuberculosis* was determined. Significant reduction of INH MIC was observed after combination with VP for isolates susceptible to anti-TB drugs. Briefly, gene expression assays revealed INH pressure response patterns that correlated with each resistance profile, presence or absence of mutations in resistance and combinatorial genes with VP.

Keywords: tuberculosis, efflux pumps, isoniazid, verapamil.

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Dissertação elaborada e formatada conforme as normas da ABNT, com referências e citações seguindo a norma Vancouver (Capítulo I e III). O manuscrito (Capítulo II) segue as normas do periódico *Tuberculosis*, Disponível em: <<https://www.elsevier.com/journals/tuberculosis/1472-9792/guide-for-authors>>

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CAPÍTULO I

1.1 INTRODUÇÃO

1.1.1 Tuberculose: epidemiologia, transmissão e tratamento.

A tuberculose (TB) é uma doença infectocontagiosa conhecida há séculos e em humanos é causada por bacilos do complexo *Mycobacterium tuberculosis*. Recentemente, a TB ultrapassou HIV/AIDS e se tornou a doença causada por um único agente infeccioso que mais mata no mundo, com cerca de 1.6 milhões de mortes em 2017 (1).

A transmissão ocorre, em sua maioria, por inalação de gotículas expelidas por indivíduos com a forma ativa da doença. Após a inalação, a infecção segue para a forma latente ou subclínica sendo que em uma pequena proporção dos casos, pode evoluir para a TB doença, sendo a manifestação pulmonar a mais comum (2). Cerca de 15% dos casos de TB manifestam-se fora do parênquima pulmonar, logo são denominados de TB extrapulmonar (TBEP) (2).

A manifestação crônica da doença é a mais comum, gerando diversas consequências na vida do portador, muitas vezes o impedindo de ter uma vida de qualidade e privando-o de certas interações sociais. Estima-se que a TB seja responsável por gastos de até 6 bilhões de dólares anualmente, sem incluir gastos com pesquisa e desenvolvimento (1). Se levado em conta que frequentemente a doença afeta o indivíduo em sua idade mais produtiva, os prejuízos são ainda maiores.

Estima-se que cerca de um terço da população mundial esteja infectada e um número ainda maior tenha pelo menos entrado em contato com o bacilo (2). O Brasil se encontra no grupo de 30 países que concentram 87% dos casos de TB mundialmente, sendo que as maiores preocupações no país são o grande número de casos e coinfeção com o vírus HIV (1).

O primeiro tratamento para TB data de 1942, com o descobrimento do fármaco estreptomicina (3). Desde então, pesquisadores e entidades governamentais propuseram diferentes esquemas de tratamento, porém baixos índices de sucesso terapêutico e o

surgimento de resistência aos fármacos empregados inviabilizaram várias das estratégias propostas.

Na tentativa de aumentar os índices de cura e evitar o surgimento de casos de resistência, a Organização Mundial da Saúde (OMS) propõe um esquema básico de tratamento para a TB, que perdura por seis meses: dois meses de tratamento intensivo com o emprego de isoniazida (INH), rifampicina (RIF), etambutol (EMB) e pirazinamida (PZA); seguidos por uma fase de manutenção com quatro meses de INH e RIF (1). No Brasil, o tratamento completo é disponibilizado gratuitamente pelo sistema único de saúde e todos os anos as diretrizes de tratamento são atualizadas pelo plano nacional de controle à TB (4).

Mesmo no cenário em que *M. tuberculosis* seja suscetível a fármacos, vários obstáculos dificultam o tratamento da TB e fazem com que o número de opções terapêuticas seja extremamente limitado. Dentre esses podemos citar: i) resistência intrínseca a certos antimicrobianos (por exemplo, beta-lactâmicos) devido à baixa permeabilidade de sua parede celular ou produção de enzimas hidrolíticas (5); ii) seu tempo de multiplicação elevado que muitas vezes inviabiliza fármacos que tem um período de meia-vida sistêmico curto; iii) possibilidade de ocorrência de estado de dormência bacteriana durante a infecção, com bacilos metabolicamente menos ativos e conseqüente diminuição da ação antimicrobiana (6); iv) e sistemas de efluxo são capazes de reduzir as concentrações intracelulares de compostos tóxicos, tornando a bactéria tolerante a diversos fármacos (7).

Além dos mecanismos citados anteriormente, *M. tuberculosis* pode desenvolver resistência adquirida durante um tratamento inapropriado. Brevemente, ao abandonar o tratamento antes da conclusão, bacilos resistentes podem ser selecionados e assim linhagens resistentes são estabelecidas. De modo geral, a resistência para os principais fármacos anti-TB se dá por erros na replicação do material genético e falhas nos sistemas de reparo de DNA (8). Mais especificamente, se essas mutações ocorrerem em sítios importantes para a função de um fármaco (ex. sítios catalíticos), esse bacilo se torna resistente (8).

A seleção de bacilos resistentes a qualquer um dos fármacos de primeira linha anti-TB reduz grandemente a eficácia do tratamento. Para INH, a resistência é particularmente importante, uma vez que este fármaco demonstra alta atividade bactericida contra bacilos em replicação ativa, reduzindo a carga bacilar pulmonar e as chances de transmissão em poucos dias (2).

Quando a resistência a INH é acompanhada de resistência a RIF, temos a denominação de resistência a múltiplos fármacos (MDR). Nestes casos, fármacos de segunda-linha têm que

ser empregados. Infelizmente, tais fármacos (ex. aminoglicosídeos) são mais tóxicos e frequentemente causam efeitos adversos, tornando o tratamento ainda mais difícil (9). Cerca de 160 mil novos casos de TB MDR foram reportados em 2017. Do total de casos de TB MDR, 87 % iniciaram o tratamento e apenas 55 % obtiveram cura (1).

Para *Mycobacterium tuberculosis* que são classificados como MDR e adicionalmente apresentam resistência a uma fluoroquinolona e a um medicamento injetável de segunda linha (amicacina, canamicina ou capreomicina) aplica-se a denominação de resistência extensiva a fármacos (XDR). Em situações de TB XDR, os índices de sucesso de tratamento caem para cerca de 30 % (1). Adicionalmente, já foram reportados casos de TB totalmente resistente a fármacos, na qual as chances de sucesso no tratamento são ainda menores (1,2).

1.1.2 Isoniazida: mecanismo de ação e resistência

A INH é um dos principais fármacos no esquema terapêutico contra a TB. Curiosamente, o primeiro uso da molécula de INH foi como um fármaco antidepressivo (10), sendo que sua atividade antimicobacteriana foi descrita posteriormente. Middlebrook e colaboradores (11) foram responsáveis pela primeira descrição de tal atividade, juntamente com a ocorrência de resistência à INH *in vitro*. Em 1970 e 1972 foi descrito que INH age inibindo a síntese de ácidos micólicos, um dos principais componentes da parede celular micobacteriana (12) e que essa inibição está correlacionada com a morte celular (13), respectivamente. Ainda antes da virada do século, os principais mecanismos de ação e resistência a INH foram elucidados (14–17). A Figura 1, adaptada de Islam et al. (2017), evidencia as principais etapas do descobrimento do fármaco INH, bem como seu mecanismo de ação (8).

Com base nessas e em outras publicações subsequentes, o mecanismo de ação de INH, bem como as principais causas da resistência foram entendidas. Resumidamente, INH precisa ser metabolizada pela enzima catalase-peroxidase micobacteriana (KatG) para iniciar sua ação no bacilo; passando do seu estado de pró-fármaco (ácido isonicotínico hidrazida) para fármaco propriamente dito (ácido isonicotínico). Logo, mutações no gene (*katG*) que codifica a enzima necessária para a ativação do fármaco, são as principais responsáveis pelo surgimento de resistência à INH. Mudanças no sítio ativo de KatG dificultam a acomodação da INH no sítio catalítico da enzima, levando a grande aumento na concentração inibitória mínima (CIM) de INH para *M. tuberculosis* (18).

Em seu estado ativo, a INH reage não enzimaticamente com o co-fator NAD^+ transformando-se em um aduto de ácido isonicotínico- NAD^+ (8). Após essa modificação, a molécula de INH tem alta afinidade pela enzima enoil-ACP redutase NADH dependente (InhA), codificada pelo gene *inhA*. Essa enzima faz parte da via de síntese de ácidos graxos (FASII) e é responsável por adicionar carbonos na molécula de ácido micólico “pré-maturo” (15), sendo que inibição desta impede a síntese dos ácidos micólicos e leva a morte do bacilo (13). Para este gene, mutações estruturais e no promotor já foram reportadas como causadoras de resistência a INH. Enquanto mutações estruturais são responsáveis pela diminuição da afinidade da enzima pelo aduto, mutações no promotor gênico causam superexpressão da enzima, assim a inibição que seria causada por uma concentração baixa de INH ativada, se torna insuficiente, sendo necessárias concentrações maiores para um bloqueio completo (19).

O mecanismo de ação da INH está diretamente ligado a enzimas pertencentes às vias de estresse oxidativo e síntese de ácidos graxos. Assim, mutações que alterem essas vias metabólicas podem afetar a susceptibilidade à INH com maior ou menor magnitude (20,21).

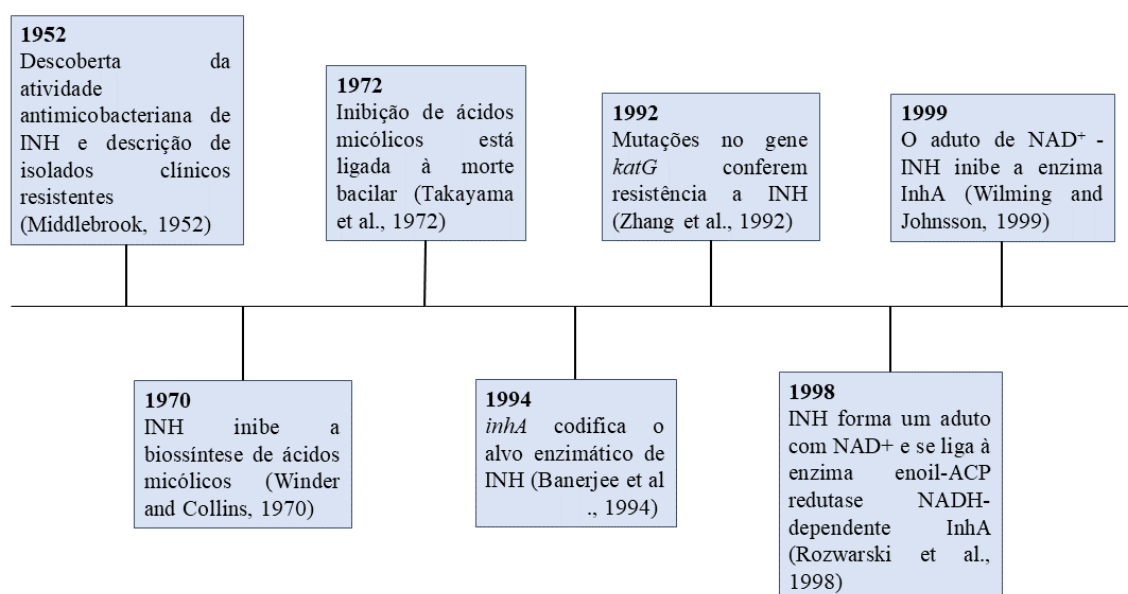


Figura 1. Histórico do descobrimento e elucidação dos principais mecanismos de resistência à isoniazida em *Mycobacterium tuberculosis*. Adaptada de Islam et al. (2017). INH, isoniazida; *inhA*, gene que codifica para a enzima enoil-ACP redutase NADH dependente; *katG*, gene que codifica para enzima catalase-peroxidase.

Apesar do avanço realizado no entendimento dos mecanismos moleculares de resistência em *M. tuberculosis*, muito ainda há a esclarecer. Em uma pequena parcela dos isolados clínicos, a resistência não pode ser explicada pela presença de mutações gênicas o

que leva a crer que mecanismos alternativos estão ativos. Dentre esses, destacam-se sistemas de efluxo bacterianos. Apesar de as bombas de efluxo (BEs) serem capazes de estrar uma grande variedade de compostos tóxicos ao microrganismo, é a sua habilidade de diminuir as concentrações intracelulares de fármacos na bactéria que gera maiores consequências no tratamento (22).

Desde 2003, publicações na literatura trazem dados de como os sistemas de efluxo micobacterianos impactam na resistência a fármacos (23,24) e atualmente o papel desses sistemas já é amplamente aceito (25–27). Além de colaborarem para a redução na susceptibilidade a um fármaco, comumente podem causar resistência cruzada a outros, uma vez que cada BE tem uma grande variedade de possíveis substratos (22).

Para estudar a fundo o papel de sistemas de efluxo na resistência micobacteriana, pesquisadores vem combinando moléculas capazes de inibir tais bombas com os antimicrobianos comumente utilizados na terapia anti-TB. Dentre os inibidores de bombas de efluxo (IBEs) mais utilizados, podemos citar verapamil (VP) por ser capaz de inibir BEs de diversas famílias (28) e ter características farmacológicas bem estabelecidas (29–31). Em humanos, VP é utilizado como um inibidor de canais de cálcio, para o tratamento de condições cardiovasculares, como arritmias e hipertensão arterial. Em mamíferos, já foi comprovado que VP inibe a glicoproteína P (P-gp), um transportador presente na maior parte do trato gastrointestinal, classificado na família *ATP-binding cassette* (ABC) que também é encontrada em micobactérias (7).

Proteínas das famílias *resistance nodulation division* (RND), *small multidrug resistance family* (SMR) e *major facilitator superfamily* (MFS) já foram relacionadas a algum grau de resistência a INH (32–34), porém o papel das BEs na resistência a este fármaco ainda é controverso, uma vez que a molécula do pró-fármaco INH é pequena, solúvel em água e sem carga iônica em pH fisiológico, características essas que garantem a difusão passiva por membranas (35).

1.1.3 Avaliação da susceptibilidade de *Mycobacterium tuberculosis* a fármacos

Avaliar os níveis de susceptibilidade a fármacos de *M. tuberculosis* fornece informações importantes que devem ser levadas em conta para o tratamento de um paciente com TB (36). Baseado na resposta indicada pelo teste realizado, o clínico pode optar por esquemas terapêuticos que sejam mais eficazes ou que apresentem maior possibilidade de cura.

Devido às características peculiares de *M. tuberculosis*, métodos convencionais de determinação de susceptibilidade *in vitro* apresentam algumas limitações. Dentre essas características, o tempo de crescimento elevado de bactérias do complexo *M. tuberculosis* e o alto nível de biossegurança necessário para manipular amostras clínicas possivelmente positivas e culturas desse microrganismo são as que se apresentam como os maiores obstáculos (37).

O método mais utilizado em centros de referência é o equipamento BD BACTET MGIT (Becton e Dickinson). O sistema baseia-se no método das proporções, ou seja, possui concentrações fixas dos fármacos de primeira linha anti-TB. Caso haja detecção de crescimento bacteriano no tubo contendo o fármaco em questão, o isolado clínico é considerado resistente [38].

O equipamento possui sensores que detectam pequenas mudanças na concentração de oxigênio presente na cultura a cada 60 minutos, assim possibilitando maior sensibilidade na detecção de crescimento. Apesar de prover agilidade, sensibilidade e confiança no diagnóstico de resistência micobacteriana, o sistema BD BACTEC MGIT tem limitações importantes como alto custo de implementação e falta de resultados quantitativos de susceptibilidade que indiquem qual é a concentração inibitória mínima (CIM) do fármaco em questão (37). Desta forma, pesquisadores lançam mão de métodos que sejam ao mesmo tempo eficientes e com melhor custo-benefício.

1.1.3.1 Resazurin Microtiter Assay Plate (REMA)

Dentre os métodos mais utilizados por pesquisadores para a avaliação de susceptibilidade a fármacos, destaca-se o método REMA, proposto por Palomino et al. (2002) (38). Brevemente, o método trata-se de uma microdiluição de fármacos em meio de cultura Middlebrook 7H9, com posterior adição de cultura micobacteriana padronizada na escala de turbidez de McFarland 1.0 e diluída 1:20 em Middlebrook 7H9. Por ser realizado em microplacas de 96 orifícios, uma variedade de fármacos pode ser testada, garantindo grande versatilidade ao ensaio.

Após término do preparo da placa, a preparação é incubada por sete dias em estufa em temperatura de 35 ± 2 °C, à atmosfera normal. Decorrido o tempo de incubação, adiciona-se 30 µL de uma solução de 0,01 % de resazurina. Após 24 h de incubação nas mesmas condições, faz-se a leitura da microplaca. A mudança visual da coloração azul para rosa,

indica redução da resazurina e, portanto, crescimento bacteriano. A CIM é determinada como o último orifício que não demonstrou viragem da cor.

O método de REMA apresenta-se como uma alternativa interessante para o pesquisador, uma vez que não são necessários equipamentos específicos além dos já disponíveis em laboratórios que trabalham com bactérias do complexo *M. tuberculosis* (38).

Adicionalmente, a característica de microdiluição permite a avaliação quantitativa da susceptibilidade dos *M. tuberculosis* testados. Assim, possibilitando a classificação dos níveis de resistência de forma acurada.

1.1.4 Avaliação do efeito combinatório entre fármacos anti-TB

Tendo em vista o regime poliquimioterápico utilizado para o tratamento da TB, o estudo sobre a interação entre fármacos é de suma importância para a proposta de novas estratégias. Diversas metodologias podem ser utilizadas para estudar a interação entre fármacos, desde análises de atividade anti-micobacteriana *in vitro*, até ensaios *in vivo* com modelos de infecção em animais de laboratório (39).

Dentre as metodologias disponíveis *in vitro*, podemos destacar o método *rezasurin drugs combination microtiter assay* (REDCA), padronizado por Caleffi-Ferracioli et al. (2013) (40). Brevemente, o método tem preparo similar ao REMA mencionado anteriormente, porém permite a diluição simultânea de dois fármacos. No eixo x da microplaca de 96 orifícios, é adicionado fármaco A, que é diluído verticalmente em série; após, é adicionado no eixo y o fármaco B, que é então diluído horizontalmente. As condições de incubação, revelação e leitura do ensaio são as mesmas do método REMA (40).

Dependendo do tipo de combinação utilizado, pode-se aplicar diferentes cálculos para interpretar os efeitos combinatórios entre os fármacos. Se realizada combinação entre dois antimicrobianos, pode-se calcular índices como o FICI, do inglês índice de concentração inibitória fracional, por meio da equação “ $FICI = CIM_{(\text{fármaco A combinado ao B})} / CIM_{(\text{fármaco A})} + CIM_{(\text{fármaco B combinado à A})} / CIM_{(\text{fármaco B})}$ ”. A interpretação desse índice revela se a interação entre os fármacos foi sinérgica ($FICI \leq 0,5$); indiferente ($0,5 > FICI \leq 4$); e antagônica ($FICI > 4$) (41).

Alternativamente, quando os testes são feitos para avaliar a interação entre um antimicrobiano e um candidato a fármaco (ex. inibidor de bomba de efluxo), o uso do FIC, do inglês concentração inibitória fracional é mais adequado. Diferentemente do FICI, o FIC considera apenas o efeito do candidato sobre o antimicrobiano em questão. A caracterização

da interação entre os fármacos se dá como sinérgica ($FIC \leq 0,25$); indiferente ($0,25 > FIC \leq 2$); e antagônica ($FIC > 2$), de acordo o resultado obtido na equação “ $FIC = CIM_{(fármaco A)} / CIM_{(Fármaco A combinado ao B)}$ ”(42).

De modo similar ao FIC, pode-se calcular o fator modulador (FM) através da equação “ $FM = CIM_{(Fármaco A combinado ao B)} / CIM_{(fármaco A)}$ ”. Comparado aos outros índices, FM tem a vantagem de evidenciar diretamente qual foi o efeito combinatório que o fármaco B exerceu no fármaco A. Por exemplo, um valor de FM de oito, significa que a combinação foi capaz de reduzir a CIM do fármaco A em oito vezes (42).

1.2 JUSTIFICATIVA

O desenvolvimento de resistência a qualquer um dos fármacos de primeira linha contra a TB dificulta muito o tratamento da doença, podendo aumentar o tempo de tratamento para 24 meses ou mais. Apesar de os mecanismos mais clássicos de resistência em *M. tuberculosis* já estarem bem caracterizados, bombas de efluxo também têm mostrado papel importante neste fenômeno. Graças a tais sistemas de efluxo, a bactéria é capaz de resistir à pressão inicial do tratamento até surgirem mutações espontâneas no genoma que acarretam em reduzida susceptibilidade.

Considerando o impacto da resistência micobacteriana no tratamento da TB, o estudo destes mecanismos de resistência secundários são de suma importância para o controle da doença, bem como o desenvolvimento de novas estratégias que possam ser efetivas no enfretamento da TB.

1.3 OBJETIVOS

1.3.1 Geral

Avaliar o papel de bombas de efluxo na resistência ao fármaco isoniazida de *M. tuberculosis*.

1.3.2 Objetivos específicos

- Determinar a concentração inibitória mínima (CIM) dos fármacos isoniazida (INH) e verapamil (VP) para *M. tuberculosis*, através do método *Resazurin microtiter assay plate* (REMA);
- Avaliar o efeito combinatório entre INH e VP em *M. tuberculosis*, por meio do método *resazurin drugs combination microtiter assay* (REDCA);
- Selecionar *M. tuberculosis* com diferentes CIMs e efeitos combinatórios de INH e VP para estudos de expressão gênica de bombas de efluxo, de acordo com os resultados dos métodos anteriores;
- Analisar a expressão relativa de 10 genes que codificam bombas de efluxo nos *M. tuberculosis* selecionados.

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CAPÍTULO II

Efflux pump expression and isoniazid resistance in *Mycobacterium tuberculosis*

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Abstract

Mycobacterial efflux is now widely accepted as a secondary mechanism of resistance to anti-TB drugs, but much is unclear about the relation between efflux and INH resistance. Thus, the aim of this study was to evaluate the role of efflux pumps (EPs) in the INH resistance, as well as the combinatory effect of INH and VP in *M. tuberculosis*. Minimal inhibitory concentrations and combinatory effects of INH+VP were accessed through resazurin microtiter assay plate (REMA) and resazurin drugs combination microtiter assay (REDCA), respectively. From the results of REMA and REDCA, we selected four bacilli with different susceptibility profiles and assessed their expression of 10 EPs genes through RT-qPCR after exposure to INH and INH+VP for 48 h. Significant reduction of INH MIC upon combination with VP was observed for INH susceptible isolates. Briefly, gene expression assays revealed responses against INH pressure that correlated with each resistance profile, presence or absence of *katG* mutations and combinatory effect with VP. Although the blockage of EPs does not impart in increased susceptibility for INH in bacteria with *katG* mutations, the conjunct administration of an EPI could cause important results for isolates with a low degree of INH resistance and act in the prevention of resistance in susceptible bacilli.

Keywords: Drug resistance, efflux pump inhibitors, verapamil, efflux pump, isoniazid, tuberculosis.

1. Introduction

1 Despite the existence of standard therapeutic against susceptible *Mycobacterium*
2 *tuberculosis*, first line anti-tuberculosis (TB) drugs such as isoniazid (INH) and rifampicin (RIF) are
3 ineffective to treat patients infected by multidrug resistant (MDR) bacilli. Unfortunately, second
4 line drugs (e.g. aminoglycosides) happen to be more toxic and often cause adverse effects, making
5 the back-up treatment even harder [1].

6 INH is one of the most important drugs in TB's treatment and usually is the first drug
7 resistance to emerge [2]. The development of resistance for INH is influenced by mutations in
8 several genes [3,4]. Of these, the two most common are in *katG*, which codifies the catalase-
9 peroxidase responsible for the activation of INH, and *inhA*, the target of the activated drug [3].
10 Although there are clear evidences that mutations on the *M. tuberculosis* genome are the main
11 responsible for INH resistance, efflux pump systems seem to also be involved [4,5].

12 During stress caused by a drug, the bacilli increases the expression of genes that encode
13 membrane transport proteins, as known as efflux pumps (EPs) in an attempt to protect itself [6]. In
14 addition, several EPs have a multi-substrate (multidrug EPs) specificity and are able to extrude a
15 wide variety of compounds [5].

16 Although there is clear evidence that EPs play important roles in resistance to many anti-TB
17 drugs [5], the impact of these efflux systems in INH resistance is not yet fully understood. The INH
18 pro-drug is a very small molecule, that has good dissolubility in water and is non-ionized in
19 physiological pH. These characteristics render great membrane diffusibility to INH through cell
20 membranes [7].

21 Verapamil (VP) is one of the molecules used to test EP activity, due to its properties as an
22 efflux pump inhibitor (EPI) [8]. In addition, VP is also been studied as an important adjuvant for
23 TB therapy, since the reduce of efflux activity increases the susceptibility of *M. tuberculosis* to
24 many anti-TB drugs [9].

25 Taking into consideration the limited number of drugs available to treat TB, the high burden
26 of the disease in the world and the fast emergence of resistance, the development of new strategies
27 to deal with TB is a primal need. Thus, the aim of this study was to evaluate the role of EPs in the
28 INH resistance, as well as the effect of VP in combination to INH in *M. tuberculosis* isolates
29 harboring different resistance profiles.

33 2. Material and methods

34

35 2.1 Bacterial strains and growth conditions

36

37 Fifteen *M. tuberculosis* isolates and the H₃₇Rv (ATCC 27294) reference strain belonging to
38 the Laboratory of Medical Bacteriology of the State University of Maringa, Brazil were selected
39 based upon previous detection of INH resistance by the automated method BD BACTEC
40 TMMGITTM960 (BD, Franklin Lakes – NJ, USA).

41 The clinical isolates selected had already been previously characterized by MIRU-VNTR,
42 Spoligotyping and tested by the commercial kit GenoType MTBDRplus® (Hain Lifescience
43 GmbH, Nehren, Germany) to detect mutations related to INH and RIF resistance [10]. All data of
44 the previous characterization is available in Table 1.

45 Bacilli were grown in Middlebrook 7H9 medium (Difco Laboratories, Detroit, MI, USA),
46 added with 0.2% glycerol (vol/vol), 0.025% tween 80 (vol/ vol) and supplemented with OADC
47 (Oleic acid, albumin, dextrose and catalase) (BBL/Becton-Dickinson, Sparks, MD, USA) for 15
48 days at 35 ± 2 °C.

49

50 2.2 Drugs

51 Isoniazid and verapamil were purchased from Sigma-Aldrich Co. (St. Louis, MO, USA).
52 The stock solutions were prepared following the manufacturer's instructions and further dilutions
53 were prepared in OADC-supplemented Middlebrook 7H9.

54

55 2.3 Minimal inhibitory concentration (MIC) and combinatory assays

56 The MIC of each drug was determined by the means of Resazurin Microtiter Assay Plate
57 (REMA) as described by Palomino et al. [11].

58 The combinatory effect of INH+ VP was determined by resazurin drugs combination
59 microtiter assay (REDCA), as previously described by Calleffi-Ferracioli et al.[12]. Combinations
60 of INH+VP were applied to all *M. tuberculosis* tested. To evaluate the combinatory effect of the
61 association, the modulation factor (MF) was calculated by the means of: $MF = MIC_{(INH)} /$
62 $MIC_{(INH+VP)}$. MF reflects the MIC reduction of a given antimicrobial when combined to an EPI. A
63 MF value ≥ 4 (four fold reduction) was considered significant [13].

64

65 2.4 Efflux pump gene expression

66 Four *M. tuberculosis* were selected to access gene expression: H37Rv (reference strain); BRF
67 47 (susceptible isolate); BRF 14 (INH monoresistant isolate, harboring S315T *katG* mutation); and
68 3614, an MDR isolate resistant to INH, RIF and EMB (harboring c-15t and I21T mutations in the
69 *inhA* gene) (Table 1). The bacilli were exposed to 0.5×MIC of INH or INH+VP for 48 h at 35 ± 2
70 °C. Additionally, pure bacterial culture was used as a control sample. RNA was extracted and
71 purified using RNeasy Plus Mini Kit (Qiagen Biotechnology, Valencia, CA, USA) and quantified in
72 Qubit 2.0 fluorimeter (Invitrogen, Carlsbad, CA, USA). DNA contaminants were removed with
73 RNase-free DNase I (Invitrogen, Carlsbad, CA, USA). The cDNA was synthesized using random
74 primers of total RNA and SuperScript® III Reverse Transcriptase (Invitrogen, Carlsbad, CA, USA).
75 The qPCR was performed using Fast SYBR Green PCR Master Mix (Applied Biosystems, Foster
76 City, CA, USA) with specific primers for the EPs studied (Table 2) in the StepOne Real-Time PCR
77 Systems (ThermoFisher Scientific, Waltham, MA, USA). Melting curves and negative controls were
78 performed in all reactions. The 16S RNA gene (*rrs*) was used to normalize all reactions and samples
79 were tested in triplicate. Relative quantification of target gene expression was calculated by the
80 means of $2^{-\Delta\Delta CT}$ [14] method and rates above two (>2) and lower than 0.5 (<0.5) were considered as
81 significantly up-regulated and down-regulated, respectively.

82

83 3. Results and Discussion

84 The major resistance mechanism to INH is due to spontaneous mutations on genes
85 responsible either for the activation or which are the target of this drug [3]. Although that is widely
86 known, alternative resistance mechanisms such as EP extrusion seem to also take part in reduced
87 susceptibility for this drug. Thus, we sought to evaluate the role of EPs in INH resistance as well as
88 the effect of the EPI VP in combination to INH in EP genes expression of *M. tuberculosis*. To our
89 knowledge, this is the first study that assayed 10 EP genes in *M. tuberculosis* harboring different
90 resistance profiles exposed to INH and INH+VP.

91 From the 15 clinical isolates tested, 12 (80%) were monoresistant to INH, two (13.33%)
92 susceptible to first-line anti-TB drugs and one MDR (6.67%). Minimal inhibitory concentrations of
93 VP and INH for the reference strain and the susceptible clinical isolates, ranged from 125 to 250
94 µg/mL and 0.06 to 0.125 µg/mL, respectively. For the INH resistant and MDR clinical isolates,
95 MICs of VP and INH had a range from 62.5 to 250 µg/mL and 4 to 32 µg/mL, respectively (Table
96 3).

97 From these results, we tested INH and VP combined to access if the blockade of EPs would
98 impart in increased susceptibility for INH (Table 3). Both susceptible bacilli tested (BRF 35 and 47)
99 presented great reductions (MF= eight and 16) of INH MIC upon combination to VP, while the

100 reference strain did not (MF = 2). Regarding the INH monoresistant isolates, all of them (12) did
101 not present great reductions of INH MIC upon combination with VP (MF= 1 or 2). One possible
102 explanation for these results is the presence of *katG* (S315T) mutations (Table 1), which greatly
103 hinder the access of INH to the catalytic site of the catalase-peroxidase enzyme [15]. In these
104 isolates, INH is not metabolized to its active form (or at least less metabolized) and so will not
105 cause damage to the bacteria. In this sense, the reduction of EP activity by VP would not act in a
106 synergic way with INH. Similarly, the MDR isolate (3614) tested did not show significant reduction
107 of INH MIC (*inhA* mutations). Differently from our results, a previous study revealed great
108 modulatory effects of INH+VP in MDR and XDR isolates harboring similar *inhA* mutations [9].

109 Although at first look our modulatory results seem discouraging, the great reduction of INH
110 MIC when combined to VP observed for the susceptible isolates provide interesting potential for
111 future therapy. As it is now consensus [6,16], efflux systems act as one of the first defense
112 mechanisms for bacteria to survive the early pressure of antibiotics. Additionally, these pumps act
113 as the “middleman” until a random mutation in resistance genes appears and impart in a more
114 definitive resistance phenotype for *M. tuberculosis* [16]. In this sense, the combination of INH+VP
115 could aid in the initial clearance process of bacilli by INH and reduce the persistence of bacteria
116 that one day might acquire *de novo* genetic resistance.

117 In our study, only susceptible isolates showed significant reductions of INH MIC upon
118 combination to VP. Differently from our findings, Machado et al. [17] observed that after *in vitro*
119 induction of resistance to INH, the conjunct administration of INH and VP reduced INH MIC at the
120 same level of the completely susceptible predecessor strain. Agreeing with our results, no
121 significant MIC reductions was observed in isolates who acquired *katG* mutations during the
122 induction process.

123 In order to evaluate how the EPs of *M. tuberculosis* respond to INH pressure, we selected
124 four *M. tuberculosis* with different susceptibility profiles to access EP gene expression (Table 3).
125 For H₃₇Rv, an overall low response in gene expression was observed for the majority of exposures
126 and genes studied (Figure 1), in comparison with the control. Exposure INH caused no significant
127 overexpression and, in fact, three genes (*Rv1410c*, *Rv1458c* and *Rv1819c*) were found
128 downregulated. Our results agree with a previous study that showed no significant overexpression
129 of EPs genes for the H₃₇Rv strain, upon exposure to INH [4]. Considering the origin and the genetic
130 background of the reference strain, this pattern could be explained by its pan-susceptibility and lack
131 of some virulence genes [18]. Seemingly, the combination with VP was able to improve the
132 pressure of INH on the strain. This is exemplified by two genes (*Rv2942* and *Rv3065*) that did not
133 present differential expression when exposed to INH alone. Similar results were also found for
134 these genes in the other isolates tested (Figures 1-4). From the cited genes, *Rv2942* has been related

135 to maintenance of virulence factors, lipid transport and efflux of INH in *M. tuberculosis* [17,19,20]
136 and *M. smegmatis* [21]; and *Rv3065* has been associated with resistance to INH and RIF [16,22].

137 Although H₃₇Rv (Figure 1) and BRF 47 (Figure 2) are both susceptible to first-line anti-TB
138 drugs, the gene expression profiles observed were quite different. Here, INH exposure caused
139 overexpression of seven genes versus none for the reference strain. All of them (*Rv2942*, *Rv2846*,
140 *Rv3065*, *Rv1258*, *Rv1458*, *Rv1218* and *Rv1819*) have been related to some degree of INH resistance
141 and/or found overexpressed in resistant isolates [16,23–27]. A previous report [4] has pointed out
142 that *Rv2846*, one of such genes, is overexpressed in INH exposure mainly in susceptible isolates.
143 This information agrees with our data, in which BRF 47 (Figure 2) isolate presented overexpression
144 of this gene, whereas the mono-resistant one (BRF 14) did not (Figure 3). The combination of the
145 anti-TB drug + EPI caused a noticeable increase in the expression rates for the majority of genes
146 tested. This overall increase in the gene expression rates of INH+VP when compared to INH alone,
147 agrees with the results from REDCA, in which the combination of drugs was able to reduce INH
148 MIC by as much as 16-fold for BRF 47 isolate (Table 3).

149 For the BRF 14 isolate, RT-qPCR revealed an overall low response in EP gene expression
150 upon exposure to INH alone and in combination to VP (Figure 3). As discussed earlier, the S315T
151 mutation in *katG* greatly hinders the activation of INH by the catalase enzyme (KatG), thus the drug
152 probably will not cause any stress to the bacteria. In spite of limited to four genes (*Rv3065*,
153 *Rv1456c*, *Rv1458c* and *Rv1218c*), the combination of INH+VP was able to improve the pressure of
154 INH in this isolate. The limited action of the combination agrees with the lack of modulation seen in
155 REDCA for this bacillus (Table 3). Unfortunately, BRF 14 did not show quantifiable levels of
156 *Rv2942*, so its response remains unsure.

157 Although VP was not able to reduce INH MIC in a significant manner for the 3614 isolate
158 (MF=2), the gene expression assays revealed patterns different from the other resistant organism.
159 Here, exposure to INH caused overexpression of all genes studied (Figure 4) despite the fact that
160 this isolate has c-15t and I21T mutations in the *inhA* gene. Mutations in the promoter region of *inhA*
161 (e.g. c-15t) increase the expression of the NADH-dependent enoyl-ACP reductase (InhA), the target
162 of activated INH, up to 20-fold, rendering the inhibitory activity of INH insufficient [28].
163 Differently from mutations in the *katG* gene that mostly nullify INH action, here the drug is still
164 able to cause damage. In addition, it has been proposed that activated INH has secondary targets
165 besides InhA [29]. Regarding the combination with VP, there was apparently no improvement for
166 this isolate with the exception of *Rv3065*. In fact, most genes demonstrated a decrease in expression
167 rates compared to INH alone.

168 Of all genes studied, *Rv1410c* stands out since it is associated with the transport of INH,
169 lipids and important virulence factors from the cytosol to the cell wall of *M. tuberculosis* [30]. Of

170 note, this gene was found downregulated or without significant change for H₃₇Rv, BRF 47 and BRF
171 14, while upregulated for 3614 isolate under INH exposure. Agreeing with our results, previous
172 reports [4,31] have shown that *Rv1410c* is found overexpressed in MDR/XDR isolates and without
173 significant change for H₃₇Rv and susceptible isolates exposed to INH. Additionally, the operon
174 formed *Rv1410* and *Rv1411* have been proposed as a possible target for drug development, since
175 deletion of this operon caused great attenuation of *M. tuberculosis* growth inside macrophages
176 [32,33].

177 Although limited to one microorganism for each resistance profile, comparing each gene
178 expression pattern provides us some data regarding the evolution of each bacilli as well as the
179 emergence of resistance to INH. Taking a closer look to isolates BRF 47 and BRF 14 (Figures 2 and
180 3, respectively) a striking difference in the expression rates upon INH exposure can be noted.
181 Possibly, BRF 47 is in an evolutionary stage that precedes the emergence of a mutation related to
182 resistance and so relies solely in the expression of EPs to tolerate the damage caused by INH.
183 Following this line of thought, an inappropriate treatment has led to the selection of BRF 14, an
184 INH resistant *M. tuberculosis* with a mutation in the *katG* gene. Although BRF 14 went through
185 similar pressures and is able to express EPs in defense, now INH will not be activated and will not
186 cause damage. Previous report by Machado et al exemplify this line of thought showing how
187 exposure to INH maintained a constant high-level EP gene expression in stages that preceded the
188 appearance of mutations in *katG* [17].

189 **4. Conclusions**

190 To this date, many papers have shown that efflux pumps play pivotal roles in resistance to
191 many anti-TB drugs. For isoniazid, these systems allow that susceptible bacillus survive the initial
192 pressure of the drug until a mutation in resistance genes randomly appears. Although the blockage
193 of EPs does not impart in increased susceptibility for bacteria with *katG* mutations, the conjunct
194 administration of an EPI could cause important results for isolates harboring more permissive
195 mutations (e.g. *inhA*) and act in the prevention of resistance in susceptible bacilli.

197 **Financial disclosure**

198 This study was supported by the National Council of Technological and Scientific Development
199 (CNPq) (grant 2016/14957-5). The funding agency did not participate in study design, data
200 collection and analysis, decision to publish or preparation of the manuscript.

201 **Acknowledgements**

202 The authors are thankful to the staff of the Medical Bacteriology Laboratory from State University
203 of Maringa - Parana, Brazil, Coordination of Superior Education Personnel Improvement (CAPES)

204 and National Council of Technological and Scientific Development (CNPq) for grant 2016/14957-
205 5.

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319 Expression in a Clinical Isolate *Mycobacterium tuberculosis* by Real-Time Reverse
320 Transcription PCR. *Microb Drug Resist* 2008;14:7–11. doi:10.1089/mdr.2008.0772.

321 **Table 1.** Molecular characterization of the *Mycobacterium tuberculosis* selected by resistance-related mutation detection by GenoType MTBDRplus®,
 322 12/24 *loci* mycobacterial interspersed repetitive unit (MIRU) and Spoligotyping.

Strain/Clinical isolate	Susceptibility profile	12/24 <i>loci</i> MIRU	Spoligotyping	Mutations			Reference
				<i>katG</i>	<i>inhA</i>	<i>rpoB</i>	
H ₃₇ Rv	S	-	777777477760771	WT	WT	WT	[34]
BRF 04	INH ^R	124326143324224234434132	776177400000171	S315T	WT	WT	[10]
BRF 14	INH ^R	223326143324224234414132	777777607760731	S315T	WT	WT	[10]
BRF 16	INH ^R	223326143324224234414132	777777607760731	S315T	WT	WT	[10]
BRF 23	INH ^R	125326143323224234324132	777777607760731	S315T	WT	WT	[10]
BRF 26	INH ^R	224326143324224234314132	777777607760731	S315T	WT	WT	[10]
BRF 35	S	224326143226224283334132	777737606560771	WT	WT	WT	[10]
BRF 47	S	225215163323323474324333	777777200000771	WT	WT	WT	[10]
BRF 57	INH ^R	124326143324224234334132	777777607760731	S315T	WT	WT	[10]
BRF 70	INH ^R	124326133323224264434232	777777607760731	S315T	WT	WT	[10]
BRF 75	INH ^R	124326143424224214214132	777777607760731	S315T	WT	WT	[10]
BRF 81	INH ^R	125326153122224263332132	777637607400031	S315T	WT	WT	[10]
BRF 84	INH ^R	125326153122224263332132	777637607400031	S315T	WT	WT	[10]
BRF 100	INH ^R	124326143324224234333132	777777607760731	S315T	WT	WT	[10]
BRF 101	INH ^R	124326143324224234333132	777777607760731	S315T	WT	WT	[10]
3614	INH ^R /RIF ^R /EMB ^R	224225163321	677737607769771	WT	c-15t I21T	S531L	[34]

323 VP: verapamil; INH: isoniazid; RIF: rifampicin; MF: modulation factor; S: susceptible to first-line anti-TB drugs; ^R: resistant to the indicated drug; WT: wild type; -: not performed.

Table 2. Primers used to assess relative quantification of the selected efflux pump gene in *Mycobacterium tuberculosis* by RT-qPCR

Efflux pump gene	Transporter family	Sequences (5' - 3')	Amplicon size (bp)	Reference
<i>Rv2942</i>	RND	Fw-TACCCAAGGTGGAAACAA Rv-CGTCAGAATAGAGGAACCAG	214	[25]
<i>Rv1410c</i>	MFS	Fw-AGTGGGAAATAAGCCAGTAA Rv-TGGTTGATGTCGAGCTGT	198	[25]
<i>Rv2846</i>	MFS	Fw-ATGGTAATGCCTGACATCC Rv-CTACGGGAAACCAACAAAG	131	[25]
<i>Rv3065</i>	SMR	Fw-AACCAGCCTGCTCAAAG Rv-CAACCACCTTCATCACAGA	221	[25]
<i>Rv1258c</i>	MFS	Fw-AGTTATAGATCGGCTGGATG Rv-GTGCTGTTCCCGAAATAC	268	[25]
<i>Rv1456c</i>	ABC	Fw-GAGTCGCACCAGAATCGC Rv-TCGCTGTTGGTTGCCTAC	90	[23]
<i>Rv1457c</i>	ABC	Fw-GTAGCACCGAGTCGTTTG Rv-ATCTCCACCGCATTACC	80	[23]
<i>Rv1458c</i>	ABC	Fw-CAGTCCAAGTACCTCAATG Rv-GCGATACGGGTCAATAAC	163	[23]
<i>Rv1218c</i>	ABC	Fw- CCGCAAGGCGTCTAGTGAA Rv- TGGACCCGTTGATGGAAAA	173	[35]
<i>Rv1819c</i>	ABC	Fw- CGGTGATTTCTTTCACAGC Rv- CCGACAGATTCCATCCATT	351	[35]
<i>16s RNA</i>		Fw- CAAGGCTAAAACCTCAAAGGA Rv- GGACTIONAACCAACATCTCA	197	[25]

Fw, sense primer; Rv, antisense primer; bp, base pairs; MFS, major facilitator superfamily; SMR, small multidrug resistance family; ABC, ATP-binding cassette; RND, resistance-nodulation-cell division superfamily

Table 3. Minimal inhibitory concentration of verapamil and isoniazid and modulation factor of the tested combinations in *Mycobacterium tuberculosis*.

Strain/Clinical isolate	Susceptibility profile	MIC ($\mu\text{g/mL}$)			MF
		VP	INH	INH+VP	
H ₃₇ Rv*	S	125	0.06	0.03	2
BRF 35	S	250	0.06	0.0078	8
BRF 47*	S	250	0.125	0.0078	16
BRF 04	INH ^R	125	4	2	2
BRF 14*	INH ^R	125	32	16	2
BRF 16	INH ^R	125	8	4	2
BRF 23	INH ^R	125	8	8	1
BRF 26	INH ^R	125	16	16	1
BRF 57	INH ^R	125	8	4	2
BRF 70	INH ^R	250	8	8	1
BRF 75	INH ^R	250	16	8	2
BRF 81	INH ^R	125	8	4	2
BRF 84	INH ^R	250	8	4	2
BRF 100	INH ^R	125	32	16	2
BRF 101	INH ^R	125	32	16	2
3614*	INH ^R /RIF ^R /EMB ^R	62.5	6.25	3.125	2

VP: verapamil; INH: isoniazid; RIF: rifampicin; MF: modulation factor; *: isolates selected for gene expression assays; S: susceptible to first-line anti-TB drugs; ^R: resistance to the indicated drug; Results in bold indicate synergism.

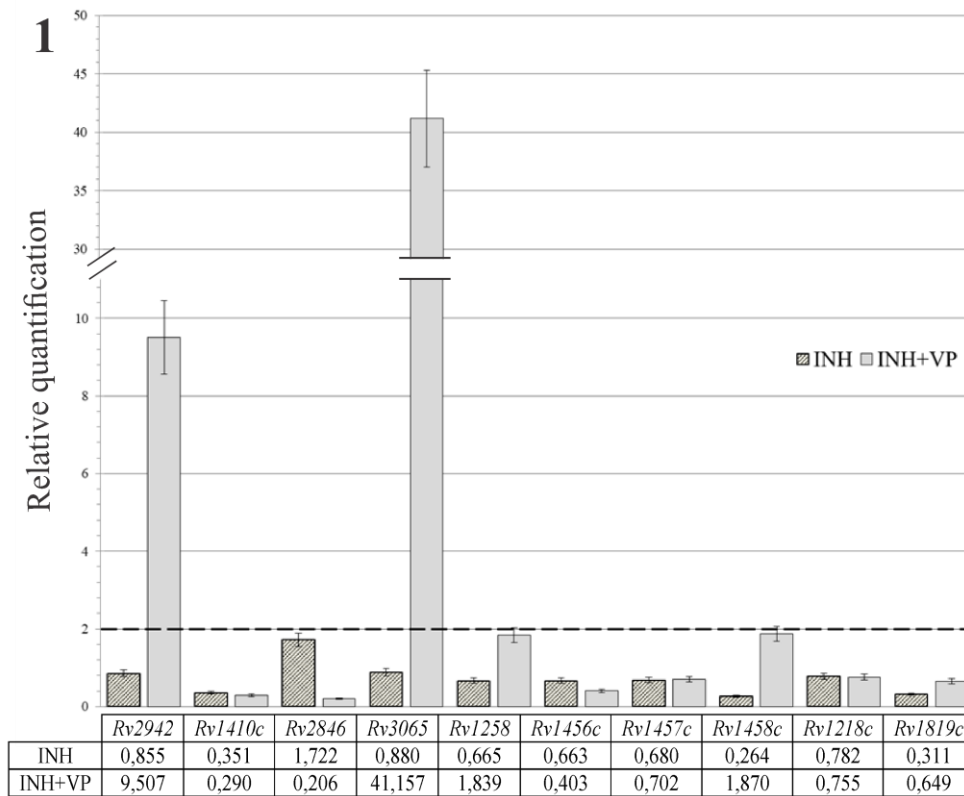


Figure 1. Relative quantification of 10 efflux pump genes in *Mycobacterium tuberculosis* H₃₇R_v, accessed by RT-qPCR after 48 h exposure to 0.5xMIC of isoniazid (INH) and the combination INH+VP. Relative quantification was determined by the means of $2^{-\Delta\Delta CT}$. Relative quantification rates above two (> 2) and lower than 0.5 (< 0.5) were considered as significantly overexpressed and underexpressed, respectively. Dashed line represents the cutoff of significant overexpression. Results were normalized using 16S RNA gene and presented in linear scale.

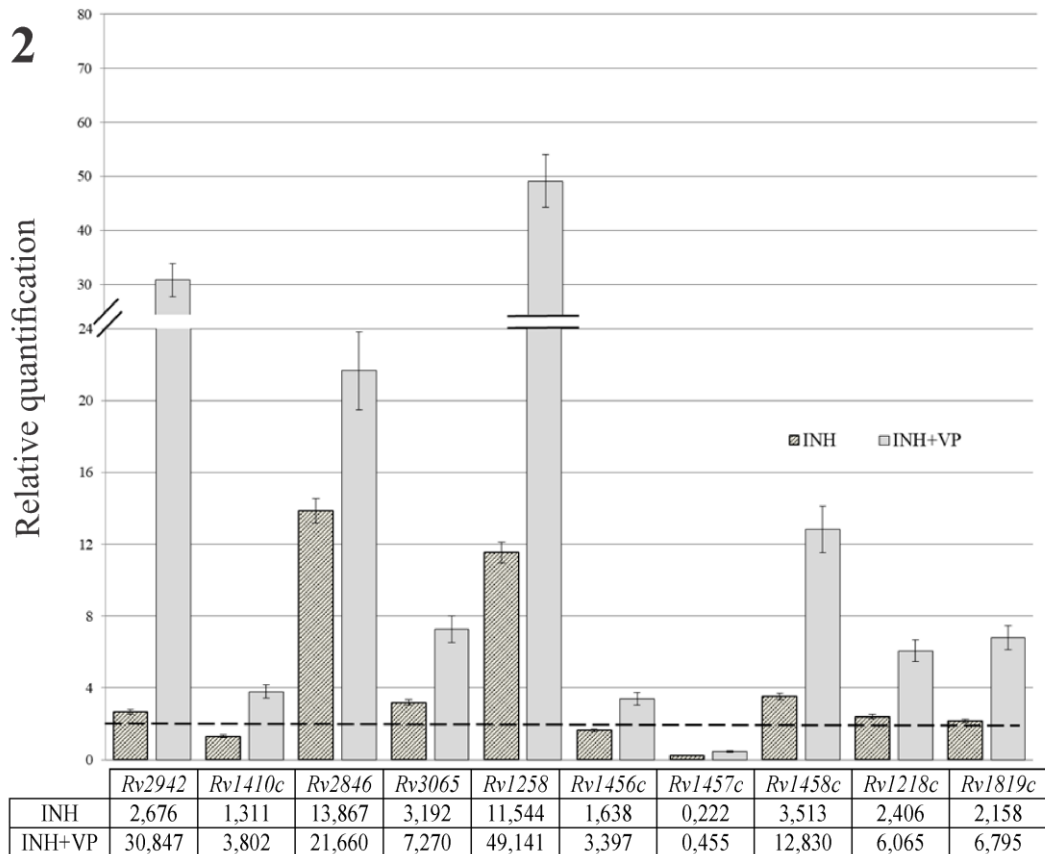


Figure 2. Relative quantification of 10 efflux pump genes in *Mycobacterium tuberculosis* BRF 47, accessed by RT-qPCR after 48 h exposure to 0.5xMIC of isoniazid (INH) and the combination INH+VP. Relative quantification was determined by the means of $2^{-\Delta\Delta CT}$. Relative quantification rates above two (> 2) and lower than 0.5 (< 0.5) were considered as significantly overexpressed and underexpressed, respectively. Dashed line represents the cutoff of significant overexpression. Results were normalized using 16S RNA gene and presented in linear scale.

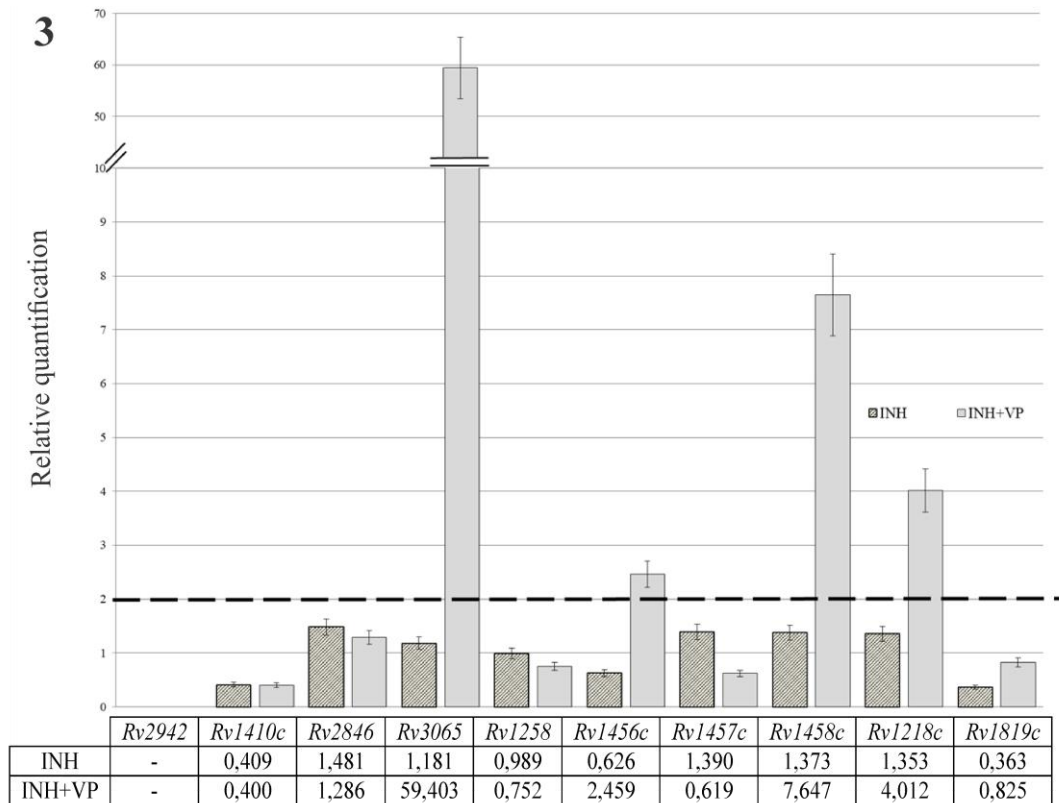


Figure 3. Relative quantification of 10 efflux pump genes in *Mycobacterium tuberculosis* BRF 14, accessed by RT-qPCR after 48 h exposure to 0.5xMIC of isoniazid (INH) and the combination INH+VP. Relative quantification was determined by the means of $2^{-\Delta\Delta CT}$. Relative quantification rates above two (> 2) and lower than 0.5 (< 0.5) were considered as significantly overexpressed and underexpressed, respectively. Dashed line represents the cutoff of significant overexpression. Results were normalized using 16S RNA gene and presented in linear scale. -, relative transcription levels not detected.

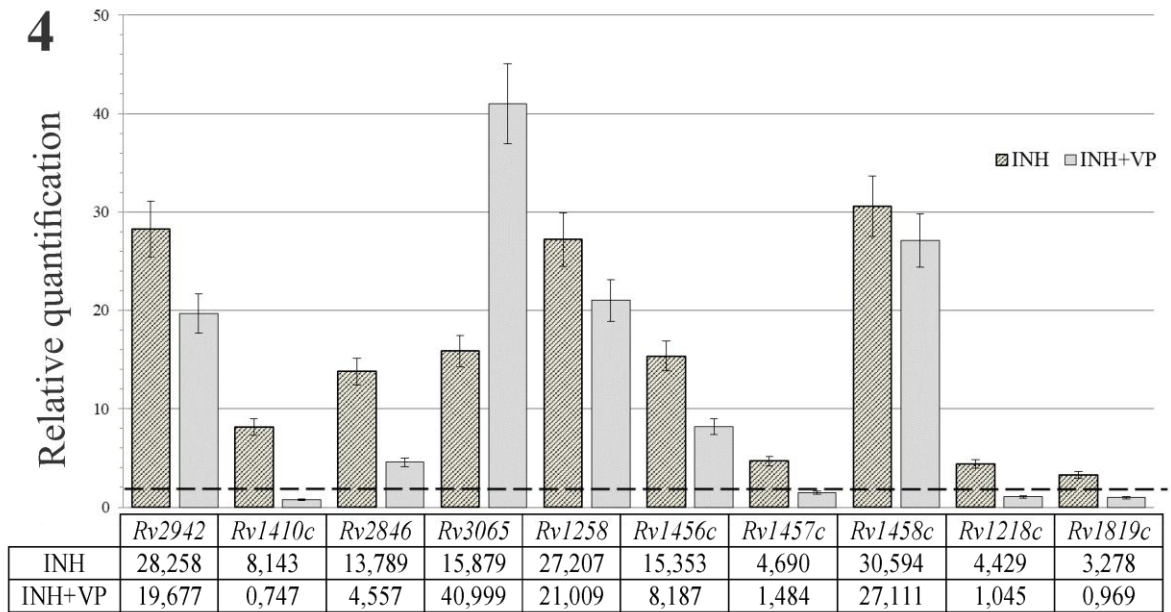


Figure 4. Relative quantification of 10 efflux pump genes in *Mycobacterium tuberculosis* 3614, accessed by RT-qPCR after 48 h exposure to 0.5xMIC of isoniazid (INH) and the combination INH+VP. Relative quantification was determined by the means of $2^{-\Delta\Delta CT}$. Relative quantification rates above two (> 2) and lower than 0.5 (< 0.5) were considered as significantly overexpressed and underexpressed, respectively. Dashed line represents the cutoff of significant overexpression. Results were normalized using 16S RNA gene and presented in linear scale.

CAPÍTULO III

3.1 CONCLUSÕES

Os sistemas de efluxo têm se mostrado como mecanismos importantes no desenvolvimento de resistência aos antimicrobianos em *Mycobacterium tuberculosis*, visto que cada proteína transportadora é capaz de expulsar uma grande variedade de substratos. Com base nos resultados obtidos no presente estudo e que corroboram com publicações disponíveis na literatura, podemos concluir que estas bombas de efluxo têm papel fundamental na resistência à isoniazida, principalmente em isolados sensíveis.

3.2 PERSPECTIVAS

Dando continuidade aos resultados aqui obtidos, nosso grupo de pesquisa buscará identificar novas estratégias que possam superar os mecanismos de resistência à isoniazida mediados por sistemas de efluxo. Dentre essas estratégias, buscaremos comparar a taxa de surgimento de mutantes expostos somente a INH e compara-las à exposição conjunta de INH+VP.

ANEXO I: INSTRUÇÃO PARA AUTORES DO PERIÓDICO TUBERCULOSIS



TUBERCULOSIS

AUTHOR INFORMATION PACK

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ISSN: 1472-9792

6. DESCRIPTION

Tuberculosis is a speciality journal focusing on basic experimental research on tuberculosis, notably on bacteriological, immunological and pathogenesis aspects of the disease. The journal publishes original research and reviews on the host response and immunology of tuberculosis and the molecular biology, genetics and physiology of the organism, however discourages submissions with a metaanalytical focus (for example, articles based on searches of published articles in public electronic databases, especially where there is lack of evidence of the personal involvement of authors in the generation of such material).

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Immunology

Immunogenetics

Pathogenetics

Microbiology

Microbial physiology

Pathogenesis

Pathology

Molecular epidemiology

Diagnostics

Vaccine development Drug resistance The resurgence of interest in tuberculosis has accelerated the pace of relevant research and Tuberculosis has grown with it, as the only journal dedicated to experimental biomedical research in tuberculosis.

7. IMPACT FACTOR

2017: 2.727 © Clarivate Analytics Journal Citation Reports 2018

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Biological Abstracts

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